

Catalogue of alien plants of the Czech Republic

Katalog zavlečených druhů flóry České republiky

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The first author dedicates this paper to the memory of his father Antonín Pyšek

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Pyšek P., Sádlo J. & Mandák B. (2002): Catalogue of alien plants of the Czech Republic. – Preslia, Praha, 74: 97–186.

Alien flora of the Czech Republic is presented. In Appendix 1, 1378 alien taxa (33.4% of the total flora) are listed with information on the taxonomic position, origin, invasive status (casual, naturalized, invasive; a new category post-invasive is introduced), time of immigration (archaeophytes vs. neophytes), habitat type invaded (natural, seminatural, human-made), vegetation invaded (expressed as occurrence in phytosociological alliances), mode of introduction into the country (accidental, deliberate), and date of the first record. Number of phytogeographical as well as biological and ecological attributes were compiled for each species in the database; its structure is presented in Appendix 2 as a suggestion for similar work elsewhere. Czech alien flora consists of 24.1% of taxa which arrived before 1500 (archaeophytes) and 75.9% neophytes. There are 891 casuals, 397 naturalized and 90 invasive species. Of introduced neophytes, 21.9% became naturalized, and 6.6% invasive. Hybrids contribute with 13.3% to the total number of aliens, and the hybridization is more frequent in archaeophytes (18.7%) than in neophytes (11.7%). If the 184 hybrids are excluded from the total number of aliens, there are 270 archaeophytes and 924 neophytes in the Czech flora, i.e. total of 1195 taxa. Accidental arrivals account for 53.4% of all taxa and deliberate introduction for 46.6%; the ratio is reversed for neophytes considered separately (45.5 vs. 54.5%). Majority of aliens (62.8%) are confined to human-made habitats, 11.0% were recorded exclusively in natural or seminatural habitats, and 26.2% occur in both types of habitat. Archaeophytes and neophytes occur in 66 and 83 alliances, respectively, of the phytosociological system. Flora is further analysed with respect to origin, life histories, life forms and strategies. Only 310 species (22.4% of the total number of all alien taxa) are common or locally abundant; others are rare, based on a single locality or no longer present. The following 19 taxa are reported as new for the Czech alien flora: *Agrostis scabra*, *Alhagi pseudalhagi*, *Allium atropurpureum*, *Bromus hordeaceus* subsp. *pseudothomini*, *Carduus tenuiflorus*, *Centaurea xgerstlaueri*, *Centaurea nigra* × *phrygia*, *Cerastium xmaureri*, *Gilia capitata*, *Helianthus strumosus*, *Hieracium pannosum*, *Hordeum leporinum*, *Oenothera coronifera*, *Papaver atlanticum* subsp. *mesatlanticum*, *Parietaria pennsylvanica*, *Polypogon fugax*, *Rodgersia aesculifolia*, *Sedum pallidum* var. *bithynicum*, *Sedum stoloniferum*; these represent results of our own field research as well as of herbaria search, and unpublished data from colleagues. Other 44 taxa are reported as escaping from cultivation for the first time. Twenty two archaeophytes are listed in the Red List of the Czech flora.

Key words: Alien flora, complete list of taxa, immigration status, casual, naturalized, invasive, time of immigration, abundance, mode of introduction, habitat type, hybridization, life history, life form, life strategy, taxonomy, species characteristics, Czech Republic

Introduction

The core of modern research in plant invasions is in ecological studies, fuelled by an effort to predict invasiveness of particular species and vulnerability of various communities to

invasions. In the last decade, the field became fully comparable with other areas of modern ecology and “hard theories“ appear to emerge (Rejmánek 1996, Rejmánek & Richardson 1996, Lonsdale 1999). The focus on ecology reflects the fact that studies on invasions are among biological disciplines with the strongest practical appeal and recently, this aspect has become its focal point. Papers devoted to impact and its quantitative description as well as to critical evaluation of prediction possibilities have been published recently (e.g. Daehler & Carino 2000, Williamson 1996, 1999, 2001, Kolar & Lodge 2001, Pyšek 2001). Debate on risks associated with GMOs as a subset of potentially invasive taxa further increases practical importance of the field (Regal 1986, Kowarik 2002). Nevertheless, despite increasing focus on experimental approaches papers comparing species lists are a popular and useful tool, especially for generating hypotheses which can be then tested by experimental and comparative methods (Weber 1997, Daehler 1998). Importance of taxonomy for studies of alien flora has been repeatedly stressed and awareness of this has been increasing, too (McNeely 2001). In a field like this, where species move dynamically over the globe and one of the frequent and basic situations workers have to face is determination of a species new to their region, and often coming from distant areas, quality background data are needed. Having a reliable database of alien species of a given territory, summarizing historical knowledge of generations of botanists, may therefore prove as a very useful tool.

Some information on alien floras is available for many European countries (see e.g. Weber 1997), although the quality of such data is highly variable. Generally, there is a remarkable difference between data drawn from standard floras and checklists commenting on species immigration status and studies focussing specifically on alien plants. Williamson (2002) has shown for the British flora how careful one must be when trying to make conclusions about the number of native species; numbers of aliens are undoubtedly even more difficult to estimate. Particular sources often give very different figures (Williamson 2002).

In Europe, the best available data on alien flora, in terms of completeness of the species list, is that of the British Isles (Clement & Foster 1994, Ryves et al. 1996). This data set, in association with databases of biological attributes available for British flora (Fitter & Peat 1994) and geographical information, proved to be a powerful tool and yielded interesting results explaining the pattern of alien floras (Crawley et al. 1996). A comprehensive list of alien species with floristic status, degree of naturalization, date and mode of introduction, and chorological, biological and ecological data will be soon available for Germany (Kuehn & Klotz 2002). Moreover, detailed information on the structure and composition of alien flora, its historical dynamics and factors underlying its development have been available for Germany, too (see Kowarik 2002 and references therein). Switzerland is another European country with solid information on its alien flora (Weber 1999), and a project on this topic has started in Italy (Celesti Grapow, pers. com.). For Poland, a detailed information is available for a subset of alien species, i.e. archaeophytes (Zajac 1979). Detailed alien floras have been published for number of large European cities which are recognized as extremely species-rich centres of aliens (Pyšek 1998a), but complete catalogues of alien species for particular countries are still rather rare, unlike in other parts of the world more affected by plant invasions (e.g. Wells 1986 for South Africa).

Geographical conditions in the Czech Republic, history of human colonization and its relevance to plant invasions

The Czech Republic covers an area of 78, 864 square kilometers and has 10.3 million inhabitants, creating a human population density of 131 inhabitants per km². The network of roads (0.71 km per km²) and railways (0.11 km per km²) is rather dense. These features certainly contribute to the richness of alien flora (Pyšek & Prach 2002). Several historical and geographical factors have significantly affected the course of human-induced plant invasions over the past 6 000 years:

1. The Czech Republic is an ecotone between large continental landscape sections: the Alps on the south, Carpathians on the east, Pannonian basin located southeast, region of oceanic climatic on the west, and the north-located region of low habitat diversity resulting from the Quarternary glaciation. There are number of natural and human-created migration routes which provide possibilities for colonization; these are oriented E-W, and SE-NW. Many species reach their northwestern distribution limits near SE political boundaries of the Czech Republic (Slavík 1988).
2. Compared to similar regions of Central and Western Europe, the landscape mosaic is diverse and remarkably heterogeneous in the Czech Republic. Diverse geological, soil and climatic conditions create suitable environments for many different types of plants (Hejný & Slavík 1988), and the majority of Central-European habitat types are present (except for coastal and alpine habitats). The human impact and types of land-use are also rather heterogeneous, both on historical time scale and recently.
3. The dynamics of plant migrations are similar to those in other central- and western-European regions; there has been a continuous stream of plant invasions since the Neolithic agricultural colonization which started in about 5300 B. C. and represented the first milestone in the history of alien plant invasions. The main landscape changes that accompanied particular plant invasion waves followed during the Aeneolite (3800 B. C.), Bronze Age (2200 B. C.) (Opravil 1980), Medieval (13th to 15th century), and recent time (since the 19th century). As early as the Aeneolite, there was a rather high proportion of deforested landscape in lowlands (Ložek 1999), and divergent development dividing the landscape into warm cultural lowlands and cold forested highlands started during this period. The landscape was gradually colonized between the Neolithic period (Central Bohemia, South Moravia) and the Medieval (cold highlands), but the highest mountains were only colonized between the 17th to 19th century. Until the Late Medieval, there were still large portions of closed forests and these acted as barriers to migrations.
4. There was little and very local exchange of goods until the beginning of the Late Medieval. Not until this period were there developed towns and large scale migration of humans and goods (Le Gof 1982), although trading routes specialized on salt, gold and amber were used in prehistoric times. The industrial revolution started in the region in 1850s and in the first half of the 20th century, the Czech Republic was one of the most highly developed industrial countries in Europe. In 1945–1989 the country was isolated from the Western Europe because of the socialistic political regime which brought about economic orientation towards the East and specific features of land-use (including so-called “collectivization”, involving the concentration of agricultural production into large production units, and the evacuation of border areas and their subsequent colonization). Many plant species of Asian and southeast-European origin entered the

central part of the continent via one of the largest European railway stations in Čierna nad Tisou in the Slovak part of the former Czechoslovakia (Jehlík & Hejný 1974, Jehlík 1998). Besides railways and roads, river traffic on the Elbe river, the Danube river and their tributaries significantly contributed to the richness of present alien flora (Jehlík 1998).

5. Despite isolation and political differences, the country went through the same process as other European regions between the 1940s and 1990s. Traditional economical and land-use models based on Neolithic scheme have ceased (Hobsbawm 1991). A new landscape type came to a large-scale existence since the 1990s. This landscape model can be characterized by the following features in the Czech Republic: (i) humans are less present in the open landscape, (ii) direct human intervention into the landscape are less frequent but more powerful, (iii) environmental stress associated with traditional agricultural management has been decreasing, (iv) the role of disturbances associated with industrial activities and urbanization increases, and (v) so do the migration possibilities (Cílek 1999, Sádlo & Storch 2000).

History of floristic research and its relevance to studies on alien species

The remarkable tradition of studies on floristics in the Czech Republic provides a solid background for compiling a list of alien species of reasonable historical relevance. Floristic research in the territory of the Czech Republic dates back to the second half of the 18th century. The first attempt at producing the flora of the whole Czechia was carried out by Schmidt (1793–1794); unfortunately some data given by him are doubtful (Skalický et al. 1988). In the first half of the 19th century, several floral works mention introduced plants and can be therefore used to infer information about plant invasions at that time (Pohl 1809, Presl & Presl 1819, Opiz 1823, 1852). For the following period, the wealth of information on alien plants can be found in the remarkable work of Čelakovský (1867–1881, 1882–1894) who recognized the alien status and origin of some plants present in the Czech flora and commented in considerable detail on their distribution. In the early 20th century botanists started to recognize human-made and disturbed habitats as a source of important additions to native floras (e.g. Laus 1908, Domin 1917–1919). Alien plants started to be systematically recorded, thanks to the founding of a specialized research section at the Institute of Botany, Průhonice, in the 1960s. Research triggered then has focussed on specific habitats (ports, railways, oilseed or wool processing factories, grain silos, mills, rubbish tips, arable land, etc.), taxonomically relevant groups and on the distribution of alien plants, as well as on their ecology and impact (see Hejný et al. 1973, Pyšek 1995a, Jehlík 1986, 1998 for references). The tradition of recording plant distribution made it possible to produce some valuable data sets providing detailed information on distribution of selected alien species (e.g. Jehlík 1998); unfortunately, they were not always adequately analyzed. In addition to this primary research, valuable floristic information about species immigration status can be derived by a careful analysis of old floral works, some of which date to 1600 (Hendrych 2001).

Surprisingly, despite this background, no comprehensive catalogue of alien plants occurring at the territory of the country has been available until now. The need for such a list has become more urgent as the research in plant invasions has been intensifying (Pyšek &

Prach 2002), and comparative studies on alien floras started to receive considerable attention (Weber 1997). Until now, data for the territory of the Czech Republic used to be taken mostly from the work of Dostál (1948–1950, 1954, 1958) on which the importance of the reliability of the data can be demonstrated. The flora of Dostál (1948–1950) which became a modern standard for Czech botanists of the second half of the 20th century gives an indication of alien origin of 599 taxa. The vast majority of the taxa are neophytes, as Dostál did not consider archaeophytes as aliens. The 599 taxa listed by Dostál represent approximately 20% of the the flora (the total number of species was 3120; Dostál 1954). Of the neophytes reported in this flora, 109 were excluded from the present list for various reasons: they were re-classified as native (e.g. *Acer tataricum*, *Plantago indica*, *Solanum alatum*, *Potentilla norvegica*), reported erroneously as they probably never grew at the territory of the country (e.g. *Cuscuta australis*, *Capsella rubella*, *Lupinus perennis*, *Erysimum perofskianum*), or, most frequently, it is uncertain whether or not they ever escape from cultivation (e.g. *Linaria purpurea*, *Mesembryanthemum crystallinum*, *Helianthus debilis*, *Ptelea trifoliata*). This phenomenon certainly deserves more attention and careful analysis since the data from Dostál's flora were taken as a basis for comparison with other regions (Pyšek 1989) because there was no other source available. Given that there are 924 neophytes (excluding hybrids) listed in the present paper, which number reflects better the real situation, and only as few as 490 are common with the Dostál's flora, the value of Jaccard coefficient of similarity between the two data sets is as low as 0.47. Even if we take into account the increase in the number of aliens in the Czech flora during the last four decades (Pyšek et al. 2002) reflecting accelerated translocation of species over the Earth surface in the second half of the 20th century, the difference between the recent and former data sets is too big to be attributed only to such an explanation and indicates lower reliability of the earlier data. Moreover, the Dostál's data set was for the territory of former Czechoslovakia which includes Slovakia. The species number in this larger region should be therefore higher. However, this fact influences the comparison of past and present alien floras in two contrasting ways: species introduced only to Slovakia are missing from the present data referring only to the Czech Republic but there are taxa whose native distribution ends in Slovakia and their occurrence in the Czech Republic is therefore secondary (e.g. *Trifolium angulatum*, *Cotinus coggygria*, *Orobanche gracilis*, *Scutellaria altissima*, *Beckmannia eruciformis*, *Pulsatilla slavica*, *Silene viridiflora*). Such taxa did not appear among aliens on Dostál's list but are included on the present one.

Different reliability of data in earlier floras is not the only reason for need to update lists of aliens of a given territory. Plant invasions are, by their very nature, extremely dynamic, and there is a constant influx of new species. Records of these newcomers are usually scattered in the local literature, much of which is not covered in international abstracting journals; such records are thus generally unavailable to international readers. An attempt to compile a list for any territory brings data to light which would otherwise be lost. Even very good and detailed floras do not pay the same attention to alien species; the quality of information and the attention paid namely to casual aliens varies. The reasons are that any such work must necessarily rely upon many contributors who feel differently about nonindigenous members of floras, and also, publication of such works usually spans over a considerable time period. The Flora of the Czech Republic (Hejný & Slavík 1988–1992, Slavík 1995–2000) is no exception in this respect. However, a decade in the contemporary world is a lifetime in plant invasions!

A good species list is clearly the prime task of such a work; another important aspect is the kind of information associated with particular taxa on the list. Databases specifically focussed on alien species have an advantage, compared to standard floras, that such information can be compiled in a considerable detail (see Appendix 2).

Data sources

The work “Flora of the Czech Republic”, of which 6 out of 8 planned volumes have been published, served as a general information source for that part of the flora which has been covered so far (Hejný & Slavík 1988–1992, Slavík 1995–2000). The newly prepared Key to the flora of the Czech Republic (Kubát et al. 2002) was also checked for the coverage of alien species. Earlier modern floral works from the second half of the 20th century were also critically evaluated (notably Dostál 1948–1950, 1954, 1958, 1989). The list of Opravil (1980) served as a basic source on residence time for species introduced before 1500. General information on biological and ecological attributes was further completed by using synthetic floral works from other regions (Tutin et al. 1964–1980, Frank & Klotz 1990, Stace 1991), and specialized compendia on chromosome numbers and ploidy levels (Májovský et al. 1987), dormancy and germination behaviour (Baskin & Baskin 1999), seed bank formation (Thompson et al. 1997), invasive behaviour elsewhere in the world (Clement & Foster 1994, Ryves et al. 1996, Frank & Klotz 1990, Kartesz & Meacham 1999), dispersal mode (Lhotská & Chrtková 1978, Lhotská et al. 1987), endangerment and conservation status (Holub & Procházka 2000), history of introduction (Hejný et al. 1973, Jehlík 1998), and planting and cultivation aspects (Walters et al. 1984–1989, Cullen et al. 1995–2000, Brickell 1989). For other information not given in these sources, we searched the primary literature (see References). We also used herbaria (mostly National Museum Prague – PR, Charles University – PRC, and Institute of Botany Průhonice – PRA), unpublished information from colleagues, and results of our own field research in 1999–2001.

Terminology, approach, and classification measures

Former floras and works related to plants non-native to the territory of the Czech Republic (see References) were considered when evaluating species status. However, each particular taxon was carefully re-assessed to confirm its native/alien status (Webb 1985, Pyšek 1995b, Richardson et al. 2000), its invasive status (Richardson et al. 2000) and residence time. For this evaluation, knowledge of species ecology and habitats occupied was used, in association with historical dynamics and role it plays in the landscape. The knowledge of landscape history since Neolithic times was also employed (Ložek 1999).

All alien species ever recorded at the territory of the country at least once in the wild were included¹. Another important condition for inclusion was that a species is alien to the

¹ Only those species which occurred in the wild were considered; i.e. we did not take into account those kept exclusively in cultivation but considered escapees. In some cases, it can be rather tricky to decide; this concerns especially plants thrown away from gardens. We adopted the following criterion: a plant was included on the list if it reproduced on its own at least once outside the space where it was sown or planted (i.e. outside the flower bed or garden). In plants reproducing by seed, germination outside such space was considered as escape from cultivation. A plant reproducing clonally was considered as an escape from cultivation only if it survived winter and persisted in a given site until the following growing period.



Fig. 1a. – Herbarium specimen of *Angelica archangelica* subsp. *archangelica* from 1893 (leg. J. Košťál PR). Presence of this neophyte at the territory of the Czech Republic has been proved as early as in 1517 (Slavík 1997).



Fig. 1b. – *Rodgersia aesculifolia*, the most recent addition to the alien flora (2001). There is a span of more than four centuries and minimum 1044 species that have immigrated during the time between the introduction of *Angelica archangelica* subsp. *archangelica* and that of this species.

whole territory of the country. If a species has or had a single locality at the territory of the country where it is considered native, it was not included onto the list. Similarly, no consideration of so-called “apophytes” (native species occurring on secondary habitats, see e.g. Holub & Jirásek 1967) was given; once a species has native habitats in the Czech Republic, its expansion into other communities did not qualify it for inclusion on the list. Similarly, species which occurred at the territory as native in the past were excluded². A strictly geographical approach to plant invasions, as opposed to one based on human values and relying on the realization of some form of “impact” (Davis & Thompson 2001), was therefore adopted (Rejmánek 1995, Richardson et al. 2000, Daehler 2001, Rejmánek et al. 2002). Doubtful records, which are sometimes listed without evidence from one flora to another, were excluded; we adopted a conservative approach. On the other hand, once a declaratively complete work on alien flora of any territory has been published, it is tempting for future researchers to start with that and pay less attention to scattered information sources from earlier times. This brings about a danger that most of what is not covered might be overlooked in the future. For that reason we included some records that are

² *Sorbus intermedia* (Ehrh.) Pers. is an example of such an approach; this taxon is commonly planted and escapes from cultivation but its native occurrence in the Krkonoše Mts from the turn of the 19th and 20th century cannot be excluded (Kovanda in Hejný & Slavík 1992). For that reason, this species is not included in the list.

not steadfastly proved because the respective herbarium specimen is not available, but other circumstances, such as the personality of the author, make it probable that they were correctly determined (e.g. *Hyoscyamus albus*, see Slavík 2000)³.

Crosses between natives and aliens are always considered as aliens, even if they have arisen in the Czech Republic. They are non-indigenous species in the sense of not having been at the territory before the onset of Neolithic agriculture (see also Williamson 2002). If a native species was taken into cultivation, its cultivars were produced and subsequently escape into the wild (e.g. *Achillea ptarmica*), such species was not included on the list because at the taxonomic levels considered in the present paper, i.e. species and subspecies, it is native in the territory (the only exception to this rule is rather common and well recognizable *Phalaris arundinacea* var. *picta*).

For taxa which are covered by it, the nomenclature follows the determination key to the Czech flora (Kubát et al. 2002). To avoid confusion, authorities are consistently given in Appendix 1. Nomenclature of higher taxa follows the Cronquist system as presented in Mabberley (1997). We only distinguished taxa up to the intraspecific level of subspecies; the only exceptions are *Reynoutria japonica* (var. *japonica* vs. var. *compacta*), *Physalis alkekengi* (var. *alkekengi* vs. var. *franchetii*), *Datura stramonium* (var. *stramonium* vs. var. *tatula*), and *Kochia scoparia* subsp. *scoparia* f. *trichophylla*.

The evaluation of the invasive status of a taxon, i.e. its stage in the “naturalization-invasion process”, should be a key point in any study dealing with alien floras. We followed the scheme proposed by Richardson et al. (2000), which is based on overcoming different kind of barriers an invading plant must face. The following categories were distinguished: casuals, naturalized, and invasive aliens⁴. On top of the standard classification introduced by Richardson et al. (2000), which describes the highest degree of invasiveness reached by a given species, we included another category which is supposed to reflect the historical dynamics, i.e. changes in a species’ invasive status. There is no reason to doubt that archaeophytes went, after their introduction, through processes similar to those we witness today with neophytes, and that their recent distribution is in many cases only a remnant of

³ In other cases, however, the support for inclusion was considered too weak. For example, *Mibora minima* (L.) Desv. has been reported as an adventive species of the Czech flora (Dostál 1989, based on Chrték 1965). The record was based on a single herbarium specimen from the Zahlbrückner collection located in PRC (s.a., s.d.) who was not, however, the collector of this plant. The sheet bears location “E Bohemia: Pohl” in Zahlbrückner’s handwriting (J. Hadinec, pers. com.). Obviously, it is too risky to include the species on the basis of second-hand information without precise location. In addition, the species is native to the neighbouring Germany so even if it was collected in Bohemia, it might have represented native occurrence. Its unclear status is reflected by it being listed among “uncertain cases of extinct and missing” taxa of the Red list of the Czech flora (Holub & Procházka 2000).

⁴ **Alien plants:** Plant taxa in a given area whose presence there is due to intentional or accidental introduction as a result of human activity.

Casual alien plants: Alien plants that may flourish and even reproduce occasionally in an area, but which do not form self-replacing populations, and which rely on repeated introductions for their persistence.

Naturalized plants: Alien plants that reproduce consistently (cf. casual alien plants) and sustain populations over more than one life cycle without direct intervention by humans (or in spite of human intervention); they often recruit offspring freely, usually close to adult plants, and do not necessarily invade natural, semi-natural or human-made ecosystems.

Invasive plants: Naturalized plants that produce reproductive offspring, often in very large numbers, at considerable distances from parent plants (approximate scales: > 100 m / < 50 years for taxa spreading by seeds and other propagules; > 6 m / 3 yrs for taxa spreading by roots, rhizomes, stolons, or creeping stems), and thus have the potential to spread over a considerable area.

After Richardson et al. (2000).

the past abundance and distribution. We made an attempt to take this into account by including a category “post-invasive”. Criteria for including archaeophytes as post-invasive were that the species now has a stable (not increasing) or even decreasing population and does not invade new, modern types of habitats, but its population dynamics and type of occurrence suggests that it might have belonged to the vegetation dominants in the past. Examples include: *Atriplex rosea*, *Spergula arvensis*, *Agrostemma githago*, *Chenopodium polyspermum*, or *Gagea villosa* (see Appendix 1). We also labelled some neophytes as post-invasive; this concerns species for which it is rather striking that, after having reached a distribution peak in the past, they either retreated or their distribution is more or less stable – i.e., they no longer spread (e.g. *Elodea canadensis*, *Mimulus guttatus*, *Imperatoria ostruthium*). It should, however, be borne in mind that the post-invasive category is, more than any other, based on our personal opinion; it is therefore more speculative than the others. Nonetheless, we consider this term to be a useful and informative addition to the traditionally applied criteria of a species’ position within the dynamics of invasion process. When a species occurs in the locality for a long time, seemingly naturalized but it is there as a remnant of past planting (e.g. *Rosa rugosa*, *Filipendula kamtschatica*, *Cotoneaster horizontalis*, *Potentilla fruticosa*, *Lonicera tatarica*, etc.) it is termed “cultivation relic” and indicated in Appendix 1.

With respect to the residence time, i.e. the time since the arrival of a species in the territory, we distinguish archaeophytes (introduced before the discovery of America, approx. 1500 A. D.) and neophytes (introduced after that date). Discussion is required here on how these terms were used by previous authors. Some terms were introduced by original sources in a slightly different sense but their meaning has shifted since then. This is most remarkable in the usage of the term “neophyte”. Strictly speaking, deliberately introduced species are not neophytes in the sense of e.g. Holub & Jirásek (1967) and should be termed “xenophytes”. For simplicity and compatibility with recent usage of the term, we use it without any relation to whether the given species arrived accidentally or was brought in by humans. It only reflects the residence time (species introduced after the year 1500) regardless of the mean of introduction.

Terminological frameworks for classifying alien species by reflecting their relationship to humans as vectors of introduction, dispersal, and habitat transformation (e.g. Thellung 1905, Kreh 1957, Holub & Jirásek 1967, Schroeder 1969) are traditionally used in Central-European countries. We used the system of Richardson et al. (2000) because: (a) it is simpler than traditional classification schemes; still it is compatible with them (Table 1; Pyšek 1995b). It differs in that it answers the basic classification questions⁵ by combining independent criteria, rather than creating specific terms for each possible situation. (b) It is being partly recognized by international scientific bodies such as Global Invasive Species Programme (McNeely 2001), hence it is a candidate for becoming generally acceptable basis for terminology in plant invasions. (c) The traditional Central-European classification of alien plants has never received notable recognition in English speaking scientific community; the terms “archaeophytes” and “neophytes” represent an exception in this respect (Williamson 2002).

However, there are several categories, between which the distinction is sometimes blurred and depends, more than elsewhere, on the level of knowledge of species ecology

⁵ When did the species arrive? Why did it arrive, i.e. what means made it possible? Where does it invade, i.e. in which habitats? How far did it get in the invasion process?

Table 1. – Comparison of terminology associated with alien plants which has been traditionally used in Central-European classification schemes with the one adopted for the present paper. The former is based on the classification of Holub & Jirásek (1967), the latter on Richardson et al. (2000). Criteria used by Holub & Jirásek (1967) for classification of particular categories are indicated: T = time of immigration, M = means of introduction, H = type of encountered habitat.

Term in Holub & Jirásek (1967)	Criteria	Meaning	Corresponding term in the present paper and its meaning
Anthropophytes		introduced by humans regardless of time and means	alien
I. Hemerophytes	M	introduced deliberately	
1. Ergasiophytes	MH	kept only in cultivation	not included on the list
2. Ergasiophytophytes	MH	kept in cultivation and occasionally escaping	deliberately introduced aliens (mostly casual)
3. Ergasiolipophytes	MH	formerly planted, currently occurring in the territory without need of human intervention	deliberately introduced aliens (naturalized or invasive)
II. Xenophytes	M	accidentally (unintentionally) introduced	any accidentally introduced alien
1. Archaeophytes	MT	accidentally introduced before ca. 1500 ⁶	archaeophytes (introduced before ca. 1500, both deliberately or accidentally)
2. Neophytes	MT	accidentally introduced after ca. 1500	neophytes (introduced after ca. 1500, both deliberately or accidentally)
(a) Ephemerophytes	MTH	occurring temporarily in human-made habitats	neophytes in human-made habitats (casual)
(b) Epekiophytes	MTH	established in human-made habitats	neophytes in human-made habitats (naturalized or invasive)
(c) Neoindigenophytes ⁷	MTH	established in the region, occurring in human-made habitats and penetrating to natural habitats, too	neophytes in natural and/or seminatural habitats (naturalized or invasive)

and perception of landscape history. This includes a decision whether a species is (i) native or archaeophyte, (ii) archaeophyte or neophyte, as long as residence time is concerned, and (iii) casual or naturalized, and (iv) naturalized or invasive when classifying the invasive status. Some special cases have been considered in the present paper too. These include, for example, *Oxalis debilis* and *O. latifolia*, species which often spread in greenhouses of garden centres where they survive without being further dispersed by humans (Holub & Holubičková 1980, Jehlík 1995). However, such species were considered as casuals because of their dependence on glasshouse environment; the outdoor climate of Central Europe does not permit their persistence.

⁶ Approximate date corresponding to the discovery of America (1492).

⁷ Some authors use the term “agriophytes” (Schroeder 1969, Lohmeyer & Sukopp 1002) for this category which is sometimes further divided into „hologriophytes“ (in natural vegetation) and „hemagriophytes“ (in seminatural vegetation) (see e.g. Kornas 1990).

Type of abundance in the landscape was estimated for each species on the list using the following criteria: single locality, rare, scattered, locally abundant, and common at the whole territory. A special category termed “extinct” relates to the situation when no records have been known for a long period, and where it is highly improbable that the species would appear again. In addition, estimates were made of the number of localities using the scale of Clement & Foster (1994): 1–4; 5–14; 15–49; 50–499; and at least 500 localities. Above 15, the number of localities is an estimate.

The first record of the species in the territory was, for obvious reasons, only determined for neophytes. It should be noted, that this date tells us only that the species has been present in the territory since at least the given year. In fact, it could have been, and in many cases undoubtedly was, present for a longer time. This category crucially depends on earliest floras available, and on their quality and completeness. Fortunately, these are regularly spread over the 19th century (Pohl 1809, Presl & Presl 1819, Opiz 1823, 1852, Čelakovský 1867–1894, Polívka 1900–1903) and provide us with solid information about the gradual enrichment of flora by alien species.

For each species, types of habitat in which it is recorded were distinguished: (i) natural (forested landscape and naturally treeless habitats), (ii) seminatural (cultural landscape excluding arable land, communication and human settlements), and (iii) human-made habitats (Chytrý et al. 2001). The type of invaded landscape was also evaluated, distinguishing (i) traditional agricultural landscape, and (ii) industrial urban landscape (Hobsbawm 1991). Particular habitats in which the species is found were classified according to the Council Directive 92/43/EEC on the conservation of natural habitats and of wild fauna and flora (1992).

The type of invaded vegetation was assessed by using alliances of the Zürich-Montpellier phytosociological system (Chytrý et al. 2001). This classification level most reasonably reflects the vegetation diversity of Central-European landscape (Ellenberg 1988).

The database

Various characteristics have been compiled for each species, where such information was available. These can be divided into several topics: (i) species identity and taxonomic position, (ii) invasiveness, i.e. occurrence and behaviour of the species in the Czech Republic. These two topics were treated in detail in the previous section and represent the original information presented in this paper. Other two spheres concern (iii) occurrence and behaviour in primary area, and (iv) biological and ecological attributes collated for each species (see Appendix 2 for details).

How many species, how large a proportion?

The Czech alien flora contains 1378 taxa belonging to 542 genera and 99 families (Appendix 1). Of these, there are 141 taxa at subspecific and 12 at varietal levels; 54 subspecies and 5 varieties are the only representatives of their species. Of the total number of taxa, 184 are hybrids.

This figure can be used to estimate the contribution of aliens to the total floristic richness of the territory of the Czech Republic. However, such estimates even with solid data at

hand, can be rather tricky and depend on several factors. As discussed by Williamson (2002), unfortunately not only numbers of aliens are uncertain but those of native species too, and the figure depends on whether microspecies and hybrids are included in calculations. Williamson (2002) argues that while hybrids are perfectly satisfactory taxa, and there are around 400 of them in British Isles, accounts of the British flora usually omit them. Stace (1997) estimates that there are around 900 native micro-species in British Isles, concentrated namely in genera *Rubus*, *Hieracium* or *Taraxacum*. Including hybrids and/or microspecies in the total of native species makes a huge difference to comparison with other regions; in the case of the relatively species-poor British flora, the total number of native species is about twice as high. Moreover, number of species reported in these critical groups strongly reflects the level of taxonomical knowledge in the country and/or simply the presence or absence of a specialist in a given group; this brings about even more bias to comparison between countries (Daehler & Carino 2001).

The number of native taxa at species and subspecies level listed in the most recent account of the Czech flora (Kubát et al. 2002) is 2754⁸. Microspecies were included in this count; there are good specialists for most critical groups in the Czech Republic and there has been sound research on apomictic species (e.g. Marhold et al. 1999). Kubát et al. (2002) list 114 species of *Hieracium*, 112 native species of *Rubus* and 72 species of *Taraxacum*⁹, to name the three most critical genera mentioned by Williamson (2002). Even if we bear in mind that not all of them are critical, as there are “ordinary” species in those genera as well, we are left with a total exceeding 300 species. The corresponding figures for native representatives of these genera in British flora are approximately 400, 250 and 115, respectively, i.e. giving the total of 765. The contribution of microspecies to the total richness of Czech flora seems to be therefore lower than in British Isles – this is further pronounced by the fact that Czech native flora is richer in “macrospecies” than that of British Isles. For this reason, we consider it justified to include microspecies in the calculation.

Accepting the totals of 1378 aliens and 2754 native species means that aliens contribute 33.4% to the total number of taxa reported for the Czech Republic. Kubát et al. (2002) list 498 crosses of native species so the number of native taxa excluding hybrids is 2256. The corresponding figure for the alien flora, excluding hybrids, is 1194. If the hybrids are not taken into account, contribution of aliens to the total number of taxa increases to 34.6%. This minor difference reflects the fact that hybridization rate is lower in aliens than in native species, possibly due to shorter common occurrence of potential parental species in the territory, their often limited distribution and smaller population sizes, and resulting lower chance to meet; given the diversity of geographical origins among aliens, other barriers to hybridization may play role, too (Briggs & Walters 1997). The native flora is also better known because of tradition of floristic research and historical focus, and crosses of alien species might be therefore also under-recorded compared to those of native flora. A sound answer to the question how large a proportion of Czech flora is formed by aliens, seems to be between 33 and 35%, depending on how the species numbers are derived.

⁸ The counts are based on the final stage of manuscript kindly provided by the editors. They may therefore slightly differ from data contained in the printed version.

⁹ This count does not include sect. *Ruderalia*, of which there are another 105 species (J. Kirschner, pers. com.); total number of microspecies in the flora of the Czech Republic will be therefore somewhat higher.

Composition and structure of Czech alien flora

Of the total number of 1378 taxa in the Czech alien flora, 24.1% arrived before 1500, while 75.9% are neophytes (Table 2). As to the invasive status, 64.7% are casuals, 28.8% were classified as naturalized, and 6.6% as invasive (Fig. 2). Four neophytes and 188 archaeophytes were classified as post-invasive (Appendix 1). Of 891 casual taxa, 91.7% are neophytes and 8.3% archaeophytes; similarly, 76.7% of the total number of 90 invasive taxa are neophytes and 23.3% archaeophytes. The group of 397 naturalized taxa of the Czech alien flora consists of 59.7% of archaeophytes and 40.3% of neophytes (Table 2). The reverse ratio of casual and naturalized taxa in both residence-time groups (Fig. 2) reflects the fact that archaeophytes which would not have become naturalized could hardly be recorded in our times; the 74 casuals in this group represent long cultivated species escaping occasionally from cultivation.

These figures make it possible to calculate ratios of how large a proportion of introduced plants is able to naturalize or invade (Fig. 3). It only makes sense to express this for neophytes because for archaeophytes, the information on the initial stage, i.e. that of casuals, is missing. Of introduced taxa, 21.9% are considered naturalized, while 817 are casuals, and 231 of them are considered extinct. Finally, 6.6% of introduced neophytes are invasive, a figure which corresponds well to theoretical rules and predictions in invasion biology (Williamson 1996).

The vast majority of archaeophytes came from the Mediterranean area, whereas neophytes have their origin in all continents, with other parts of Europe (39.8%), Asia (27.6%), and North America (15.1%) contributing most taxa (Fig. 4).

The taxonomic structure of the alien flora involves families whose representatives commonly invade in temperate climates (Pyšek 1998b), with *Compositae*, *Gramineae*, and *Brassicaceae* most represented (Table 3). Some differences between archaeophytes and neophytes are obvious: *Chenopodiaceae*, *Apiaceae*, *Scrophulariaceae* and *Caryophyllaceae* tend to be better represented among the former, whereas *Fabaceae*, *Solanaceae*, *Polygonaceae*, *Onagraceae* and *Amaranthaceae* are typical “neophytic families”. Compared to the native flora, *Gramineae*, *Brassicaceae*, *Chenopodiaceae*, and *Solanaceae* are over-represented among the aliens, whereas *Rosaceae*, *Cyperaceae*, *Salicaceae*, and *Orchidaceae* are those with remarkable contribution of native taxa (Fig. 5). Some large families contain almost exclusively native (e.g. *Orchidaceae*, 97 native species – Kubát et al. 2002, and only one alien – *Cypripedium reginae*) or exclusively alien (e.g. *Amaranthaceae*, 25 species) representatives. There are 39 families and 162 gen-

Table 2. – Composition of the Czech alien flora. Number of taxa in particular categories of immigration time and invasive status (see text for definitions). Hybrids are included (for their numbers see Table 4). Distribution of archaeophytes and neophytes with respect to invasive status are significantly different (G-test on contingency tables, $G = 379.04$, $df = 2$, $P < 0.001$).

	Casual	Naturalized	Invasive	Total
Archaeophytes	74	237	21	332
Neophytes	817	160	69	1046
Aliens total	891	397	90	1378

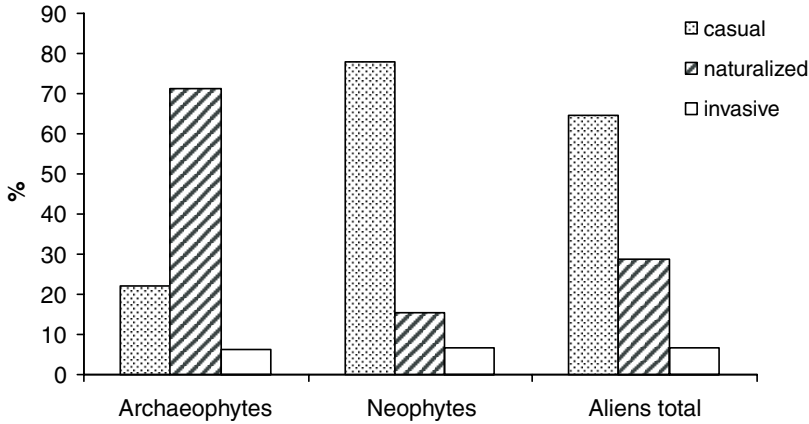


Fig. 2. – Invasive status in groups of alien flora classified according to the immigration time.

era containing archaeophytes and 98 families and 477 genera (including one nothogenus) with neophytes in the Czech alien flora. The total number of families with native species is 138. The genera with the highest number of alien taxa are: *Chenopodium* (27), *Amaranthus* (24), *Oenothera* (23), *Bromus* (21), and *Vicia* (18).

Annuals contribute to the total number of archaeophytes with 57.8%, significantly more than to that of neophytes (39.4%). Perennials (38.2%) and woody plants (14.1%) are more frequent among neophytes than among archaeophytes (Fig. 6). In total, the Czech alien flora comprises 44.0% annuals, 9.3% biennials, 34.4% perennials, 7.7% shrubs and 4.5% trees.

The distribution of Raunkiaer’s life forms (see e.g. Ellenberg 1988) in aliens is different from that in the native flora, with all groups except for therophytes and phanerophytes over-represented in the latter (Fig. 7).

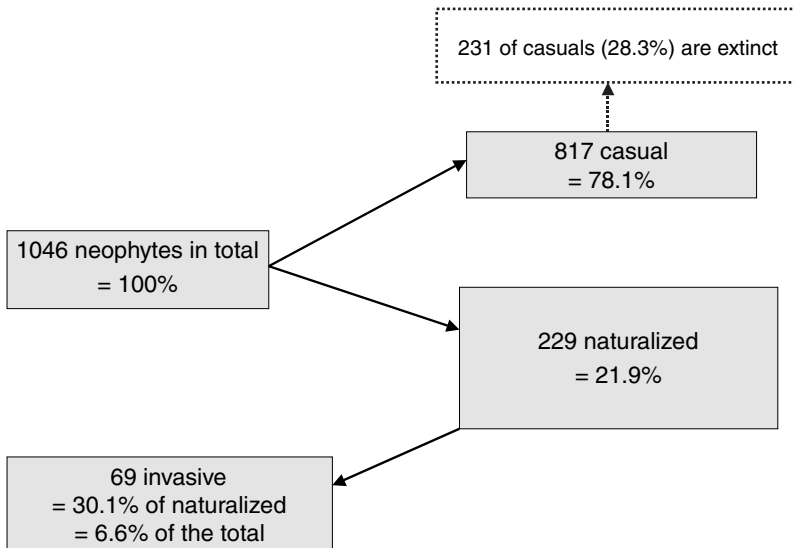


Fig. 3. – Transition rates between particular categories of invasive status in Czech aliens (see text for explanation).

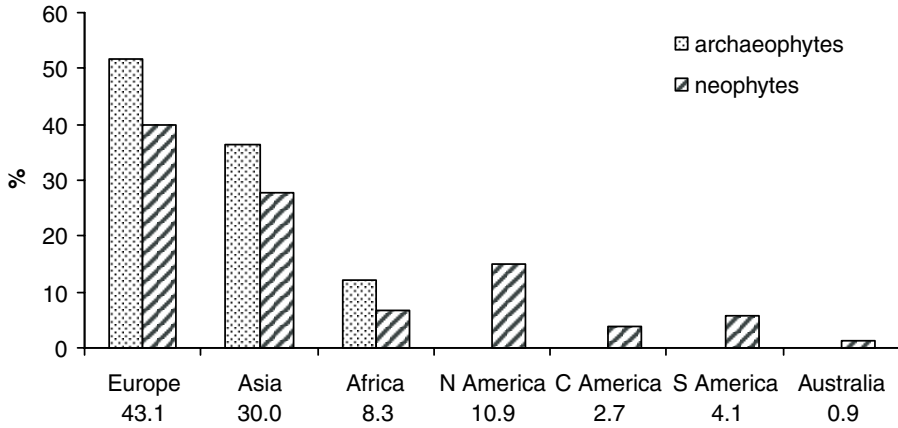


Fig. 4. – Structure of the Czech alien flora with respect to origin. If a species distribution area covers more than one continent, it is considered as a representative of each of them. Percentage contribution of areas of origin to the total number of aliens follows the name of the continent.

Table 3. – The most represented families in the Czech alien flora. Only those with percentage contribution to the total number of alien taxa ($n = 1378$) exceeding 1% are displayed. Archaeophytes and neophytes differed significantly with respect to the representation of plant families (G-test on contingency tables, $G = 133.17$, $df = 35$, $P < 0.001$, only families with at least 5 species were considered in calculation).

	Number of species			%		
	Archaeo-phytes	Neo-phytes	Aliens	Archaeo-phytes	Neo-phytes	Aliens
<i>Compositae</i>	52	135	187	15.7	12.9	13.6
<i>Gramineae</i>	38	113	151	11.4	10.9	11.0
<i>Brassicaceae</i>	29	72	101	8.7	6.9	7.3
<i>Fabaceae</i>	13	76	89	3.9	7.3	6.5
<i>Rosaceae</i>	16	62	78	4.8	5.9	5.7
<i>Lamiaceae</i>	18	46	64	5.4	4.4	4.6
<i>Chenopodiaceae</i>	22	33	55	6.6	3.2	4.0
<i>Apiaceae</i>	17	24	41	5.1	2.3	3.0
<i>Scrophulariaceae</i>	15	24	39	4.5	2.3	2.8
<i>Onagraceae</i>	0	38	38	0.0	3.6	2.8
<i>Caryophyllaceae</i>	17	20	37	5.1	1.9	2.7
<i>Solanaceae</i>	3	33	36	0.9	3.2	2.6
<i>Polygonaceae</i>	2	27	29	0.6	2.6	2.1
<i>Boraginaceae</i>	11	14	25	3.3	1.3	1.8
<i>Amaranthaceae</i>	2	23	25	0.6	2.2	1.8
<i>Ranunculaceae</i>	5	18	23	1.5	1.7	1.7
<i>Malvaceae</i>	6	14	20	1.8	1.3	1.5
<i>Violaceae</i>	7	10	17	2.1	1.0	1.2
<i>Geraniaceae</i>	5	11	16	1.5	1.1	1.2
<i>Liliaceae</i>	1	14	15	0.3	1.3	1.1

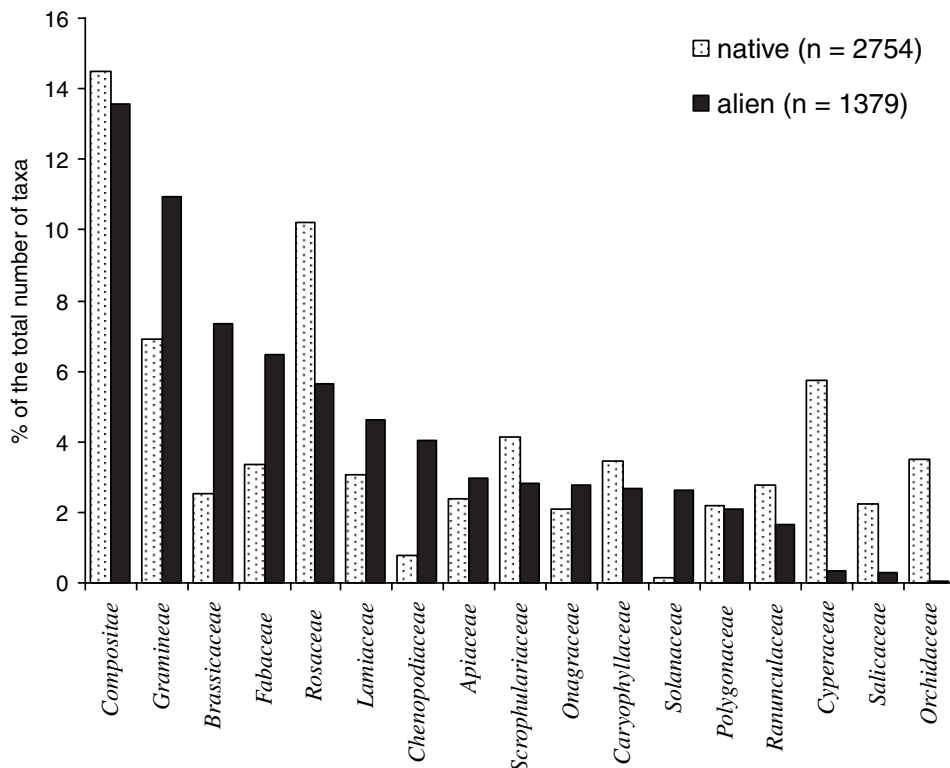


Fig. 5. – Comparison of taxonomic structure of alien and native floras. The most represented families are arranged according to the decreasing contribution to the alien flora. Data on native flora were taken from Kubát et al. (2002). Alien and native floras significantly differed with respect to the representation of plant families (G-test on contingency tables, $G = 865.41$, $df = 76$, $P < 0.001$).

As shown previously on a limited data set (Pyšek et al. 1995b), Grime's scheme of life strategies (Grime 1979) is a convenient predictor of invasive success. In contrasting environments, different life strategies are more likely to result in naturalization and invasion. The present data set shows that the C-strategy is a convenient one for naturalization but those species which possess combination of all three kinds of strategies have a better chance of becoming invasive (Table 4). The sometimes raised caution that the use of Grime's strategies brings about the danger of circular reasoning is not justified here since the invasive ability is a mixture of both capability to survive in disturbed habitats (typical of R-strategy) and to compete successfully (favoured by C-strategy). The question of which kind of life strategy favours invasion success is therefore a legitimate one, because the classification of species into particular strategies was not directly affected by the fact how good invader a species is.

As to the mode of introduction, 49.9% of all aliens were introduced into the country accidentally, and 42.7% deliberately; the remaining 7.4% were likely introduced by both means (Table 5). If the last group is not considered separately but the species belonging to it are considered in both accidental and deliberate category, accidental arrivals account for 53.4% of taxa and deliberate introductions for 46.6%. Since most archaeophytes reached the country as agricultural weeds, i.e. not on purpose of humans, the ratio for total aliens is

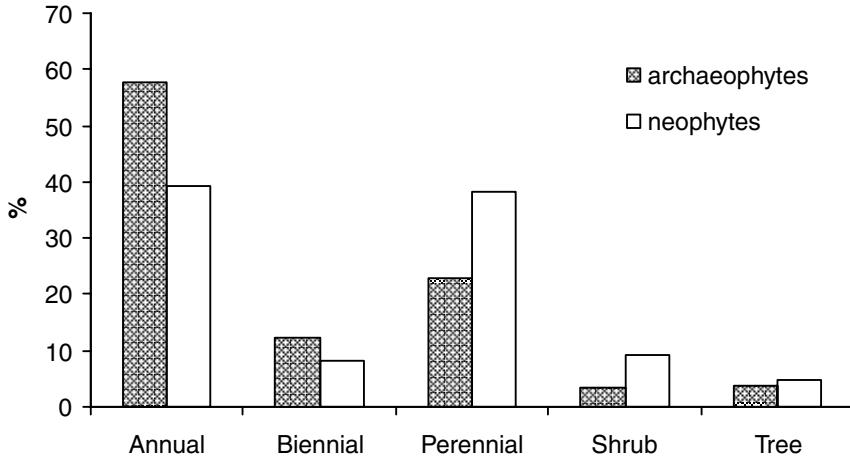


Fig. 6. – Distribution of life histories in archaeophytes and neophytes. Species known to occur as more than one life history were considered as representatives of each of them. Distribution of life histories in archaeophytes was significantly different from neophytes (G-test on contingency tables, $G = 64.24$, $df = 4$, $P < 0.001$).

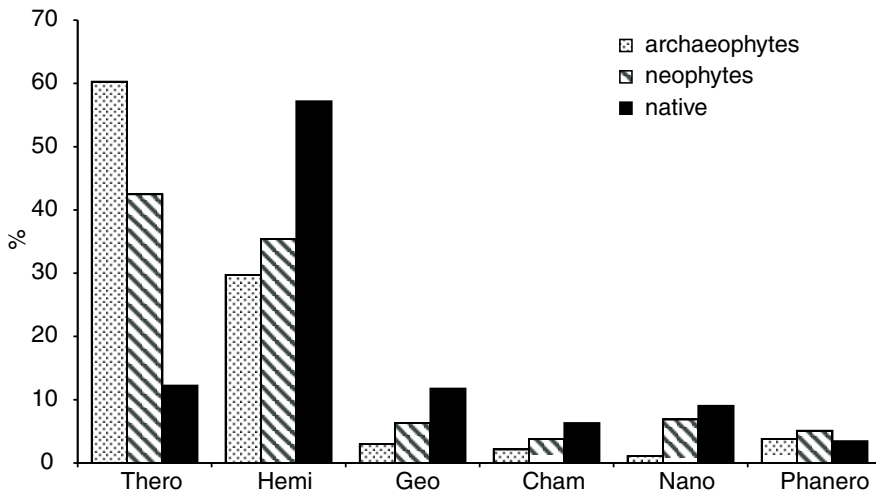


Fig. 7. – Distribution of Raunkiaer life forms (see Ellenberg 1988 for definitions) in archaeophytes, neophytes, and native flora. Species exhibiting more than one life form were considered as representatives of each of them. Data on native flora were taken from Kubát et al. (2002). Thero = therophytes, Hemi = hemicryptophytes, Geo = geophytes, Cham = chamaephytes, Nano = nanophanerophytes, Phanero = phanerophytes. Distribution of life forms was significantly different between archaeophytes and neophytes ($G = 52.83$, $df = 5$, $P < 0.001$), and between aliens and native ($G = 587.89$, $df = 5$, $P < 0.001$).

biased towards accidental introductions. Neophytes, on the contrary, include many taxa planted on purpose and escaping from cultivation, hence the ratio is reversed: more were introduced deliberately (54.5%) than accidentally (45.5%). More than a half of taxa are cultivated as ornamentals, other frequently encountered purposes are for food, medical purpose, landscaping, and bee-keeping (Table 6).

Table 4. – Distribution of Grime’s life strategies according to residence time, and with respect to invasive status in neophytes. Data shown are percentages based on 288 classified archeophytes and 611 neophytes. Distribution of life strategies was significantly different between archeophytes and neophytes ($G = 94.58$, $df = 6$, $P < 0.001$), and between aliens and native ($G = 49.95$, $df = 12$, $P < 0.001$).

	Archeophytes	Neophytes			
		Total	Casual	Naturalized	Invasive
C	15.3	38.8	33.1	58.1	47.4
CR	35.4	27.3	29.7	25.8	20.7
CS	2.8	5.9	5.6	4.8	7.4
CSR	8.3	8.0	6.8	6.5	12.6
R	31.3	15.2	20.0	1.6	6.7
S	0.0	2.6	2.7	0.0	3.7
SR	6.9	2.1	2.2	3.2	1.5

Table 5. – Structure of the Czech alien flora with respect to the presumed type of introduction into the country (deliberately or accidentally) and type of habitat. Number of taxa are shown for particular habitat/introduction categories. Natural and seminatural habitats (see text for definition) are grouped. Species occupying particular habitat types are significantly different with respect to the type of introduction (G -test on contingency tables, $G = 48.35$, $df = 4$, $P < 0.001$).

Type of habitat ¹⁰	Type of introduction			
	Accidental	Both ways	Deliberate	Total
Human-made habitats	486	61	315	862
Both types of habitats	147	34	178	359
Natural/seminatural habitats	51	7	93	151
Total	684	102	586	

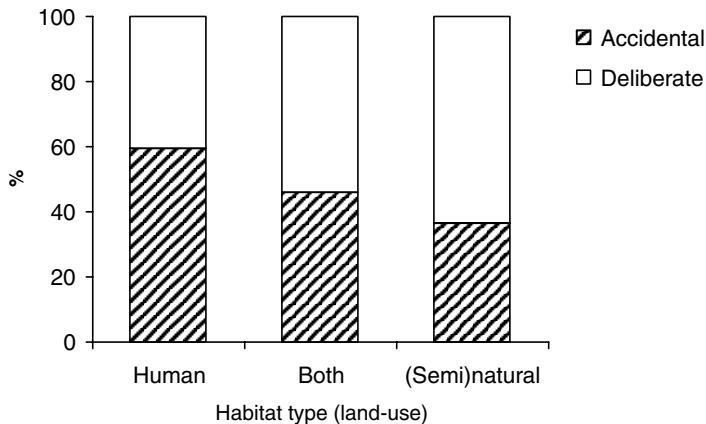


Fig. 8. – Type of introduction of alien species into habitat types classified according to the land-use. Species which were introduced into the country by both means (see Table 5) are included in both groups. See Table 5 for details on the grouping of habitats.

¹⁰ Human-made = H in Appendix 1; Both = NSH, SH; Natural/seminatural = N, S, NS

Table 6. – Deliberately introduced taxa of the Czech alien flora classified with respect to means of planting. Species with multiple planting purposes were considered in each of them.

Planting purpose	Species number	%
Ornamental	511	53.3
Food	149	15.5
Medical	99	10.3
Fodder	74	7.7
Landscaping	44	4.6
Bees	37	3.9
Oil	13	1.4
Wood	13	1.4
Dye	8	0.8
Textile	6	0.6
Agriculture (other than food)	5	0.5

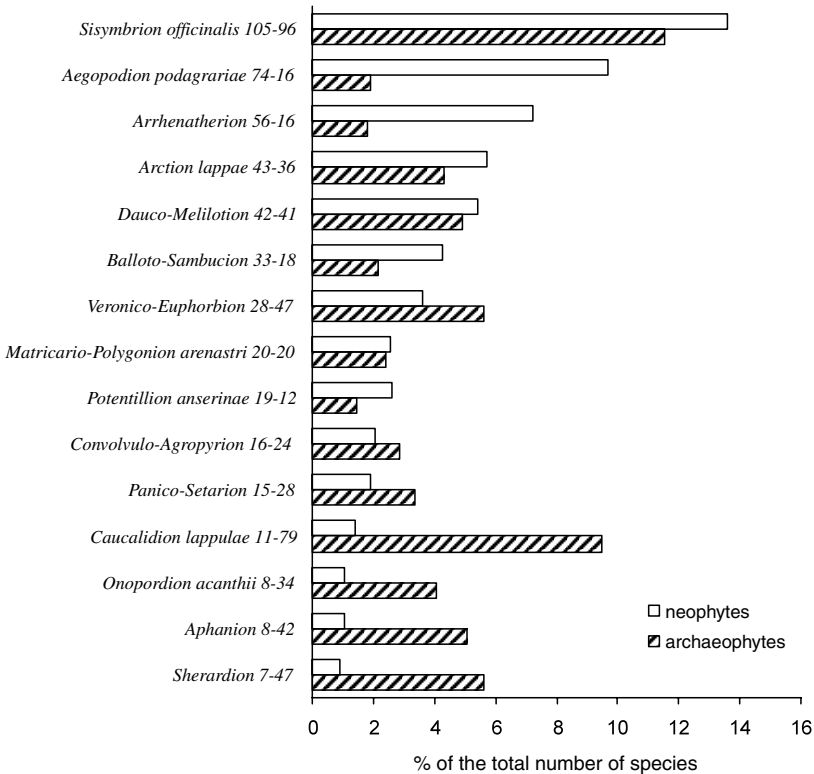


Fig. 9. – Occurrence of alien species in phytosociological units. Only the 10 alliances with the highest representation of neophytes and archaeophytes, respectively, are shown. Number of taxa of alien origin that commonly occur in a given phytosociological unit follow its name; the former value is the number of neophytes, the latter that of archaeophytes. Length of the bar represents the proportional contribution of the alliance to the total number of alien species in a given residence-time group. Rare and exceptional presence of species in alliances was not considered. Archaeophytes and neophytes significantly differ in their occurrence in phytosociological alliances (G-test on contingency tables, $G = 962.10$, $df = 177$, $P < 0.001$). See Appendix 1 for classification of particular species according to their occurrence in the alliances.

Majority of aliens (62.8%) are confined to human-made habitats, 11.0% were recorded exclusively in natural and/or seminatural habitats, and 26.2% occur in both types of habitat (Table 5). Plants which were introduced deliberately more often invade seminatural and natural habitats than taxa arriving by accidental means: 63.3% of aliens recorded in either natural or seminatural types of habitats are or used to be planted in the past whereas the corresponding figure for human-made habitats is only 40.7% (Fig. 8).

Archaeophytes and neophytes occur in 66 and 83 alliances, respectively, of the phytosociological system; alien species as a whole are present in 91 alliances (classified according to Chytrý et al. 2001). Some vegetation types, such as *Sisymbrium officinalis*, *Dauco-Melilotion*, and *Arction lappae* harbour species of both groups with a comparable frequency (Fig. 9). *Aegopodium*, *Arrhenatherion* (including its ruderalized stands), and *Balloto-Sambucion* are alliances with more neophytes than archaeophytes present while *Caucalidion lappulae*, *Onopordion acanthii*, *Aphanion*, and *Sherardion* are units containing high number of archaeophytes. Alien species are thus concentrated in vegetation of deforested mesic habitats with frequent disturbances such as rubbish tips, waste land, arable land, or fringe communities.

Hybridization is an important event contributing to the diversity in alien floras (Vilà et al. 2000). In the Czech flora, hybrids contribute 13.3% to the total number of aliens, and the hybridization is more frequent in archaeophytes (18.7%) than in neophytes (11.7%). Sixty six crosses of aliens with native taxa were recorded (Table 7). If hybrids are excluded from the total number of aliens, there are 270 archaeophytes and 924 neophytes in the Czech flora. The extent of hybridization is difficult to compare with other regions since, as pointed out by Vilà et al. (2000), the number of hybrids reported reflects the level of detail aimed at by particular floras. British flora has been reported to include 70 hybrids between an introduced and native species and 21 hybrids between two introduced species. In Ontario, there are 31 hybrids directly introduced or resulting from hybridization among introduced species. However, the quantitative data are rather scarce. Whereas hybridization between native species may produce novel genotypes and increase genetic diversity at both the population and species level, spontaneous hybridization involving alien species may have the reverse effects and threaten the genetic integrity and persistence of native species (Vilà et al. 2000). Range expansion of hybrids can be rapid and hybrids can become weeds (Abbott 1992). *Reynoutria xbohemica* can serve as an example from the territory of the Czech Republic (Bímová et al. 2001).

Majority of aliens are diploids and tetraploids. In neophytes, there are more species with high ploidy levels compared to archaeophytes. Native flora has, compared to aliens, lower proportion of diploids and higher representation of tetraploids, triploids and pentaploids (Table 8).

Only 310 species (22.4% of the total number of all alien taxa) are common or locally abundant; others are rare, based on a single locality or no longer present (Fig. 10). The proportion of neophytes represented by a single locality (14.3%) indicates the importance of chance in records of alien flora. There are species which qualified for the list on the basis of the successful establishment of a single plant on a single locality. For example, *Rumex brownii* and *R. dentatus* subsp. *halacsyi* were present in their localities as single, fruitful specimens which ended up in a herbarium. What would happen if these plant got a chance to spread their seed and became a potential founder of a population? Other species, e.g. *Datura ferox*, *Polypogon fugax* or *Alhagi pseudalhagi* were also included on the basis of

Table 7. – Overview of hybridization in the Czech alien flora. Numbers of hybrid taxa, classified according to the immigration time or native status of their parents and hypothesized to occur at the territory studied are shown. Hybrid arrivals are crosses and hybridogenous species which originated outside the territory of the Czech Republic.

	Number of hybrid taxa with			Species originated in cultivation	Hybrid arrivals	Total number of hybrids	% of total number of taxa
	archaeophytes	neophytes	native				
Archaeophytes	12		31	12	7	62	18.7
Neophytes	6	28	35	32	21	122	11.7
Aliens total				44	28	184	13.3

a find of a single plant. These numbers demonstrate that quantifying biological invasions is a difficult task, and hunting for the number of casuals is probably the hardest part of it! Also, these examples show that the role of humans is crucial in every step of the process and sometimes difficult to predict (Kowarik 2002).

Comparison of the distribution of the number of localities between archaeophytes and neophytes reveals an opposite pattern which reflects historical consequences (Table 9). Most archaeophytes are rather frequent; 72.3% are supposed to have more than 50 localities.

Some rare archaeophytes are on the Red List of Czech flora: *Ajuga chamaepitys*, *Arnoseris minima*, *Bromus arvensis*, *B. commutatus*, *B. secalinus*, *Bupleurum rotundifolium*, *Galium tricorntum*, *Linaria arvensis*, *Kickxia spuria* subsp. *spuria*, *K. elatine* subsp. *elatine*, *Lolium remotum*, *L. temulentum*, *Marrubium peregrinum*, *M. vulgare*, *Papaver lecoqii*, *Polycnemum arvense*, *P. majus*, *Sagina apetala*, *Stellaria pallida*, *Veronica opaca*, *V. agrestis*, *V. triloba* (Holub & Procházka 2000).

Table 8. – Overview of ploidy levels in Czech alien flora. Note that the data were not available for all taxa hence the species totals are lower than for other presented characteristics. Data on chromosome numbers of native flora were taken from Kubát et al. (2002) where available (n = 2005). Archaeophytes and neophytes are not significantly different with respect to representation of ploidy levels (G-test on contingency tables, G = 0.71, df = 3, NS, ploidy levels with at least 5 taxa were considered). Aliens significantly differed from native taxa in distribution of ploidy levels (G=120.15, df 7, P<0.001).

Ploidy level	Number of taxa			%		
	Archaeophytes	Neophytes	Native	Archaeophytes	Neophytes	Native
2x	183	427	896	64.2	60.3	44.7
3x		9	105		1.3	5.2
4x	67	164	644	23.5	23.2	32.1
5x	2	1	49	0.7	0.1	2.4
6x	23	56	174	8.1	7.9	8.7
7x	1	1	4	0.4	0.1	0.2
8x	8	26	82	2.8	3.7	4.1
> 8x	1	24	51	0.4	3.4	2.5

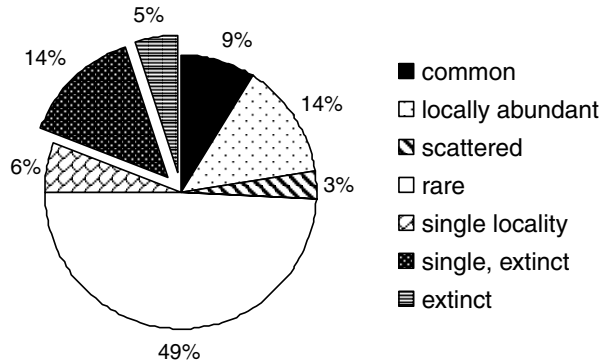


Fig. 10. – Distribution of the types of abundance in alien flora (see text for explanation). Categories including “extinct” species are disconnected.

Table 9. – Distribution of alien flora according to their abundance expressed as the number of localities (see text for details). Abundance scale corresponds to that used in Clement & Foster (1994) and Ryves et al. (1996). Archaeophytes and neophytes differed significantly with respect to their distribution of abundance (G-test on contingency tables, $G = 477.78$, $df = 4$, $P < 0.001$).

Number of localities	Number of taxa			%		
	Archaeophytes	Neophytes	Aliens total	Archaeophytes	Neophytes	Aliens total
1–4	22	571	593	6.6	54.6	43.1
5–14	36	208	244	10.8	19.9	17.7
15–49	34	124	158	10.2	11.8	11.5
50–499	68	72	140	20.5	6.9	10.2
> 499	172	71	243	51.8	6.8	17.6

Potential use of databases of alien species: what are they good for?

Databases of alien species have, in addition to the historical value (they can be used for future comparisons), several other functions. (i) The scientific importance lies in the possibility to generate hypotheses about the effect of species characters on probability of naturalization and invasive success (Crawley et al. 1996, Pyšek et al. 1995b). (ii) Prediction possibilities. Pyšek (2001) has shown that prediction systems screening species on the basis of the number of their characteristics (Daehler & Carino 2000) can be very powerful, definitely more so than those based on mere intuition and autecological knowledge. (iii) Regional databases represent stones for a mosaic of databases covering larger geographical areas. A European-scale project of a continental database of alien species could make use of sharing the information on species characteristics, consolidating the measures of naturalization and invasiveness and providing information on potentially arriving species into the country prior to their naturalization. Such better sharing of information might contribute to adopting appropriate measures in advance rather than after the invasion has started. To our knowledge, the database presented here is one of the first of that kind in terms of taxonomical, ecological and geographical detail. For that reason, its detailed structure is presented as a suggestion for the work of similar kind (Appendix 2).

Acknowledgments

Very special thanks are due to K. Kubát, V. Řehořek, V. Grulich, J. Chrtek jun., J. Hadinec, M. Štech, J. Zázvorka, and M. Pejchal for providing many unpublished data and sharing their knowledge. Comments of V. Řehořek, K. Kubát and D. M. Richardson on various aspects of the manuscripts are greatly appreciated. V. Jarošík kindly conducted statistical analyses. Other colleagues who contributed with information on particular species are listed in alphabetical order: J. Burda, P. Bauer, P. Bureš, M. Dančák, J. Danihelka, T. Francírková, H. Gruberová, H. Härtel, P. Havlíček, L. Hrouda, V. Chán, J. Chrtek sen., J. Jedlička, V. Jehlík, Z. Kaplan, J. Kirschner, L. Kirschnerová, P. Kovář, F. Krahulec, M. Král, B. Křísa, M. Lepší, K. Martínek, S. Mihulka, E. Opravil, P. Petřík, P. Pokorný, K. Prach, F. Procházka, †A. Pyšek, Bohumil Slavík, K. Sutorý, P. Špryňar, O. Šída, J. Štěpánek, J. Štěpánková, B. Trávníček, J. Vicherek, W. L. Wagner, and E. Weber. David M. Richardson and John H. Brock kindly improved our English. Logistic support from I. Ostrý, J. Pergl, and I. Koukolíková is highly appreciated. The project was made possible by a grant no. 206/99/1239 from the Grant Agency of the Czech Republic, and partly supported by a grant no. AV0Z6005908 from the Academy of Sciences of the Czech Republic.

Souhrn

Práce je pokusem podat kompletní přehled nepůvodních (člověkem zavlečených, adventivních) druhů české flóry, které se vyskytují či v minulosti vyskytovaly ve volné přírodě; nezahrnuje tedy druhy pěstované, jež nezplaňují. Jsou zahrnuty pouze druhy, které jsou nepůvodní na celém území ČR; pokud má druh v určité části země, případně na ekologicky specializovaném stanovišti, původní výskyt, není považován za zavlečený, byť byl jinak pěstován a zplaňoval. Přehled zavlečených druhů je uveden v Appendixu 1; jsou klasifikovány podle několika kritérií. Postavení druhu v invazním procesu odpovídá třídění navrženému v práci Richardsona et al. (2000) a je vyjádřeno následovně: náhodný výskyt (odpovídá anglickému termínu „casual“) – druh se ve volné přírodě pravidelně nereprodukuje a pokud se v krajině vyskytuje v delším časovém horizontu, je závislý na opakovaném, člověkem zprostředkovaném přísunu disper; naturalizace – druh se ve volné přírodě rozmnožuje, generativně či vegetativně, jeho výskyt není závislý na dalších introdukcích a jeho přítomnost na určité lokalitě či v určitém území je dosti trvalý; invaze – druh se v krajině šíří a vytváří více či méně rozsáhlé populace. V Appendixu 1 jsou dále označeny druhy, které považujeme za „postinvazní“; invaze u nich proběhla v minulosti a v současné době se již nešíří (vzhledem ke své náplni je tento termín zatížen větší názorovou subjektivitou, než termíny standardního členění). Jsou také označeny druhy, u nichž má výskyt výrazný charakter pozůstatku z dřívějšího pěstování na dotyčné lokalitě; přesto jsou i tyto druhy zahrnuty, pokud se na lokalitách udržují po mnoho let a nezdá se výrazně rozrůstat.

Další použitá kritéria jsou, zda se jedná o archeofyt či neofyt (tedy druh zavlečený před objevením Ameriky nebo až poté); náplň pojmu „neofyt“ je v zájmu jasnější terminologie poněkud posunuta proti dřívějšímu chápání (např. Holub & Jirásek 1967) v tom smyslu, že za neofyty považujeme všechny druhy zavlečené po roce ca. 1500, bez ohledu na to, zda k tomu došlo úmyslně či náhodně. Způsob zavlečení (úmyslné nebo náhodné), typ stanoviště (původní, polopřirozená, antropogenní), společenstva, ve kterých se druh vyskytuje (na úrovni svazů curyško-montpelliérského systému) a kontinent, ze kterého pochází, jsou dalšími charakteristikami uvedenými v Appendixu 1. Abundance druhu na území ČR byla hodnocena pomocí pětičlenné semikvantitativní stupnice založené na odhadu počtu lokalit (Clement & Foster 1994): 1–4, 5–14, 15–49, 50–499, 500 a více lokalit. Je uveden také rok prvního známého výskytu z území ČR. K 19 druhům, které jsou udávány z našeho území jako nové jsou v Appendixem 1 připojeny komentáře. Jedná se o *Agrostis scabra*, *Alhagi pseudalhagi*, *Allium atropurpureum*, *Bromus hordeaceus* subsp. *pseudohominii*, *Carduus tenuiflorus*, *Centaurea xgerstlaui*, *Centaurea nigra* × *phrygia*, *Cerastium xmaureri*, *Gilia capitata*, *Helianthus strumosus*, *Hieracium pannosum*, *Hordeum leporinum*, *Oenothera coronifera*, *Papaver atlanticum* subsp. *mesatlanticum*, *Parietaria pennsylvanica*, *Polypogon fugax*, *Rodgersia aesculifolia*, *Sedum pallidum* var. *bithynicum* a *Sedum stoloniferum*. Pro dalších 44 taxonů jsou uvedeny první údaje o zplanění.

Adventivní flóra ČR obsahuje celkem 1378 taxonů patřících do 542 rodů a 99 čeledí; z toho je 184 kříženců nebo hybridogenních taxonů. Podíl zavlečených taxonů na flóře ČR činí 33,4 %. Pokud z hodnocení vyjmeme křížence adventivních i původních druhů, činí tento podíl 34,6 %. Flora obsahuje 332 archeofytů a 1046 neofytů; 892 taxonů je považováno za náhodně se vyskytující, 397 za naturalizované a 90 za invazní (tab. 1). Z celkového počtu 1046 neofytů došlo k naturalizaci u 229 druhů (21,9%) a z nich je 69 invazních (tj. 6,6 % z celkového počtu introdukcí). Naopak 231 náhodně se vyskytнувších neofytů z flóry vymizelo.

Většina archeofytů pochází ze Středozeší; neofyty mají svůj původ převážně v ostatních částech Evropy (39,8 %) a Asie (27,6 %) a v Severní Americe (15,1 %). Z čeledí jsou nejzastoupenější *Compositae*, *Gramineae* a *Brassicaceae* (tab. 3). Objevují se v tomto ohledu i určité rozdíly mezi archeofyty a neofyty: *Chenopodiaceae*, *Apiaceae*, *Scrophulariaceae* a *Caryophyllaceae* mají více archeofytů, zatímco *Fabaceae*, *Solanaceae*, *Polygonaceae*,

Onagraceae a *Amaranthaceae* představují typické “neofytní” čeledi. Mezi rody s největším počtem nepůvodních taxonů patří *Chenopodium* (27), *Amaranthus* (24), *Oenothera* (23), *Bromus* (21) a *Vicia* (18).

Jednoleté druhy tvoří 57,8 % všech archeofytů, zatímco vytrvalé bylinné druhy (38,2 %) a dřeviny (14,1 %) jsou častěji zastoupené mezi neofyty (obr. 3). Celkem česká adventivní flora sestává z 44,0 % jednoletých, 9,3 % dvouletých, 34,4 % vytrvalých bylin, 7,7 % keřů a 4,5 % stromů. 49,9 % všech taxonů se na území ČR dostalo bez úmyslného přispění člověka, 42,7 % bylo zavlečeno úmyslně; na zavlečení zbývajících 7,4 % se podílely oba způsoby. U neofytů hodnocených samostatně je tento poměr posunut ve prospěch záměrných introdukcí (54,5 %).

Většina druhů (62,8 %) je vázána na antropogenní stanoviště; 26,2 % se vyskytuje jak na člověkem vytvořených, tak na přirozených či polopřirozených stanovištích a 11,0 % (151 druhů) bylo zaznamenáno pouze na (polo) přirozených typech stanovišť (tab. 5). Rostliny introdukované záměrně se objevují častěji v přirozené vegetaci než druhy zavlečené neúmyslně (obr. 7). Archeofyty se objevují ve vegetaci patřící do 66 svazů curyško-montpelliérského systému, neofyty v 83 svazech. *Sisymbrium officinalis*, *Dauco-Melilotion* a *Arction lappae* hostí stejně často druhy obou skupin; *Aegopodion*, *Arrhenatherion* a *Balloto-Sambucion* jsou svazy typické výskytem neofytů; archeofyty jsou soustředěny především ve vegetaci svazů *Caucalidion lappulae*, *Onopordion acanthii*, *Aphanion* a *Sherardion* (obr. 8).

Kříženci a hybridogenní taxony tvoří 13,3 % celkového počtu nepůvodních taxonů; kříženci archeofytů (18,7 %) jsou přitom častější než kříženci neofytů (11,7 %). Bylo zaznamenáno 66 kříženců nepůvodních druhů se zástupci domácí flóry. Vyloučením hybridů dospějeme k celkovému počtu 1194 taxonů (270 archeofytů, 924 neofytů).

Dvacet dva archeofytů je na Červeném seznamu české flóry (Holub & Procházka 2000): *Ajuga chamaepitys*, *Arnosaris minima*, *Bromus arvensis*, *B. commutatus*, *B. secalinus*, *Bupleurum rotundifolium*, *Galium tricorneratum*, *Linaria arvensis*, *Kickxia spuria* subsp. *spuria*, *K. spuria* subsp. *elatine*, *Lolium remotum*, *L. temulentum*, *Marrubium peregrinum*, *M. vulgare*, *Papaver lecoqii*, *Polycnemum arvense*, *P. majus*, *Sagina apetala*, *Stellaria pallida*, *Veronica opaca*, *V. agrestis* a *V. triloba*.

Katalog je nutno chápat jako první práci svého druhu pro území ČR; údaje v ní obsažené budou postupně upřesňovány a autoři budou vděční za jakékoli připomínky a doplňky. Při sestavování katalogu jsme vycházeli ze základních florových děl vztahujících se k území ČR i z primární literatury. Obtížnost klasifikace se projevuje při hodnocení mnoha hraničních kategorií, zejména při rozhodování, zda je druh původní či archeofyt, archeofyt či neofyt, náhodně se vyskytující či naturalizovaný, naturalizovaný či invazní. Statut každého druhu byl důkladně přehodnocen a třebaže hojně konzultován s řadou kolegů, v konečném důsledku odráží především náš názor na historické postavení dotyčného druhu v naší krajině.

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¹¹ The bibliography, although not aspiring to be complete, considers majority of important literature on taxonomy, biogeography and autecology of particular alien species in the Czech Republic; other sources can be found in the Flora of the Czech Republic (Hejný & Slavík 1988–1992, Slavík 1995–2000). For that reason, the list of references includes also sources not explicitly referred to in the text. All the sources were, however, used for compiling the database.

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Received 14 January 2002

Revision received 28 March 2002

Accepted 15 April 2002

Appendix 1. – List of alien taxa of the Czech flora. Species are arranged alphabetically. **Family** codes (Fam) are formed by initial letters of the family name. The following information is given for each species, if available: **Invasive status** (Stat): cas = casual, nat = naturalized, inv = invasive. Post-invasive status (see text for explanation) is indicated by an asterisk. Species which are supposed to grow in the wild as relics of former cultivation (in all or majority of their localities) are followed by #. **Residence time** (Res): ar = archaeophyte; if known, period of the earliest evidence is indicated: N – Neolithic/Aeneolithic period (5300–2200 B. C.), B – Bronze Age (2200–750 B. C.), I – Iron Age (750–0 B. C.), R – Roman period and Migration period (0–550), P – prehistoric times (5300 B.C.–550), M – Medieval period (550–1500); neo = neophyte. **Date** of the first reported occurrence in the wild (1st). Habitat type is expressed as combination of two criteria: 1. **Land-use**: N = natural habitats, i.e. natural forests and naturally treeless habitats; S = seminatural habitats, i.e. managed landscape except of settlements, communications, and arable land; H = human-made habitats (Chytrý et al. 2001). 2. **Landscape** classification: T = traditional agricultural landscape; M – modern urban and industrial landscape. For some species, there were not enough data to classify the habitat type. **Syntaxa** in which the species occurs; alliances of the Zurich-Montpellier school are listed and their codes are explained below (see Chytrý et al. 2001 for authors' names). **Abundance type** in the wild at the territory of the country (Abund): s = single locality, r = rare, sc = scattered, la = locally abundant, c = common, e = extinct (if no records have been known for a long period), se = single locality, now extinct. LocNo: quantitative estimate of the **number of localities** using the scale of Clement & Foster (1994): 1 = 1–4 localities; 2 = 5–14; 3 = 15–49; 4 = 50–499; 5 = over 500 localities. The system used here is based on the number of separate localities from which the species has been recorded, in print, on herbarium labels, or privately communicated during the 20th century; above 15, the number of localities is an estimate. Minimum arbitrary distance between localities in order to be considered as separate was 1 km. **Introduction mode** (Intr) of the species into the country: d = deliberate (by planting); a = accidental; ad = both means. For hybrids, those spontaneously originated at the territory of the Czech Republic are considered as “accidental”, whereas hybrids escaped from cultivation are considered “deliberate”. **Origin**: E = Europe, AS = Asia, AMN = North America, AMC = Central America, AMS = South America, AF = Africa, AU = Australia; if not given, the taxon is of a hybrid origin or it is obscure. **Life history** (LH): a = annual, af = annual fern, ap = parasitic or semiparasitic annual, b = biennial, bp = parasitic biennial, pe = perennial, ss = semi-shrub, s = shrub, t = tree, f = fern (in multiple entries, the life form more common at the territory is given first). Species in which the life form is not possible to determine such as hybrids of parents belonging to different life forms are indicated by questionmark. **Source**: F – Flora of the Czech Republic, Vol. 1–6 (Hejny & Slavík 1988–1992, Slavík 1995–2000), K – Key to the flora of the Czech Republic (Kubát et al. 2002); as accounts published in the Flora are more detailed, reference to the Key is only given if the Flora does not cover the taxon. Additional source is only given if there is a detailed specialized literature account on the species or if the species is reported neither in F nor in K. Detailed information on taxa which represent either additions to the Czech flora or the first record of escape from cultivation is given at the end of the Appendix together with other remarks on e.g. invasion status, history, or taxonomy.

Codes of syntaxa: **Ab** – *Arabidopsis thalianae*; **Ad** – *Adenostylion*; **Ae** – *Aegopodium podagrariae*; **AF** – *Alyso-Festucion pallentis*; **Ah** – *Aphanion*; **Ai** – *Alnion incanae*; **Al** – *Arction lappae*; **An** – *Alnion glutinosae*; **Ap** – *Alopecurion pratensis*; **AQ** – *Aceri tatarici-Quercion*; **Ar** – *Arrhenatherion*; **AS** – *Alyso alyssoidis-Sedion albi*; **At** – *Atropion*; **Bd** – *Berberidion*; **Bf** – *Batrachion fluitantis*; **Bi** – *Bidention tripartitae*; **BR** – *Balloto nigrae-Robinion*; **Br** – *Bromion erecti*; **BS** – *Balloto-Sambucion*; **CA** – *Convolvulo-Agropyron*; **Cb** – *Chenopodion rubri*; **CE** – *Carici piluliferae-Epilobion angustifolii*; **Cl** – *Caucalidion lappulae*; **Cm** – *Cymballario-Asplenion*; **CM** – *Cardamino-Montion*; **Co** – *Corynephorion canescentis*; **Cr** – *Carpinion*; **CR** – *Chelidonio-Robinion*; **Ct** – *Calthion*; **Cy** – *Cynosurion*; **DM** – *Dauco-Melilotion*; **DS** – *Diantho lumitzeri-Seslerion albicantis*; **EC** – *Euphorbio-Callunion*; **Er** – *Eragrostion*; **Es** – *Eleocharition soloniensis*; **Fg** – *Fagion*; **Fv** – *Festucion valesiacae*; **GA** – *Galio-Alliarion*; **Ge** – *Geniston*; **GQ** – *Genisto germanicae-Quercion*; **Gs** – *Geranion sanguinei*; **HF** – *Helianthemum cani-Festucion pallentis*; **HS** – *Hyperico perforati-Scleranthion perennis*; **IS** – *Impatienti-Stachyion sylvaticae*; **KP** – *Koelerio-Phleion phleoidis*; **Le** – *Lemnon minoris*; **LF** – *Luzulo-Fagion*; **Ma** – *Magnocaricion elatae*; **Mn** – *Malvion neglectae*; **Mp** – *Magnopotamion*; **MP** – *Matricario-Polygonion arenastris*; **Na** – *Nardion*; **NA** – *Nardo-Agrostion tenuis*; **Nc** – *Nanocyperion flavescens*; **NJ** – *Nardo-Juncion squarrosi*; **Oa** – *Onopordion acanthii*; **Pa** – *Potentillion anserinae*; **Pe** – *Petasition officinalis*; **PF** – *Plantagini-Festucion ovinae*; **Ph** – *Phalaridion arundinaceae*; **Pp** – *Parvopotamion*; **Pr** – *Phragmition*; **PR** – *Pruno-Rubion radulae*; **Ps** – *Panicco-Setarion*; **Ps** – *Prunio spinosae*; **PT** – *Polygono-Trisetion*; **Qp** – *Quercion pubescenti-petraeae*; **Qt** – *Quercion petraeae*; **Ra** – *Rumicion alpini*; **Sa** – *Salicion albae*; **Sc** – *Stipion calamagrostis*; **Se** – *Salicion elaeagno-daphnoidis*; **Sf** – *Senecion fluviatilis*; **SG** – *Sparganio-Glycerion fluitantis*; **Sg** – *Saginion procumbentis*; **Sh** – *Sheraldion*; **Si** – *Sisymbrium officinalis*; **SJ** – *Scorzonero-Juncion gerardii*; **Sn** – *Scleranthion annui*; **SO** – *Spergulo-Oxalidion*; **Sr** – *Salsolion ruthenicae*; **SS** – *Sambuco-Salicion caprae*; **St** – *Salicion triandrae*; **Sx** – *Salicion incanae*; **TA** – *Tilio-Acerion*; **Th** – *Thero-Airion*; **Tm** – *Trifolion medii*; **Vc** – *Violion caninae*; **Ve** – *Veronico-Euphorbion*; **VT** – *Veronico politae-Taraxacion*.

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Abutilon theophrasti</i> Med.	Mal	cas	neo	1894	H	TM
<i>Acer ginnala</i> Maxim.	Ace	cas	neo	2001	H	M
<i>Acer monspessulanum</i> L.	Ace	cas	neo	2001	H	M
<i>Acer negundo</i> L.	Ace	inv	neo	1875	NSH	TM
<i>Acer saccharinum</i> L.	Ace	cas	neo		H	TM
<i>Achillea crithmifolia</i> W. et K.	Com	cas	neo	1886	S	T
<i>Achillea filipendulina</i> Lamk.	Com	cas	neo	1986	H	M
<i>Achnatherum calamagrostis</i> (L.) P. B.	Gra	cas	neo		N	T
<i>Aconitum xcammarum</i> L.	Ran	nat*	neo	1819	NSH	T
<i>Acorus calamus</i> L.	Ara	nat	neo	1809	NS	T
<i>Acroptilon repens</i> (L.) DC.	Com	cas	neo	1962	H	M
<i>Adonis aestivalis</i> L.	Ran	nat*	arB		H	T
<i>Adonis annua</i> L. subsp. <i>annua</i>	Ran	cas	neo	1874	H	T
<i>Adonis flammea</i> Jacq.	Ran	nat*	ar		H	T
<i>Aegilops cylindrica</i> Host	Gra	cas	neo		H	TM
<i>Aegilops geniculata</i> Roth	Gra	cas	neo		H	TM
<i>Aesculus xcarnea</i> Hayne	Hip	cas	neo	1963	H	M
<i>Aesculus hippocastanum</i> L.	Hip	cas	neo		SH	TM
<i>Aethusa cynapium</i> L.	Api	nat*	arN		H	T
<i>Ageratum houstonianum</i> Mill.	Com	cas	neo		H	TM
<i>Agropyron pectinatum</i> (M. Bieb.) P. B.	Gra	cas	neo	1823	H	M
<i>Agrostemma githago</i> L.	Car	nat*	arN		H	T
<i>Agrostis gigantea</i> Roth	Gra	nat	neo		H	TM
<i>Agrostis scabra</i> Willd.	Gra	cas	neo	2001	H	T
<i>Ailanthus altissima</i> (Mill.) Swingle	Sim	inv	neo	1874	NSH	TM
<i>Ajuga chamaepitys</i> (L.) Schreber	Lam	nat*	ar		H	T
<i>Ajuga glabra</i> C. Presl	Lam	nat	ar		H	T
<i>Alcea rosea</i> L.	Mal	nat	neo	1880	H	TM
<i>Alchemilla conjuncta</i> Bab.	Ros	cas#	neo		S	T
<i>Alchemilla mollis</i> (Buser) Rothm.	Ros	cas	neo	1985	H	TM
<i>Alchemilla sericata</i> Reichenb.	Ros	cas	neo		H	TM
<i>Alchemilla speciosa</i> Buser	Ros	cas	neo		H	TM
<i>Alchemilla tythantha</i> Juz.	Ros	cas	neo		SH	TM
<i>Alliagi pseudalliagi</i> (M. Bieb.) Desv.	Fab	cas	neo	1963	H	M
<i>Allium atropurpureum</i> W. et K.	Alli	cas	neo	1946	S	T
<i>Allium atroviolaceum</i> Boiss.	Alli	cas	neo	1922	S	T
<i>Allium cepa</i> L.	Alli	cas	neo		H	TM
<i>Allium fistulosum</i> L.	Alli	cas	neo		H	TM
<i>Allium moly</i> L.	Alli	cas	neo		H	T
<i>Allium paradoxum</i> (M. Bieb.) G. Don	Alli	nat	neo	1867	NSH	T
<i>Allium porrum</i> L.	Alli	cas	neo		H	TM
<i>Allium sativum</i> L.	Alli	nat	ar		SH	TM
<i>Allium tuberosum</i> Rottl. ex Spreng.	Alli	nat	neo		S	T
<i>Alnus rugosa</i> (Duroi) Sprengel	Bet	nat	neo	1872	NS	T
<i>Alopecurus myosuroides</i> Huds.	Gra	nat	ar		H	TM
<i>Althaea armeniaca</i> Ten.	Mal	cas	neo	1966	H	M
<i>Althaea hirsuta</i> L.	Mal	cas	neo	1870	H	T
<i>Alyssum murale</i> W. et K.	Bra	nat	neo		H	M
<i>Alyssum rostratum</i> Steven	Bra	cas	neo	1897	H	M
<i>Amaranthus xalleizettei</i> Aellen	Ama	cas	neo	1945	H	M
<i>Amaranthus acutilobus</i> Uline et Bray	Ama	cas	neo	1909	H	TM
<i>Amaranthus albus</i> L.	Ama	nat	neo	1893	SH	TM
<i>Amaranthus blitoides</i> S. Watson	Ama	nat	neo	1931	H	TM
<i>Amaranthus blitum</i> L.	Ama	nat	ar		H	TM
<i>Amaranthus bouchonii</i> Thell.	Ama	cas	neo	1948	H	M
<i>Amaranthus caudatus</i> subsp. <i>saueri</i> Jehlík	Ama	cas	neo	1838	H	M
<i>Amaranthus crispus</i> (Lesp. et Thév.) N. Terracc.	Ama	cas	neo	1926	H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Si Er	r	3	ad	AS	a	F, Hejný et al. 1973, Jehlík 1998
	s	1	d	AS	st	F
	r	1	d	E	t	F
Ai Sa BS CR	la	5	d	AMN	t	F
	s	1	d	AMN	t	F
	r	1	a	E	pe	K
	se	1	d	AS	pe	K, Sutorý 1993
	r	1	a	E	pe	K
Ae Ph Pe	sc	4	d		pe	F
Pr	sc	5	a	AS	pe	K, Pyšek & Mandák 1998a
	r	1	a	E AS	pe	K, Hejný et al. 1973, Jehlík 1998
Cl	sc	5	a	E AS AF	a	F
	r	2	d	E AF	a	F
Cl	r	3	a	E AS	a	F
	r	1	a	E	a	K
	r	1	a	E	a	K
	r	1	d		t	F
BS	la	4	d	E	t	F
Sh	c	5	a	E AS	a	F
	r	1	d	AMC AMS	pe	K
	r	1	a	E	pe	K
Sh Cl Ah	r	4	a	E AS	a	F
Pa Si Ph SJ	sc	4	a	E AS	pe	K
	s	1	a	AMN	pe	
SS	sc	4	d	AS	t	F
Cl Oa DM	r	4	ad	E AF	a b	F
Cl Oa VE	e	1	a	E AS	a b pe	F
Pa Al	r	4	d	E AS	b pe	F
	s	1	d	E	pe	F
	r	2	d	E AS	pe ss	F
	s	1	d	E AS	pe	F
	s	1	d	E	pe ss	F
	r	1	d	AS	pe	F
	se	1	a	E AS	pe	
	s	1	d	E AS	pe	Krahulec in prep.
	se	1	a	E AS	pe	Dostál 1948-1950, Krahulec in prep.
Si VE	r	4	d	AS	pe	K
	r	2	d	AS	pe	K
	r	2	d	E	pe	K
	r	2	d	E AS	pe	K, Hejný 1971, Hejný et al. 1984
	r	2	d		pe	K
VE CA	sc	4	d	AS	pe	K
	se	1	d	AS	pe	Dostál 1948-1950, Krahulec in prep.
	r	2	d	AMN	s	F
	r	2	a	E AS	a	K, Jehlík 1998
	e	1	a	E AS	pe	F, Smejkal 1966
	e	1	a	E AS	a	F
	r	2	d	E	pe	F
	e	1	a	E	a	F
	r	1	a		a	F
	e	1	d	AMN	a	F
Si PS Er	sc	4	a	AMN	a	F, Hejný et al. 1973, Jehlík 1998
Mn MP	sc	4	a	AMN	a	F, Hejný et al. 1973, Jehlík 1998
Mn VE	sc	4	a	E AF	a	F
Si	e	1	a	AMN	a	F
Si	sc	3	d	AMS	a	F
MP	r	2	a	AMS	a	F

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Amaranthus cruentus</i> L.	Ama	cas	neo	1834	H	TM
<i>Amaranthus deflexus</i> L.	Ama	cas	neo	1905	H	M
<i>Amaranthus graecizans</i> L. subsp. <i>graecizans</i>	Ama	cas	neo	1912	H	M
<i>Amaranthus graecizans</i> subsp. <i>sylvestris</i> (Vill.) Brenan	Ama	cas	ar		H	TM
<i>Amaranthus graecizans</i> subsp. <i>thellungianus</i> (Nevski) Gusev	Ama	cas	neo	1965	H	M
<i>Amaranthus hybridus</i> L.	Ama	cas	neo	1961	H	M
<i>Amaranthus hypochondriacus</i> L.	Ama	cas	neo	1853	H	TM
<i>Amaranthus ×ozanonii</i> Thell.	Ama	cas	neo	1943	H	M
<i>Amaranthus palmeri</i> S. Watson	Ama	cas	neo	1908	H	M
<i>Amaranthus powellii</i> S. Watson	Ama	inv	neo	1853	H	TM
<i>Amaranthus quitensis</i> Kunth	Ama	cas	neo	1910	H	N
<i>Amaranthus retroflexus</i> L.	Ama	inv	neo	1818	H	TM
<i>Amaranthus rudis</i> Sauer	Ama	cas	neo	1967	H	M
<i>Amaranthus spinosus</i> L.	Ama	cas	neo	1909	H	M
<i>Amaranthus ×turicensis</i> Thell.	Ama	cas	neo	1909	H	M
<i>Amaranthus viridis</i> L.	Ama	cas	neo	1964	H	M
<i>Ambrosia artemisiifolia</i> L.	Com	inv	neo	1883	H	M
<i>Ambrosia psilostachya</i> DC.	Com	cas	neo	1999	H	M
<i>Ambrosia trifida</i> L.	Com	cas	neo		H	M
<i>Amelanchier lamarcckii</i> Schroeder	Ros	cas	neo	1877	S	T
<i>Amelanchier ovalis</i> Med.	Ros	cas	neo		SH	TM
<i>Ammi majus</i> L.	Api	cas	neo	1898	H	TM
<i>Ammi visnaga</i> (L.) Lam.	Api	cas	neo	1987	H	TM
<i>Amorpha fruticosa</i> L.	Fab	inv	neo	1932	S	TM
<i>Anacyclus clavatus</i> (Desf.) Pers.	Com	cas	neo		H	T
<i>Anagallis arvensis</i> L.	Pri	nat*	arN		H	T
<i>Anagallis ×doerfleri</i> Ronniger	Pri	cas	ar		H	T
<i>Anagallis foemina</i> Miller	Pri	nat*	ar		H	T
<i>Anagallis monelli</i> L.	Pri	cas	neo	1953	H	TM
<i>Anaphalis margaritacea</i> (L.) Bentham	Com	cas	neo	1887	NSH	T
<i>Anchusa azurea</i> Mill.	Bor	cas	neo		H	TM
<i>Anchusa officinalis</i> L.	Bor	nat*	arR		SH	T
<i>Androsace elongata</i> L.	Pri	nat*	ar		NSH	T
<i>Androsace maxima</i> L.	Pri	nat*	ar		H	T
<i>Anethum graveolens</i> L.	Api	cas	ar		H	TM
<i>Angelica archangelica</i> L. subsp. <i>archangelica</i>	Api	inv	neo	1517	NSH	T
<i>Anoda cristata</i> (L.) Schlecht.	Mal	cas	neo	1973	H	M
<i>Anthemis arvensis</i> L.	Com	nat*	ar		H	T
<i>Anthemis austriaca</i> Jacq.	Com	nat*	ar		H	T
<i>Anthemis cotula</i> L.	Com	nat*	arM		H	T
<i>Anthoxanthum aristatum</i> Boiss.	Gra	cas	neo	1883	H	T
<i>Anthriscus caucalis</i> M. Bieb.	Api	nat*	arP		H	T
<i>Anthriscus cerefolium</i> (L.) Hoffm. subsp. <i>cerefolium</i>	Api	cas	neo	1834	SH	T
<i>Anthriscus cerefolium</i> subsp. <i>trichosperma</i> (Schult.) Arcang.	Api	nat*	ar		NSH	T
<i>Antirrhinum majus</i> L.	Scr	nat	neo	1819	H	T
<i>Apera spica-venti</i> (L.) P. B.	Gra	inv	ar		H	TM
<i>Apium graveolens</i> L.	Api	cas	ar		H	T
<i>Aquilegia atrata</i> Koch	Ran	cas	neo		SH	T
<i>Arabis alpina</i> L.	Bra	nat	neo		SH	T
<i>Arabis caucasica</i> Willd.	Bra	nat	neo	1957	SH	T
<i>Arabis procurrens</i> W. et K.	Bra	cas	neo		SH	TM
<i>Arctium ×ambiguum</i> (Čelak.) Beck	Com	cas	ar		H	TM
<i>Arctium ×cimbricum</i> (Krause) Hayek	Com	cas	ar		NS	T
<i>Arctium lappa</i> L.	Com	nat*	arB		H	TM
<i>Arctium ×maassii</i> (M. Schulye) Rouy	Com	cas	ar		NS	T
<i>Arctium minus</i> (Hill.) Bernh.	Com	nat	arM		H	TM
<i>Arctium ×mixtum</i> (Simk.) Nyman	Com	cas	ar		H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Si	r	3	d	AMC AMS	a	F
MP	r	1	a	AMS	pe	F
	e	1	a	E AS AF	a	F
VE PS	e	2	a	E	a	F
	e	1	a	E AS	a	F
Si VE	r	1	a	AMN AMC AMS	a	F, Grill & Priszter 1969
	r	2	d	AMC AMS	a	F
Si VE	sc	3	a		a	F
Si VE	r	3	a	AMN	a	F
Si VE PS Er	la	5	a	AMC AMS	a	F, Hejný et al. 1973, Jehlík 1998
Si	e	1	a	AMS	a	F
Si VE PS Er	c	5	a	AMN AMC	a	F
	r	2	a	AMN	a	F
	r	1	a	AMC AMS	a	F
	r	1	a		a	F
	r	2	a	AMS	a	F, Hejný et al. 1973, Jehlík 1998
PS Sr DM	la	5	a	AMN	a	K, Hejný et al. 1973
	s	1	a	AMN AMS	pe	K, Červinka & Sádlo 2000
Si	sc	3	a	AMN AMC	a	K, Hejný et al. 1973, Jehlík 1998
	r	1	d	AMN	s t	K, Čelakovský 1881
	r	3	d	E AS	s	F
	r	2	ad	E AS	a	F
	e	1	a	E AS	a	F
	la	3	d	AMN	s	F
	e	1	d	E	a	K
Cl Sh Si VE SO	c	5	a	E	a	F
Cl	r	4	a		a	F
Cl	sc	5	a	E AS	a	F
	e	1	a	E	a	F
GQ SS CE Ae	r	3	d	AMS AS	pe	K
	r	2	a	E AS	pe	F
Oa CA DM	sc	5	a	E	b pe	F
Ab AS Cl	r	4	a	E	a	F
Cl	e	2	a	E AS AF	a	F
Si	sc	5	d	E AS	a	F
Sf Pe Ae	la	5	d	E AS	b pe	F, Jehlík & Rostaříski 1975
	r	2	a	AMN AMC AMS	a pe	F
Ah Sh Cl	c	5	a	E	a	K
Sh Cl	sc	5	a	E	a	K
Mn	sc	5	a	E	a	K
	r	1	a	E	a	K
GA Si	r	4	a	E AS AF	a	F
GA	r	4	d	E AS	a	F
GA BS BR	la	4	ad	E AS	a	F
Cm	r	3	d	E	a pe	F
PS Ah Sn	c	5	a	E AS	a	K
Si	r	2	d	E AS AF	b	F
	r	2	d	E	pe	F
Cm	r	2	d	E AF	pe	F
Cm	r	2	d	E	pe	F
Cy AS	r	2	d	E	pe	F
Al	sc	4	a		pe	K
At IS Ae	r	4	a		pe	K
Al	c	5	a	E	pe	K
At IS Ae	sc	5	a		pe	K
Al	c	5	a	E	pe	K
Al	sc	5	a		pe	K

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Arctium xneumannii</i> Rouy	Com	cas	ar		NS	T
<i>Arctium xnothum</i> (Ruhmer) Weiss	Com	cas	ar		H	TM
<i>Arctium tomentosum</i> Mill.	Com	nat*	arB		H	TM
<i>Arctotheca calendula</i> (L.) Levyns	Com	cas	neo		H	M
<i>Argemone mexicana</i> L.	Pap	cas	neo	1965	H	M
<i>Armeria maritima</i> (Mill.) Willd.	Plu	cas#	neo	1890	SH	T
<i>Armoracia rusticana</i> G., M. et Sch.	Bra	nat	ar		NSH	TM
<i>Arnoseris minima</i> (L.) Schweigg. et Koerte	Com	nat	ar		SH	T
<i>Arrhenatherum elatius</i> subsp. <i>bulbosum</i> (Willd.) Schübl. et Mart.	Gra	cas	neo	1867	SH	TM
<i>Arrhenatherum elatius</i> (L.) J. Presl et C. Presl subsp. <i>elatius</i>	Gra	inv	neo		NSH	TM
<i>Artemisia abrotanum</i> L.	Com	cas	ar		H	TM
<i>Artemisia absinthium</i> L.	Com	nat*	ar		NSH	T
<i>Artemisia alba</i> Turra	Com	cas	neo		N	T
<i>Artemisia annua</i> L.	Com	nat	neo	1897	H	M
<i>Artemisia biennis</i> Willd.	Com	cas	neo		H	M
<i>Artemisia dracunculus</i> L.	Com	cas	neo		H	T
<i>Artemisia gnaphalodes</i> Nutt.	Com	cas	neo	1971	H	TM
<i>Artemisia repens</i> Willd.	Com	cas	neo	1872	H	M
<i>Artemisia scoparia</i> W. et K.	Com	nat*	ar		SH	T
<i>Artemisia sieversiana</i> Willd.	Com	cas	neo		H	M
<i>Artemisia tournefortiana</i> Rechb.	Com	nat	neo	1972	SH	T
<i>Artemisia verlotiorum</i> Lamotte	Com	nat	neo	1947	H	M
<i>Asclepias syriaca</i> L.	Asc	nat	neo	1901	H	M
<i>Asperugo procumbens</i> L.	Bor	nat*	ar		H	T
<i>Asperula arvensis</i> L.	Rub	nat*	arN		H	T
<i>Asperula orientalis</i> Boiss. et Hohen.	Rub	cas	neo	1905	H	T
<i>Aster bellidiastrum</i> (L.) Scop.	Com	cas	neo		H	TM
<i>Aster cordifolius</i> L.	Com	cas	neo	1867	H	TM
<i>Aster divaricatus</i> L.	Com	cas	neo		H	TM
<i>Aster dumosus</i> L. × <i>A. novi-belgii</i> L.	Com	cas	neo		H	TM
<i>Aster laevis</i> L.	Com	cas	neo	1851	SH	TM
<i>Aster lanceolatus</i> Willd.	Com	inv	neo		SH	TM
<i>Aster macrophyllus</i> L.	Com	cas	neo		H	TM
<i>Aster novae-angliae</i> L.	Com	cas	neo		SH	TM
<i>Aster novi-belgii</i> L.	Com	inv	neo	1850	SH	TM
<i>Aster parviflorus</i> Nees	Com	nat	neo	1872	SH	TM
<i>Aster xsalignus</i> Willd.	Com	inv	neo	1872	SH	TM
<i>Aster versicolor</i> Willd.	Com	inv	neo		SH	TM
<i>Astilbe xarendsii</i> Arends	Sax	cas	neo	1999	N	T
<i>Astragalus alopecuroides</i> L.	Fab	cas	neo	1872	S	T
<i>Astragalus glycyphylloides</i> DC.	Fab	cas	neo		S	T
<i>Astrodaucus orientalis</i> (L.) Drude	Api	nat	neo	1847	H	T
<i>Atriplex heterosperma</i> Bunge	Chen	cas	neo	1967	H	M
<i>Atriplex hortensis</i> L.	Chen	cas	neo	1872	H	TM
<i>Atriplex littoralis</i> L.	Chen	cas	neo	1977	H	M
<i>Atriplex xnorthusiana</i> Wein.	Chen	cas	ar		H	TM
<i>Atriplex oblongifolia</i> W. et K.	Chen	inv	arM		H	TM
<i>Atriplex patula</i> L.	Chen	nat*	arP		H	TM
<i>Atriplex rosea</i> L.	Chen	nat*	ar		H	TM
<i>Atriplex sagittata</i> Borkh.	Chen	inv	arP		H	TM
<i>Atriplex semilunaris</i> Aellen	Chen	cas	neo	1963	H	M
<i>Atriplex tatarica</i> L.	Chen	nat*	ar		H	TM
<i>Aubrieta deltooides</i> (L.) DC.	Bra	cas#	neo		H	T
<i>Avena barbata</i> Pott et Link	Gra	cas	neo		H	TM
<i>Avena fatua</i> L.	Gra	nat*	arB		H	TM
<i>Avena nuda</i> L.	Gra	cas	neo	1867	H	T
<i>Avena sativa</i> L. group <i>Chinensis</i>	Gra	cas	neo		H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
At IS Ae	sc	4	a		pe	K
Al	sc	5	a		pe	K
Al CA Ar	c	5	a	E	pe	K
	r	1	a	AF	pe	K
	e	1	a	AMC	a	F
	e	1	d	E	pe	F
Ph Ae Ap Sf Pa	c	5	d	E	pe	F
Sn Co	r	4	a	E	pe	K
	r	1	a	E	pe	K
Ar CA DM	c	5	a	E	pe	K
	r	1	d		s	K
Oa.Sc Al AF	sc	5	a	E AS	pe	K
	e	1	d	E	pe	K
	r	2	d	AS	a	K, Hejný et al. 1973, Jehlík 1998
	r	1	a	AMN	pe	K, Jehlík 1980
	r	2	d	AS	pe	K
	se	1	a	AMN	pe	K, Grill 1974
	r	1	a	E AS	pe	K
Ono	r	3	a	E	a	K
Si	r	3	a	E AMN	a	K, Hejný 1964, Hejný et al. 1973
	sc	1	a	AS	pe	K, Grill 1972
Si Al	r	3	d	AS	pe	K, Gutte & Pyšek 1972, Jehlík 1998
DM CA	r	3	d	AMN	pe	F
Si Al GA Oa VE	r	5	a	E AS AF	a	F
Cl	e	3	a	E AS AF	a	F
	e	2	ad	E AS	a	F
	r	1	d	E	pe	K
Ae Al Ar	r	1	d	AMN	pe	K
Ae Al Ar	r	1	d	AMN	pe	K, Pyšek & Vobořil 2002
Ae Al Ar	r	2	d		pe	K
Ae Al	r	3	d	AMN	pe	K
Ar Ae Sf	c	5	d	AMN	pe	K
Ae Al Ar	r	1	d	AMN	pe	K
	r	2	d	AMN	pe	K
Ae Al	sc	4	d	AMN	pe	K
Ae Al	sc	3	d	E	pe	K
Ae Al Ar	sc	3	d		pe	K
Ae Al Ar	r	3	d		pe	K
Ai	s	1	d	AS	pe	F
	e	1	a	E	pe	F
	e	1	a	E	pe	F
	e	1	a	E AS	b	F
	r	2	a	E AS	a	F
Si VE	r	4	d	E	a	F
	se	1	a	E AS AMN	a	F
	r	1	a		a	F
Si DM Al CA	la	5	a	E AS AF	a	F
Si Bi Oa	c	5	a	E AS	a	F
Si	r	4	a	E AS	a	F
Si	c	5	a	E AS	a	F
	e	1	a	AU	a	F
Si MP Oa	la	5	a	E AS AF	a	F
	r	1	d	E	pe	F
	se	1	a	E AS	a	Dostál 1989
Cl Sh Ah Si	c	5	a	E	a	K
	r	2	a		a	K
	r	2	a	E	a	K

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Avena sativa</i> L. group Praegravis	Gra	cas	neo		H	TM
<i>Avena sativa</i> L. group Sativa	Gra	nat	arB		H	TM
<i>Avena sterilis</i> L.	Gra	cas	neo		H	M
<i>Avena strigosa</i> Schreber	Gra	cas	ar		H	TM
<i>Axyris amaranthoides</i> L.	Chen	cas	neo	1953	H	M
<i>Azolla caroliniana</i> Willd.	Azo	cas	neo	1895	N	T
<i>Ballota nigra</i> subsp. <i>meridionalis</i> (Béguinot) Béguinot	Lam	cas	neo	1932	H	TM
<i>Ballota nigra</i> L. subsp. <i>nigra</i>	Lam	inv	arB		SH	TM
<i>Balsamita major</i> Desf.	Com	cas	neo		H	T
<i>Basella rubra</i> L.	Bas	cas	neo	1901	H	M
<i>Bassia sedoides</i> (Pallas) Aschers.	Chen	cas	neo	1960	H	M
<i>Bassia tricuspidis</i> F. Mueller	Chen	cas	neo	1966	H	M
<i>Beckmannia eruciformis</i> (L.) Host subsp. <i>eruciformis</i>	Gra	cas	neo		S	T
<i>Beckmannia syzigachne</i> (Steud.) Fernald	Gra	cas	neo		SH	M
<i>Bergenia crassifolia</i> (L.) Fritsch	Sax	cas	neo		H	TM
<i>Berteroa incana</i> (L.) DC.	Bra	nat*	ar		SH	T
<i>Berteroa stricta</i> Boiss. et Heldr.	Bra	cas	neo	1960	H	M
<i>Beta trigyna</i> W. et K.	Chen	cas	neo	1935	H	M
<i>Beta vulgaris</i> L. group Cicola	Chen	cas	ar		H	TH
<i>Beta vulgaris</i> L. group Vulgaris	Chen	cas	arM		H	TM
<i>Bidens connata</i> Willd.	Com	cas	neo		N	T
<i>Bidens frondosa</i> L.	Com	inv	neo	1894	NSH	TM
<i>Bidens pilosa</i> L.	Com	cas	neo	1981	H	M
<i>Bifora radians</i> M. Bieb.	Api	nat*	arM		H	T
<i>Bistorta amplexicaulis</i> (D. Don) Greene	Poly	cas	neo	1966	H	M
<i>Bolboschoenus glaucus</i> (Lam.) S. G. Smith	Cyp	cas	neo	1925	H	TM
<i>Borago officinalis</i> L.	Bor	cas	neo	1809	H	TM
<i>Brachypodium rupestre</i> (Host) R. et Sch.	Gra	cas	neo	1891	S	T
<i>Brassica elongata</i> Ehrh. subsp. <i>elongata</i>	Bra	cas	neo	1873	SH	TM
<i>Brassica elongata</i> subsp. <i>integrifolia</i> (L.) Koch	Bra	cas	neo	1960	H	M
<i>Brassica juncea</i> (L.) Czern. et Cosson	Bra	cas	neo	1963	H	M
<i>Brassica napus</i> L. subsp. <i>napus</i>	Bra	cas	ar		H	TM
<i>Brassica nigra</i> (L.) Koch	Bra	inv	ar		SH	T
<i>Brassica oleracea</i> L.	Bra	cas	ar		H	TM
<i>Brassica rapa</i> subsp. <i>oleifera</i> (DC.) Metzger	Bra	cas	ar		H	TM
<i>Brassica rapa</i> var. <i>sylvestris</i> (Lam.) Briggs	Bra	cas	neo	1964	H	TM
<i>Briza maxima</i> L.	Gra	cas	neo		H	TM
<i>Briza minor</i> L.	Gra	cas	neo		H	TM
<i>Bromus arvensis</i> L.	Gra	nat	arB		H	T
<i>Bromus briziformis</i> Fisch. et Mey.	Gra	cas	neo		H	M
<i>Bromus carinatus</i> Hooker et Arnott	Gra	cas	neo	1934	H	M
<i>Bromus catharticus</i> Vahl	Gra	cas	neo	1873	H	M
<i>Bromus commutatus</i> Schrad.	Gra	nat*	ar		H	TM
<i>Bromus hordeaceus</i> L. subsp. <i>hordeaceus</i>	Gra	nat	ar		H	TM
<i>Bromus hordeaceus</i> subsp. <i>pseudohominii</i> (P. Smith) H. Scholz	Gra	cas	neo	1971	H	M
<i>Bromus japonicus</i> Thunb.	Gra	nat	ar		H	T
<i>Bromus lanceolatus</i> Roth	Gra	cas	neo		H	M
<i>Bromus lepidus</i> Holmberg	Gra	cas	neo		H	TM
<i>Bromus madritensis</i> L.	Gra	cas	neo	1961	H	M
<i>Bromus pumpellianus</i> Scribner × <i>B. inermis</i> Leysser	Gra	cas	neo	1997	N	T
<i>Bromus rigidus</i> Roth	Gra	cas	neo		H	M
<i>Bromus riparius</i> Rehmman	Gra	cas	neo		H	M
<i>Bromus rubens</i> L.	Gra	cas	neo	1961	H	M
<i>Bromus scoparius</i> L.	Gra	cas	neo		H	M
<i>Bromus secalinus</i> subsp. <i>decipiens</i> Bomble et H. Scholz	Gra	cas	ar		H	T
<i>Bromus secalinus</i> subsp. <i>multiflorus</i> (Sm.) Schübl. et Mart.	Gra	cas	ar		H	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	r	2	a	E	a	K
Si Ah	c	5	d	E	a	K
	r	2	a	E	a	K
Ah Sh Cl	r	2	a	E	a	K
	se	1	a	AS	a	F
Le	r	1	d	AMN	af	F
Al	r	2	a	E	pe	F
Al BR BS Bd PR	c	5	a	E AS AF	pe	F
	r	2	d	AS	pe	K
	e	1	d	AS	pe	F
	e	1	a	E AS	a	F
	e	1	a	AU	a	F, Dvořák & Kühn 1966
	r	1	a	E AS	pe	K, Vicherek et al. 2000
	r	1	a	AMS AS	a	K
	r	3	d	AS	pe	F
Oa DM Ab PF	c	5	a	E AS	a b pe	F
	e	1	a	E	a b pe	F
	r	1	d	E AS	pe	F
	r	2	d		b a	F
Si VE	r	4	d		b a	F
	r	2	a	AMN	a	K, Lhotská 1968a
Bi	c	5	a	AMN	a	K, Hejny 1948, Hejny & Lhotská 1964, Lhotská 1966, 1968a, Jehlík et al. 1973, Gruberová et al. 2001
	r	1	a	AMN AMC AMS	a	K, Lhotská 1968a
Cl	r	4	a	E AS AF	a	F
	r	1	d	AS	pe	F
	s	1	a	E AS	pe	K, Hroudová et al. 1999
Oa Si	r	3	d	E AF	a	F
	se	1	a	E	pe	K, Schippman 1991
Br Si	r	2	a	E	b pe	F
	e	1	a	E AS	b pe	F
Si	r	2	ad	AS	a	F
MP Si	sc	5	d	E	a	F
Sf Pa	la	3	ad	E	a	F
Si	r	4	a	E	a b pe	F
Si	r	4	d	E	a	F
	r	2	a	E	a b	F, Kühn 1968
	r	1	d	E	a	K
	r	2	a	E	a	K
Sh Cl Si	r	4	a	E AS	a	K
	r	1	ad	E AS	a	K
	r	1	d	AMN	a pe	K, Svobodová & Řehořek 1996, Řehořek 2002
	r	1	a	AMS	a	K, Řehořek 2002
Cl Si	r	3	a	E	a	K
Si DM	c	5	a	E	a	K
	r	1	a	E	a	K
Cl DM	sc	5	a	E	a	K
	se	1	a	E	a	K, Dvořák & Kühn 1966
	r	1	a	E	a	K
	r	1	a	E	a	K, Dvořák & Kühn 1966
Ct	s	1	d		pe	K, Krahulec & Jiříštil 1997
	r	1	a	E	a	K
	r	1	a	E	pe	K
	se	1	a	E	a	K, Dvořák & Kühn 1966
	r	1	a	E AS	a	K, Dvořák & Kühn 1966
	e	1	a	E AS	a	K
Sh Ah	e	1	a	E AS	a	K

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Bromus secalinus</i> L. subsp. <i>secalinus</i>	Gra	nat*	arB		H	T
<i>Bromus sterilis</i> L.	Gra	nat*	arN		SH	TM
<i>Bromus tectorum</i> L.	Gra	nat*	arN		NSH	TM
<i>Brunnera macrophylla</i> (Adams) I. M. Johnston	Bor	cas	neo	1965	SH	T
<i>Bryonia alba</i> L.	Cuc	inv	ar		SH	TM
<i>Bryonia dioica</i> Jacq	Cuc	nat	ar		H	TM
<i>Buddleja davidii</i> Franchet	Bud	cas	neo	2000	H	M
<i>Bunias erucago</i> L.	Bra	cas	neo		H	M
<i>Bunias orientalis</i> L.	Bra	inv	neo	1856	SH	TM
<i>Bunium bulbocastanum</i> L.	Api	cas	neo	1879	H	T
<i>Bupleurum croceum</i> Fenzl.	Api	cas	neo	1943	H	T
<i>Bupleurum rotundifolium</i> L.	Api	nat*	ar		H	T
<i>Cakile baltica</i> (Jord. ex Rouy et Fouc.) Pobed.	Bra	cas	neo	1929	H	TM
<i>Cakile euxina</i> Pobed.	Bra	cas	neo	1960	H	M
<i>Calamintha grandiflora</i> (L.) Moench	Lam	cas	neo	1945	H	TM
<i>Calamintha menthaefolia</i> Host	Lam	cas	neo	1989	H	M
<i>Calamintha nepeta</i> subsp. <i>glandulosa</i> (Req.) P. W. Ball	Lam	cas	neo	1948	SH	T
<i>Calamintha nepeta</i> (L.) Savi subsp. <i>nepeta</i>	Lam	cas	neo	1996	N	T
<i>Calandrinia compressa</i> DC.	Por	cas	neo	1853	H	M
<i>Calendula arvensis</i> L.	Com	cas	neo	1901	H	TM
<i>Calendula officinalis</i> L.	Com	cas	neo	1872	H	TM
<i>Callistephus chinensis</i> (L.) Nees	Com	cas	neo	1872	H	TM
<i>Calystegia pulchra</i> Brummitt et Heywood	Con	nat	neo	1857	SH	T
<i>Camelina alyssum</i> (Mill.) Thell. subsp. <i>alyssum</i>	Bra	nat	ar		H	T
<i>Camelina alyssum</i> subsp. <i>integerrima</i> (Čelak.) Smejkal	Bra	nat	ar		H	T
<i>Camelina laxa</i> C. A. Meyer	Bra	cas	neo	1958	H	M
<i>Camelina microcarpa</i> DC. subsp. <i>microcarpa</i>	Bra	cas	neo		H	TM
<i>Camelina microcarpa</i> subsp. <i>sylvestris</i> (Wallr.) Hiitonen	Bra	nat*	ar		SH	T
<i>Camelina rumelica</i> Velen.	Bra	cas	neo	1963	H	M
<i>Camelina sativa</i> (L.) Crantz subsp. <i>sativa</i>	Bra	cas	neo	1852	H	T
<i>Camelina sativa</i> subsp. <i>zingeri</i> (Mirek) Smejkal	Bra	cas	neo		H	T
<i>Campanula alliariifolia</i> Willd.	Cam	cas	neo		NS	TM
<i>Campanula xiserana</i> Kovanda	Cam	cas	neo	1974	S	T
<i>Campanula medium</i> L.	Cam	cas	neo	1968	H	TM
<i>Campanula rapunculoides</i> L.	Cam	cas	neo		S	TM
<i>Campanula rhomboidalis</i> L.	Cam	nat	neo	1880	S	T
<i>Campanula speciosa</i> Hornem.	Cam	cas#	neo		SH	T
<i>Cannabis xintersita</i> Soják	Can	cas	neo	1960	H	TM
<i>Cannabis ruderalis</i> Janisch.	Can	inv	neo	1868	H	M
<i>Cannabis sativa</i> L.	Can	cas	ar		H	TM
<i>Capsella bursa-pastoris</i> (L.) Med.	Bra	nat*	arN		H	TM
<i>Cardamine chelidonia</i> L.	Bra	nat	neo	1930	NSH	T
<i>Cardamine hirsuta</i> L.	Bra	nat	ar		SH	TM
<i>Cardaria draba</i> (L.) Desv.	Bra	inv	arM		H	TM
<i>Carduus acanthoides</i> L.	Com	nat*	ar		H	TM
<i>Carduus crispus</i> L.	Com	nat*	arN		NSH	T
<i>Carduus xleptocephalus</i> Peterm.	Com	cas	ar		H	TM
<i>Carduus xorthocephalus</i> Wallr.	Com	cas	ar		H	T
<i>Carduus xsepincola</i> Hausskn.	Com	cas	ar		S	T
<i>Carduus xstangii</i> Buek	Com	cas	ar		H	T
<i>Carduus tenuiflorus</i> Curtis	Com	cas	neo	1967	H	M
<i>Carex muskingumensis</i> Schwein.	Cyp	cas	neo	1947	S	M
<i>Carthamus lanatus</i> L.	Com	cas	neo		H	TM
<i>Carthamus tinctorius</i> L.	Com	cas	neo		H	TM
<i>Castanea sativa</i> Mill.	Fag	cas	neo		NS	T
<i>Catalpa bignonioides</i> Walter	Big	cas	neo		H	M
<i>Catananche caerulea</i> L.	Com	cas	neo		H	M

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Ah	r	4	a	E AS	a	K
Si BR BS GA	c	5	a	E AS	a	K
Si Co DM Sc	c	5	a	E AS	a	K
	r	2	d	E AS	pe	F, Holub 1970
GA Al BS CR	c	5	d	E AS	pe	F
GA Al Ae BS	sc	3	d	E AS AF	pe	F
SS	r	1	d	AS	s	F
Al CA	r	1	ad	E AS	b pe	F
CA Al Ae Ar	la	4	a	E AS	b pe	F, Jehlík & Slavík 1968, Hejný et al. 1973, Jehlík 1998
	e	2	ad	E	pe	F
	se	1	a	E AS	a	Hadínek 2002
Cl	r	3	a	E AS	a	F, Šourková 1976
	e	1	a	E	a	F
	e	1	a	E	a	F
	s	1	d	E	pe	F
	se	1	a	E	pe	F
	s	1	d	E	pe	F
	s	1	d	E	pe	F
	r	1	d	AMS	a	F, Sekera 1854
	r	1	a	E	a	K
Si	sc	4	d		a	K
Si	r	3	d	AS	a	K
Ah Ae Al BS	r	4	d	AS	pe	F, Holub 1971
Ah	e	4	a	E AS	a	F
Ah	e	3	a	E AS	a	F
	e	1	a	E AS	a	F
Cl Si DM	sc	3	a	E AS	a	F
Cl Si DM Oa Fv	c	5	a	E AS	a	F
	r	2	a	E AS	b	F
Cl Si	e	3	d	E AS	a	F
Cl Si	e	2	a	E AS	a	F
	r	1	d	E	pe	F, Šuk 2001
	s	1	a		pe	F, Kovanda 1999
	r	2	d	E	b	F
	r	1	ad	E AS AF	b	F
	r	1	ad	E	pe	F, Kovanda & Husová 1976, Kovanda 1996
	r	2	d	E	pe	F
	e	1	a		a	F
Oa Si Al	la	4	a	AS	a	F, Jehlík 1998
Si Al	r	4	d	AS	a	F
MP Ab AS Si Ab	c	5	a	E	a b	F
Cr GA IS Ae	r	2	d	E	a pe	F, Kučera 1991
CE	r	3	a	E AS	a b	F
CA Si DM Al Oa	c	5	a	E AS	pe	F
DM Oa Al	la	5	a	E	pe	K
Sf Ae St Ph	sc	5	a	E AS	pe	K
	r	1	a		pe	K
Oa DM	r	2	a		pe	K
	r	2	a		pe	K
	r	2	a		pe	K
	se	1	a	E	a pe	
	se	1	a	AMN	pe	Jedlička 1949, Grill 1952
	s	1	a	E	a	K
	r	1	d	E AS	a	K
Cr GQ	r	3	d	E AS	t	F
	r	2	d	AMN	t	F
	s	1	d	E	pe	K

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Caucalis platycarpus</i> subsp. <i>muricata</i> (Čelak.) Holub	Api	nat*	ar		H	T
<i>Caucalis platycarpus</i> L. subsp. <i>platycarpus</i>	Api	nat*	arM		SH	T
<i>Celastrus orbiculatus</i> Thunb.	Cel	cas	neo		H	M
<i>Celosia argentea</i> var. <i>cristata</i> (L.) O. Kuntze	Ama	cas	neo	1902	H	M
<i>Celtis occidentalis</i> L.	Ulm	cas	neo	2001	H	M
<i>Cenchrus echinatus</i> L.	Gra	cas	neo		H	M
<i>Centaurea calcitrapa</i> L.	Com	cas	neo	1872	SH	T
<i>Centaurea cyanus</i> L.	Com	nat*	arB		H	T
<i>Centaurea dealbata</i> Willd.	Com	cas	neo		NH	TM
<i>Centaurea diffusa</i> Lam.	Com	cas	neo		SH	T
<i>Centaurea ×gerstlaui</i> Erdner	Com	cas	neo		H	M
<i>Centaurea macrocephala</i> Willd.	Com	cas	neo		H	TM
<i>Centaurea melitensis</i> L.	Com	cas	neo		H	TM
<i>Centaurea nigra</i> L.	Com	cas	neo	1872	SH	T
<i>Centaurea nigra</i> L. × <i>C. phrygia</i> L.	Com	cas	neo	1966	H	M
<i>Centaurea nigrescens</i> Willd. subsp. <i>nigrescens</i>	Com	cas	neo	1823	H	TM
<i>Centaurea ×psammogena</i> (Gáyer) Holub	Com	cas	neo		SH	T
<i>Centaurea solstitialis</i> L.	Com	cas	neo	1823	NSH	TM
<i>Centranthus ruber</i> (L.) DC.	Val	cas	neo	1880	SH	TM
<i>Cephalaria gigantea</i> (Ledeb.) Bobrov	Dip	cas	neo	1951	SH	TM
<i>Cephalaria syriaca</i> (L.) R. et Sch.	Dip	cas	neo	1948	H	T
<i>Cerastium biebersteini</i> DC.	Car	cas	neo		H	M
<i>Cerastium ×maureri</i> M. Schulze	Car	cas	neo		S	TM
<i>Cerastium tomentosum</i> L.	Car	cas	neo		SH	T
<i>Cerithe minor</i> L.	Bor	nat	ar		SH	T
<i>Chaenomeles japonica</i> (Thunb.) Spach	Ros	cas#	neo	1986	S	T
<i>Chamaecyparis lawsoniana</i> (A. Murray) Parl.	Cup	cas#	neo		N	T
<i>Chamaecytisus elongatus</i> (W. et K.) Link	Fab	cas#	neo		S	T
<i>Chelidonium majus</i> L.	Pap	nat*	arM		NSH	TM
<i>Chenopodium acuminatum</i> Willd.	Chen	cas	neo	1953	H	M
<i>Chenopodium ambrosioides</i> L.	Chen	cas	neo	1835	H	M
<i>Chenopodium berlandieri</i> subsp. <i>zschackei</i> (J. Murr) Zobel	Chen	cas	neo		H	M
<i>Chenopodium bonus-henricus</i> L.	Chen	nat*	arN		H	T
<i>Chenopodium botrys</i> L.	Chen	nat	ar		SH	TM
<i>Chenopodium capitatum</i> (L.) Aschers.	Chen	cas	neo	1809	H	TM
<i>Chenopodium ficifolium</i> Sm.	Chen	inv	arN		SH	TM
<i>Chenopodium foliosum</i> (Moench) Aschers.	Chen	cas	neo	1834	H	T
<i>Chenopodium glaucum</i> L.	Chen	nat*	arM		NSH	TM
<i>Chenopodium hircinum</i> Schrad.	Chen	cas	neo	1957	H	M
<i>Chenopodium integrifolium</i> Worosch.	Chen	cas	neo	1840	H	M
<i>Chenopodium melanocarpum</i> (J. Black) J. Black	Chen	cas	neo		H	M
<i>Chenopodium missouriense</i> Aellen	Chen	cas	neo	1963	H	M
<i>Chenopodium murale</i> L.	Chen	nat*	arN		H	TM
<i>Chenopodium nitriaceum</i> (F. Mueller) Bentham	Chen	cas	neo	1963	H	M
<i>Chenopodium opulifolium</i> Schrader	Chen	nat*	ar		H	TM
<i>Chenopodium pedunculare</i> Bertol.	Chen	inv	ar		SH	TM
<i>Chenopodium polyspermum</i> L.	Chen	nat*	arN		SH	TM
<i>Chenopodium probstii</i> Aellen	Chen	cas	neo		H	M
<i>Chenopodium prostratum</i> Herder	Chen	cas	neo		H	M
<i>Chenopodium pumilio</i> R. Br.	Chen	nat	neo	1890	H	M
<i>Chenopodium quinoa</i> Willd.	Chen	cas	neo	1966	H	M
<i>Chenopodium schradertianum</i> Schult.	Chen	cas	neo	1864	H	M
<i>Chenopodium striatiforme</i> J. Murr	Chen	nat	neo		H	M
<i>Chenopodium strictum</i> Roth	Chen	nat*	neo		H	TM
<i>Chenopodium urbicum</i> L.	Chen	nat*	arN		H	T
<i>Chenopodium vulvaria</i> L.	Chen	nat*	arM		H	T
<i>Chloris radiata</i> (L.) Swartz	Gra	cas	neo	1961	H	M

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Cl	r	2	a	E AS	a	F
Cl	r	5	a	E AS	a	F
BS	s	1	d	AS	s	F, Červinka & Sádlo 2000
Si	r	1	d	AMC AS	a	F
BS	r	1	d	AMN	t	K
	r	1	a	AMN	a	K
	r	2	a	E	a	K
Ap Sh	sc	5	a	E	a	K
Ae Ar	r	3	d	E AS	pe	K
DM Fv	r	2	a	E AS	a	K
	r	1	a		pe	
	r	1	d	E AS	pe	K
	e	1	a	E	a	K
	r	2	a	E	pe	K
	r	1	a		pe	
	r	2	a	E	pe	K
DM	s	1	a		?	Dostál 1989
Oa Fv	r	2	a	E AS	a	K
Cm	r	1	d	E	pe	F
Ar Ae	r	3	d	E AS	pe	F, Smejkal 1952
	se	1	a	E AS	a	F
	sc	2	d	E	pe	F
Cm Ar	sc	4	ad	E	pe	
Ar Cm	sc	4	d	E	pe	F
Oa GA	sc	5	a	E AS	b a pe	F
	s	1	d	AS	s	F
	r	1	d	AMN	t	F
	r	1	d	E	s	F
GA Ae SS TA	c	5	ad	E AS	pe	F
Si	e	1	a	AS	a	F
Si	r	3	d	AMS	a b	F
Si	r	1	a	AMN	a	F
Ae Al Ra Pa	c	5	ad	E	pe	F
Si PS Sr	sc	3	d	E AS	a	F
Si MP VE	r	2	d	AMN	a	F
Si Cb Bi	la	5	a	E AS	a	F, Dostálek 1983
Si MP VE	r	3	d	E AS AF	a b	F
Cb SJ Si MP Pa	sc	5	a	E AS	a	F, Dostálek 1983
Si	r	1	a	AMS	a	F
Si	r	1	d		a	F
Si	e	1	a	AU	a	F
Si	r	1	a	AMN	a	F, Hejny et al. 1973
Mn Si	sc	4	a	E AS	a	F
Si	e	1	a	AU	s	F
Si Al	sc	4	a	E AS AF	a	F
Cb Si VE	c	5	ad	E	a	F
SO Bi VE	c	5	a	E AS	a	F
Si	r	3	a	AMN	a	F
Si	e	1	a	AS	a	F
Mn MP Si PS Bi	sc	4	a	AU	a	F, Lhotská & Hejny 1979, Jehlík 1998
Si	e	1	d	AMS	a	F
Si	r	2	d	AF	a	F
Si MP Er PS	r	3	a	E AS	a	F
Si Mn Er PS	sc	5	a	AS	a	F, Dostálek 1983
Si	sc	4	a	E AS	a	F
Mn MP Si	sc	5	a	E AS	a	F, Dostálek 1983
	se	1	a	AMC AMS	a	K, Dvořák & Kühn 1966

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Chloris truncata</i> R. Br.	Gra	cas	neo	1956	H	M
<i>Chloris virgata</i> Swartz	Gra	cas	neo	1961	H	M
<i>Chlorocrepis staticifolia</i> (All.) Griseb.	Com	cas	neo	1873	H	T
<i>Chorispora tenella</i> (Pallas) DC.	Bra	cas	neo	1960	H	M
<i>Cicer arietinum</i> L.	Fab	cas	neo		H	TM
<i>Cicerbita macrophylla</i> subsp. <i>uralensis</i> (Rouy) P. D. Sell	Com	cas	neo		SH	TM
<i>Cichorium intybus</i> subsp. <i>foliosum</i> (Hegi) Janchen	Com	cas	neo		H	TM
<i>Cichorium intybus</i> L. subsp. <i>intybus</i>	Com	nat	arM		SH	TM
<i>Cirsium arvense</i> (L.) Scop.	Com	inv	arP		SH	TM
<i>Cirsium xaschersonianum</i> Čelak.	Com	cas	neo		SH	T
<i>Cirsium x bipontinum</i> F. W. Schultz	Com	cas	ar		SH	T
<i>Cirsium x celakovskyanum</i> Knaf	Com	cas	ar		SH	T
<i>Cirsium echinus</i> (M. Bieb.) Hand.-Mazz.	Com	cas	neo	1937	H	T
<i>Cirsium x gerhardtii</i> Schultz-Bip.	Com	cas	ar		SH	T
<i>Cirsium x preiseri</i> Uechtr.	Com	cas	ar		SH	T
<i>Cirsium x reichenbachianum</i> Löhrl	Com	cas	ar		SH	T
<i>Cirsium x sabaudum</i> Löhrl	Com	cas	ar		SH	T
<i>Cirsium x sextinum</i> Ausserd. ex Huter	Com	cas	ar		SH	T
<i>Cirsium x soroksarensis</i> Wagner	Com	cas	ar		SH	T
<i>Cirsium x subspinuligerum</i> Peterm.	Com	cas	ar		SH	T
<i>Cirsium tuberosum</i> (L.) All.	Com	cas	neo	1872	S	T
<i>Cirsium vulgare</i> (Savi) Ten.	Com	inv	arM		SH	TM
<i>Citrullus lanatus</i> (Thunberg) Matsumura et Nakai	Cuc	cas	neo	1969	H	TM
<i>Clarkia pulchella</i> Pursh.	Ona	cas	neo		H	M
<i>Clarkia unguiculata</i> Lindl.	Ona	cas	neo		H	M
<i>Claytonia alsinoides</i> Sims	Por	nat	neo	1951	H	TM
<i>Claytonia perfoliata</i> Willd.	Por	cas	neo		H	N
<i>Clematis flammula</i> L.	Ran	cas	neo		H	TM
<i>Clematis tangutica</i> (Maxim.) Korshinsky	Ran	cas	neo		H	T
<i>Clematis viticella</i> L.	Ran	cas	neo		H	TM
<i>Cnicus benedictus</i> L.	Com	cas	neo	1883	H	TM
<i>Cnidium silaifolium</i> (Jacq.) Simk.	Api	nat	neo	1868	H	TM
<i>Cochlearia officinalis</i> L.	Bra	cas	neo	1819	H	T
<i>Coleostephus myconis</i> (L.) Reichenb. fil.	Com	cas	neo		H	TM
<i>Collomia grandiflora</i> Lindl.	Pole	nat	neo	1880	SH	T
<i>Colutea arborescens</i> L.	Fab	nat	neo	1819	NS	T
<i>Commelina communis</i> L.	Come	cas	neo	1940	H	TM
<i>Conium maculatum</i> L.	Api	inv	arM		H	TM
<i>Conringia orientalis</i> (L.) Dumort.	Bra	nat*	arB		H	T
<i>Consolida ajacis</i> (L.) Schur	Ran	cas	neo	1880	H	TM
<i>Consolida orientalis</i> (Gay) Schrödinger	Ran	nat	neo	1913	H	TM
<i>Consolida regalis</i> S. F. Gray subsp. <i>regalis</i>	Ran	nat*	ar		H	T
<i>Convolvulus arvensis</i> L.	Con	nat*	arN		NSH	TM
<i>Convolvulus tricolor</i> L.	Con	cas	neo		H	TM
<i>x Conygeron huelsenii</i> (Vatke) Rauschert	Com	cas	neo	1887	H	T
<i>Conyza bonariensis</i> (L.) Cronq.	Com	cas	neo	1964	H	M
<i>Conyza canadensis</i> (L.) Cronq.	Com	inv	neo	1750	H	TM
<i>Conyza triloba</i> Decne.	Com	cas	neo	1971	H	T
<i>Coreopsis tinctoria</i> Nutt.	Com	cas	neo	1883	H	TM
<i>Coriandrum sativum</i> L.	Api	cas	neo	1819	H	TM
<i>Corispermum leptopterum</i> (Aschers.) Iljin	Chen	cas	neo	1960	H	M
<i>Cornus sericea</i> L. emend. Murray	Cor	nat	neo	1900	NSH	TM
<i>Coronilla scorpioides</i> (L.) Koch	Fab	cas	neo		H	M
<i>Coronopus didymus</i> (L.) Sm.	Bra	cas	neo	1903	H	M
<i>Coronopus squamatus</i> (Forskål) Aschers. subsp. <i>squamatus</i>	Bra	nat*	ar		H	T
<i>Corydalis alba</i> (Mill.) Mansf. subsp. <i>alba</i>	Fum	cas	neo	1995	H	TM
<i>Corydalis lutea</i> (L.) DC.	Fum	nat	neo	1886	H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	e	1	a	AS AU	a	K, Dvořák & Kühn 1966
	se	1	a	AMC AMS	a	K, Dvořák & Kühn 1966
	se	1	a	E	pe	Dostál 1989, Štech in prep.
	r	2	a	E AS	a	F
	r	1	d	E AS	a	F
Ae	r	1	d	AS	pe	K
	r	2	d		b	K
Cy DM Ar	c	5	a	E	pe	K
Si DM CA VE	c	5	a	E AS	pe	K
	se	1	a		pe	K
	r	2	a		pe	K
	r	3	a		pe	K
	e	1	a	AS	pe	K
	r	2	a		pe	K
	r	2	a		pe	K
	r	1	a		pe	K
	r	2	a		pe	K
	r	2	a		pe	K
	r	2	a		pe	K
	r	2	a		pe	K
	r	2	a		pe	K
	r	2	a		pe	K
	se	1	a	E	pe	K
DM Si CE	c	5	a	E AS	pe	K
Si VE	r	3	d	AF	a	F
	r	1	d	AMN	a	F
	r	1	d	AMN	a	F
	r	1	d	AMN	a	F, Holub 1975
	r	1	d	AMN AMC	a	F
	r	1	d	E AS AF	s	F
	r	1	d	AS	s	K, Pilát 1953, Procházka 1998
	r	2	d	E AS	s	F
	r	1	d	E	a	K
	e	1	a	E AS	pe	F
	r	1	d	E	b pe	F
	se	1	d	E	a	Dostál 1989
	r	3	d	AMN	a	F
Qp Bd	r	3	d	E	s	F
Si	sc	4	d	AS	a	K, Hejný et al. 1973, Jehlík 1998
Al Sf	sc	5	a	E AS AF	a b	F
Cl	r	4	a	E AS AF	a	F
Si Ah Sh	sc	3	ad	E AS AF	a	F
Cl Sh Si	r	4	ad	E AS AF	a	F, Jehlík 1998
Cl Sh	sc	5	a	E AS	a	F
CA DM Oa Cl Sh VE Br	c	5	a	E AS	pe	F
	r	2	d	E AS	a pe	F
	r	1	a		?	K, Čelakovský 1888b
	s	1	a	AMS	a	K
Si Sr PS	c	5	a	AMN	a	K
	se	1	a	AS AF	a pe	K
	r	2	d	AMN	a	K
	r	2	d	E	a	F
	e	1	a	E AS	a	F
Ai St	r	3	d	AMN	s	F
	e	1	a	E AS AF	a	F
MP Si	r	3	a	AMS	a b	F
MP	r	5	a	E	a b	F
Cm	r	1	d	E	pe	Dostál 1989
Cm	sc	4	d	E	pe	F

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Corylus colurna</i> L.	Bet	cas	neo	2001	SH	T
<i>Corylus maxima</i> Mill.	Bet	cas	neo	1902	H	M
<i>Cosmos bipinnatus</i> Cav.	Com	cas	neo		H	M
<i>Cotinus coggygia</i> Scop.	Ana	cas#	neo	1884	H	T
<i>Cotoneaster bullatus</i> Boiss.	Ros	cas	neo	2001	H	M
<i>Cotoneaster horizontalis</i> Decne	Ros	cas#	neo	1986	S	T
<i>Cotoneaster lucidus</i> Schlecht.	Ros	cas	neo		S	T
<i>Cotula australis</i> (Sieb. ex Spreng.) Hook fil.	Com	cas	neo	1961	H	M
<i>Crambe abyssinica</i> Hochst. ex R. E. Fries	Bra	cas	neo	1965	H	TM
<i>Crambe maritima</i> L.	Bra	cas	neo		H	TM
<i>Crataegus crus-galli</i> L.	Ros	cas#	neo	1900	N	T
<i>Crataegus flabellata</i> (Bosc ex Spach) C. Koch	Ros	cas	neo	1993	H	M
<i>Crataegus mollis</i> (Torrey et A. Gray)	Ros	cas	neo		SH	TM
<i>Crataegus pedicellata</i> Sarg.	Ros	cas	neo		SH	T
<i>Crataegus persimilis</i> Sarg.	Ros	cas	neo		S	T
<i>Crepis biennis</i> L.	Com	nat*	ar		SH	TM
<i>Crepis capillaris</i> (L.) Wallr.	Com	nat	ar		SH	T
<i>Crepis foetida</i> L. subsp. <i>foetida</i>	Com	cas	neo	1872	S	T
<i>Crepis foetida</i> subsp. <i>rhoeadifolia</i> (M. Bieb.) Čelak.	Com	nat	ar		SH	TM
<i>Crepis nicaeensis</i> Balb.	Com	cas	neo	1872	H	TM
<i>Crepis setosa</i> Haller fil.	Com	nat	arR		SH	T
<i>Crepis tectorum</i> L.	Com	nat*	ar		SH	T
<i>Crepis vesicaria</i> subsp. <i>taraxacifolia</i> (Thuill.) Thell.	Com	cas	neo		H	TM
<i>Crocus chrysanthus</i> Herb.	Lil	cas#	neo	1960	NSH	TM
<i>Crocus flavus</i> West.	Lil	cas	neo	1999	SH	TM
<i>Crocus heuffelianus</i> Herb.	Lil	cas	neo		S	T
<i>Crocus napolitanus</i> Mord.	Lil	cas	neo		SH	TM
<i>Crocus sativus</i> L.	Lil	cas	neo		S	T
<i>Cucumis melo</i> L.	Cuc	cas	neo	1963	H	TM
<i>Cucumis sativus</i> L.	Cuc	cas	neo		H	TM
<i>Cucurbita maxima</i> Duchesne	Cuc	cas	neo		H	TM
<i>Cucurbita pepo</i> L.	Cuc	cas	neo	1969	H	TM
<i>Cuscuta campestris</i> Yuncker	Con	inv	neo	1883	H	TM
<i>Cuscuta epilinum</i> Boenn.	Con	nat	arM		H	T
<i>Cydonia oblonga</i> Mill.	Ros	cas	ar		SH	T
<i>Cymbalaria muralis</i> G., M. et Sch. subsp. <i>muralis</i>	Scr	nat*	ar		H	TM
<i>Cymbalaria pallida</i> (Ten.) Wettst.	Scr	cas	neo		SH	TM
<i>Cynodon dactylon</i> (L.) Pers.	Gra	nat*	ar		SH	TM
<i>Cynosurus echinatus</i> L.	Gra	cas	neo		H	M
<i>Cyperus eragrostis</i> Lam.	Cyp	cas	neo	1999	N	T
<i>Cyperus rotundus</i> L.	Cyp	cas	neo		H	TM
<i>Cypripedium reginae</i> Walt.	Orch	cas#	neo	1935	S	T
<i>Cystopteris bulbifera</i> (L.) Bernh.	Dry	nat#	neo		N	T
<i>Cytisus scoparius</i> (L.) Link subsp. <i>scoparius</i>	Fab	inv*	neo	1819	NSH	T
<i>Dactyloctenium aegyptium</i> (L.) P. B.	Gra	cas	neo		H	M
<i>Dahlia pinnata</i> Cav.	Com	cas	neo		H	TM
<i>Dasypyrum villosum</i> (L.) P. Candargy	Gra	cas	neo		H	M
<i>Datura ferox</i> L.	Sol	cas	neo	1987	H	TM
<i>Datura innoxia</i> Mill.	Sol	cas	neo	1934	H	TM
<i>Datura stramonium</i> L. var. <i>stramonium</i>	Sol	nat	neo	1809	H	TM
<i>Datura stramonium</i> var. <i>tatula</i> (L.) Torrey	Sol	nat	neo	1935	H	TM
<i>Daucus carota</i> subsp. <i>sativus</i> (Hoffm.) Schübl. et Mart.	Api	cas	neo		H	TM
<i>Descurainia sophia</i> (L.) Webb ex Prantl	Bra	nat	arI		H	TM
<i>Desmazeria rigida</i> (L.) Tutin	Gra	cas	neo		H	M
<i>Deutzia scabra</i> Thunb.	Phi	cas	neo	2001	H	M
<i>Dianthus barbatus</i> L. subsp. <i>barbatus</i>	Car	cas#	neo	1874	SH	TM
<i>Dianthus caryophyllus</i> L.	Car	cas	neo		H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	s	1	d	E AS	t	F
	r	2	d	E AS	s	F
Si	r	4	d	AMC	a	K
	r	1	d	E	s t	F
Ar	s	1	d	AS	s	F
	r	2	d	AS	s	F
	s	1	d	AS	s	
	se	1	a	AU	a	Dvořák & Kühn 1966
	e	1	d	AF	a	F, Smejkal 1989a
	r	1	d	E	pe	F
	se	1	d	AMN	ts	F
	r	1	d	AMN	s t	F
	r	1	d	AMN	ts	F
	sc	3	d	AMN	ts	F
	s	1	d	AMN	s t	F
Ar	c	5	a	E	b	K
Cy PT	la	5	a	E	a	K
	e	1	a	E AS	a	K
DM Sc	la	5	a	E AS	a	K
	r	2	b	E	pe	K
	r	2	a	E	a	K
Si DM	r	4	a	E AS	a	K
	se	1	a	E	pe	K
	r	1	d	E AS	pe	K, Šuk 2001
	r	1	d	E	pe	K
	r	2	d	E	pe	K
	r	1	d	E	pe	K
	se	1	d		pe	K
	r	1	d	AS AF	a	F
	r	2	d	AS	a	F
	r	1	d	AMS	a	F
	r	2	d	AMN AMC AF	a	F
Si VE Al	sc	3	a	AMN	a	F, Jehlík 1998
Ap	e	4	a	E AS	a	F
	r	2	d	AS	ts	F
Cm	sc	5	ad	E	pe	F
	r	2	d	E	pe	F
CA Sr Er	sc	5	a	AS AF	pe	K
	r	1	a	E	a	K
	s	1	a	AMN AMC AMS	pe	K, Peřík 2002
	e	1	a	E	pe	K
	e	1	d	AMN	pe	Dostál 1948-1950, Šuk 2001
	s	1	d	AMN	pe	Marek & Procházka 2002
CE Ge GQ	sc	5	d	E	s	F
	r	1	a	AS AF	a	K
Si	r	3	d	AMC	pe	K
	r	1	a	E AS AF	a	K
	e	1	a	AS	a	F
Si VE	se	1	d	AMN AMC AMS	pe	F
Si VE	sc	5	ad	AMN	a	F
Si VE	r	3	ad	AMN	a	F
	r	3	d	AS	b	F
Si Cl Oa	c	5	a	E AS	a	F
	r	1	a	E	a	Dostál 1989
	s	1	d	AS	s	F
	r	3	d	E	pe	F
	r	2	d	E	pe	F

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Dianthus chinensis</i> L.	Car	cas	neo		H	TM
<i>Dichanthium sericeum</i> (R. Br.) A. Camus	Gra	cas	neo	1961	H	M
<i>Diervilla lonicera</i> Mill.	Cap	cas#	neo		SH	T
<i>Digitalis lanata</i> Ehrh.	Scr	cas	neo	1881	H	T
<i>Digitalis lutea</i> L.	Scr	cas	neo	1872	SH	T
<i>Digitalis purpurea</i> L.	Scr	inv	neo	1790	NS	T
<i>Digitaria ischaemum</i> (Schreber) Mühlenb.	Gra	nat*	ar		H	TM
<i>Digitaria sanguinalis</i> subsp. <i>pectiniformis</i> Henrard	Gra	nat*	arM		H	TM
<i>Digitaria sanguinalis</i> (L.) Scop. subsp. <i>sanguinalis</i>	Gra	nat*	arM		H	TM
<i>Dinebra retroflexa</i> (Vahl) Panzer	Gra	cas	neo	1972	H	M
<i>Diplotaxis muralis</i> (L.) DC.	Bra	nat*	ar		H	T
<i>Diplotaxis tenuifolia</i> (L.) DC.	Bra	nat*	ar		SH	T
<i>Dipsacus sativus</i> (L.) Honck.	Dip	cas	neo	1901	H	T
<i>Doronicum columnae</i> Ten.	Com	nat#	neo		S	T
<i>Doronicum orientale</i> Hoffm.	Com	nat#	neo	1819	S	T
<i>Doronicum parlatianches</i> L.	Com	nat#	neo	1897	S	T
<i>Draba sibirica</i> (Pall.) Thell.	Bra	cas	neo	1963	H	M
<i>Dracocephalum moldavica</i> L.	Lam	cas	neo	1854	H	T
<i>Dracocephalum thymiflorum</i> L.	Lam	cas	neo	1958	H	TM
<i>Duchesnea indica</i> (Andrew) Focke	Ros	nat	neo	1960	H	TM
<i>Ecballium elaterium</i> (L.) A. Richard	Cuc	cas	neo	1880	H	M
<i>Echinochloa colonum</i> (L.) Link	Gra	cas	neo		H	M
<i>Echinochloa crus-galli</i> (L.) P. B.	Gra	nat	arN		NSH	TM
<i>Echinochloa frumentacea</i> Link	Gra	cas	neo		H	M
<i>Echinochloa muricata</i> (P. B.) Fernald	Gra	cas	neo		H	M
<i>Echinochloa oryzoides</i> (Ard.) Fritsch	Gra	cas	neo	1950	H	M
<i>Echinochloa utilis</i> Ohwi et Yabuno	Gra	cas	neo		H	M
<i>Echinocystis lobata</i> (Michx) Torrey et A. Gray	Cuc	inv	neo	1911	NSH	T
<i>Echinops exaltatus</i> Schrad.	Com	nat	neo		SH	TM
<i>Echinops sphaerocephalus</i> L.	Com	inv	neo	1871	SH	TM
<i>Echium plantagineum</i> L.	Bor	cas	neo	1960	H	M
<i>Echium vulgare</i> L.	Bor	nat*	arN		NSH	TM
<i>Ehrharta longiflora</i> Sw.	Gra	cas	neo	1963	H	M
<i>Eichhornia crassipes</i> (C. Martius) Solms-Laub.	Pont	cas	neo	2000	H	T
<i>Elaeagnus angustifolia</i> L.	Ela	cas	neo		H	TM
<i>Eleusine indica</i> (L.) Gaertn.	Gra	cas	neo	1963	H	M
<i>Ellisia nyctelea</i> L.	Hydp	cas	neo		H	TM
<i>Elodea canadensis</i> Michx.	Hydc	inv	neo	1879	NSH	TM
<i>Elodea nuttallii</i> (Planch.) St. John	Hydc	cas	neo	1988	N	TM
<i>Elsholtzia ciliata</i> Willd.	Lam	cas	neo	1853	H	TM
<i>Elymus canadensis</i> L.	Gra	cas	neo		H	M
<i>Epilobium ciliatum</i> Rafin.	Ona	inv	neo	1926	NSH	TM
<i>Epilobium dodonaei</i> Vill.	Ona	nat	neo	1794	NSH	TM
<i>Epilobium ×floridulum</i> Smejkal	Ona	cas	neo	1980	H	TM
<i>Epilobium ×fossicola</i> Smejkal	Ona	cas	neo		SH	T
<i>Epilobium ×iglaviense</i> Smejkal	Ona	cas	neo	1979	SH	TM
<i>Epilobium ×interjectum</i> Smejkal	Ona	cas	neo	1987	SH	TM
<i>Epilobium ×josefi-holubi</i> Krahulec	Ona	cas	neo	1997	SH	T
<i>Epilobium komarovianum</i> H. Lévêille	Ona	cas	neo	1964	H	TM
<i>Epilobium ×mentiense</i> Smejkal	Ona	cas	neo	1987	SH	TM
<i>Epilobium ×novae-civitatensis</i> Smejkal	Ona	cas	neo	1972	H	TM
<i>Epilobium ×nutantiflorum</i> Smejkal	Ona	cas	neo	1976	SH	TM
<i>Epilobium ×prochazkae</i> Krahulec	Ona	cas	neo	1997	SH	T
<i>Epilobium ×vicinum</i> Smejkal	Ona	cas	neo	1971	SH	T
<i>Epimedium alpinum</i> L.	Ber	cas	neo	1874	H	TM
<i>Eragrostis albensis</i> H. Scholz	Gra	cas	neo	1984	H	M
<i>Eragrostis cilianensis</i> (All.) F. T. Hubbard	Gra	cas	neo		H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	r	1	d	AS	a b	F
	se	1	a	AU	pe	K, Dvořák & Kühn 1966
	r	1	d	AMN	s	F
	r	2	ad	E	b pe	F
	r	2	ad	E	pe	F
CE LF	la	5	d	E	b pe	F, Domin 1948, Mackeová 1999
Er PS Si	sc	5	a	E	a	K
MP PS Er	r	4	a	E	a	K
Er PS MP	c	5	d	E	a	K
	r	1	a	AS AF	a	K, Dvořák & Kühn 1966
Si VE Mn Si	sc	5	ad	E AF	a b	F
CA DM	sc	4	a	E AS AF	pe	F
	r	3	ad	E	b	F
	r	1	d	E	pe	K, Štech in prep.
	r	1	d	E AS	pe	K, Štech in prep.
	s	1	d	E	pe	K
	r	1	d	AS	pe	F
	e	2	d	AS	a	F
	e	2	a	E AS	a	F, Hejný et al. 1973
Ar Pa	r	3	d	AS	pe	F, Smejkal 1975b
	r	1	d	E AS AF	pe	F
	e	1	a	E	a	K
PS Bi Ne Es Si SO	c	5	a	E AS	a	K
	e	1	a	AS	a	K
	r	1	a	AMN	a	K
	e	1	a	E	a	K, Hejný 1950-51, Dvořák & Kühn 1966, Hejný et al. 1973
	r	1	a	AS	a	K
Sf Al Ae	la	4	d	AMN	a	F, Slavík & Lhotská 1967, Sutorý 2000, Rydlo 2000
Ae	r	3	d	E	pe	K
DM Al CA	c	5	d	E AS	pe	K, Hendrych 1987
	se	1	a	E	b	F
DM Ab AS Si AF Fv HF	c	5	a	E AS	b pe	F, Klotz 1963
	se	1	a	AF	a	Dvořák & Kühn 1966
	r	1	d	AMS	pe	Rydlo 2001
	r	1	d	E AS	t	F
MP Er	r	1	a	AF	a	K, Dvořák & Kühn 1966, Jehlík 1998
	r	1	ad	AMN	a	F
Mp Bf Pp	c	5	a	AMN	pe	K, Pyšek & Mandák 1998b
	r	1	d	AMN	pe	K, Husák 1992
Si	r	3	d	AS	a	F, Cejp 1948
	e	1	a	AMN	pe	K
Ma Ar Pa CE Ae Al Bi Si	c	5	a	AMN AMC	pe	F, Holub 1966, Smejkal 1986
DM Sx Se	la	3	a	E	pe	F, Slavík 1986
	r	1	a		pe	F, Smejkal 1995
	r	2	a		pe	F, Smejkal 1995
	s	1	a		pe	F, Smejkal 1995
	r	2	a		pe	F, Smejkal 1995
	s	1	a		pe	K, Krahulec 1999
	r	1	ad	AU	pe	F, Řehořek 1974, Holub 1978a
	r	1	a		pe	F, Smejkal 1995
	r	1	a		pe	F, Smejkal 1974
	r	3	a		pe	F, Smejkal 1995
	r	1	a		pe	K, Krahulec 1999
	r	1	a		pe	F, Smejkal 1995
Ae GA	r	2	d	E	pe	F
	s	1	a		a	K
	r	1	a	AS AF AMC AMS	a	K, Dvořák & Kühn 1966

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Eragrostis gracilis</i> Schrader	Gra	cas	neo		H	M
<i>Eragrostis mexicana</i> (Lag.) Link	Gra	cas	neo	1966	H	M
<i>Eragrostis minor</i> Host	Gra	nat*	ar		H	TM
<i>Eragrostis multicaulis</i> Steud.	Gra	cas	neo	1961	H	M
<i>Eragrostis suaveolens</i> Becher.	Gra	cas	neo	1961	H	M
<i>Eragrostis tef</i> (Zuccagni) Trotter	Gra	cas	neo	1965	H	M
<i>Eranthis hyemalis</i> (L.) Salisb.	Ran	cas	neo		NSH	T
<i>Erechtites hieraciifolia</i> (L.) Rafin. ex DC.	Com	nat	neo		NSH	T
<i>Erigeron annuus</i> subsp. <i>septentrionalis</i> (Fern. et Wieg.) Wagenitz	Com	inv	neo		H	TM
<i>Erigeron annuus</i> (L.) Pers. subsp. <i>annuus</i>	Com	nat	neo	1884	H	TM
<i>Erigeron speciosus</i> (Lindl.) DC.	Com	cas	neo	1888	H	TM
<i>Erigeron strigosus</i> Willd.	Com	nat	neo		H	TM
<i>Eriochloa procera</i> (Retz.) C. E. Hubb.	Gra	cas	neo	1961	H	M
<i>Erodium botrys</i> (Cav.) Bertol.	Ger	cas	neo	1956	H	M
<i>Erodium cicutarium</i> (L.) L'Hér. subsp. <i>cutarium</i>	Ger	nat*	arB		NSH	T
<i>Erodium gruinum</i> (L.) L'Hér.	Ger	cas	neo	1897	H	TM
<i>Erodium moschatum</i> (L.) L'Hér.	Ger	cas	neo	1855	H	M
<i>Erodium neuradifolium</i> Delile	Ger	cas	neo	1986	H	M
<i>Eruca sativa</i> (L.) Mill.	Bra	cas	neo	1900	H	TM
<i>Erucastrum gallicum</i> (Willd.) O. E. Schulz	Bra	nat	neo	1867	H	T
<i>Erucastrum nasturtiifolium</i> (Poirot) O. E. Schulz	Bra	nat	neo	1870	SH	T
<i>Eryngium amethystinum</i> L.	Api	cas	neo	1966	H	T
<i>Eryngium giganteum</i> M. Bieb.	Api	cas	neo	1995	H	T
<i>Erysimum argillosum</i> (Greene) Rydberg	Bra	cas	neo	1942	S	T
<i>Erysimum cheiranthoides</i> L. subsp. <i>cheiranthoides</i>	Bra	nat*	ar		SH	TM
<i>Erysimum cheiri</i> (L.) Crantz	Bra	cas	neo	1819	H	T
<i>Erysimum repandum</i> L.	Bra	nat*	ar		SH	T
<i>Erythronium dens-canis</i> L.	Lil	nat	neo	1819	NS	T
<i>Eschscholzia californica</i> Cham.	Pap	cas	neo		H	TM
<i>Euclidium syriacum</i> (L.) R. Br.	Bra	nat*	ar		SH	T
<i>Euphorbia chamaesyce</i> L.	Eup	cas	neo		H	TM
<i>Euphorbia exigua</i> L.	Eup	nat*	arI		H	T
<i>Euphorbia falcata</i> L.	Eup	nat*	ar		H	T
<i>Euphorbia helioscopia</i> L.	Eup	nat*	arB		H	TM
<i>Euphorbia humifusa</i> Willd.	Eup	cas	neo		H	TM
<i>Euphorbia lagascae</i> Sprengel	Eup	cas	neo	1974	H	TM
<i>Euphorbia lathyris</i> L.	Eup	cas	neo	1872	H	TM
<i>Euphorbia maculata</i> L.	Eup	cas	neo		H	TM
<i>Euphorbia marginata</i> Pursh.	Eup	cas	neo		H	TM
<i>Euphorbia peplus</i> L.	Eup	nat*	arM		H	T
<i>Euphorbia taurinensis</i> All.	Eup	cas	neo	1930	H	TM
<i>Fagopyrum esculentum</i> Moench	Poly	cas	neo	1872	H	T
<i>Fagopyrum tataricum</i> (L.) Gaertn.	Poly	cas	neo	1880	H	T
<i>Fallopia aubertii</i> (L. Henry) Holub	Poly	nat	neo		H	M
<i>Fallopia convolvulus</i> (L.) Á. Löve	Poly	nat*	arN		NSH	TM
<i>Ficus carica</i> L.	Mor	cas	neo	1966	H	TM
<i>Filago gallica</i> L.	Com	cas	neo	1872	SH	T
<i>Filipendula kamschatica</i> (Pallas) Maxim.	Ros	cas#	neo	1940	SH	T
<i>Filipendula rubra</i> (Hill) Robinson	Ros	cas	neo		H	TM
<i>Foeniculum vulgare</i> Mill.	Api	cas	ar		H	M
<i>Forsythia suspensa</i> (Thunb.) Vahl	Ole	cas	neo		SH	M
<i>Fragaria x magna</i> Thuill.	Ros	cas	neo		H	TM
<i>Fraxinus ornus</i> L.	Ole	nat	neo	1950	NS	T
<i>Fraxinus pennsylvanica</i> Marshall	Ole	inv	neo		NSH	TM
<i>Fritillaria meleagris</i> L.	Lil	cas	neo	1819	SH	T
<i>Fumaria capreolata</i> L.	Fum	cas	neo		H	TM
<i>Fumaria officinalis</i> L. subsp. <i>officinalis</i>	Fum	nat*	arN		H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	e	1	a	AMS	a	K
	r	1	a	AMN	a	K
Er	sc	5	a	E AS	a	K
	e	1	a	AS	a	K, Dvořák & Kühn 1966
	se	1	a	E AS	a	Dvořák & Kühn 1966
	e	1	a	AF AS	a	K, Kubát 1979
Ae Ai	r	2	d	E	pe	F
CE At	r	4	a	AMN	pe	K, Vopravil 1950-1951
DM Al CA Ar	c	5	a	AMN	ab	K
Si DM	sc	4	a	AMN	a	K, Jehlík 1998
	se	1	d	AMN AMS	pe	Dostál 1989
	r	3	a	AMN	a	K
	se	1	a	AS	a	Dvořák & Kühn 1966
	e	1	a	E AS	ab	F, Slavík 1996b
AS Ab PF KP AF Cl DM Oa Si HF Fvc	5	a	a	E AS	ab	F
	e	1	ad	E AS	ab	F
	r	2	ad	E AS	ab	F
	e	1	a	E AS AF	ab	F, Slavík 1996c
	r	3	ad	E	a	F
Si PS SO CA	sc	4	a	a	ab	F, Štěpánek 1983
Oa DM CA	la	3	a	E	b pe	F, Štěpánek 1983
	se	1	d	E	pe	F
	se	1	d	E	pe	F
	e	1	d	AMN	b pe	F, Kirschner & Štěpánek 1984
VE SO Bi Si	c	5	a	E AS AF	a	F
	r	2	d	E	pe	F
Cl CA Oa	r	4	a	E	a	F
Cr	s	1	d	E	pe	K
	r	2	d	AMN	a	F
	r	2	a	E AS	a	F, Sutorý 1982
	e	1	a	AMN	a	F
Cl Sh	sc	5	a	E	a	F
Cl	sc	4	a	E AS AF	a	F
VE	c	5	a	E AS AF	a	F
MP	e	1	a	AS	a	F
	e	1	a	E	a	F, Unar 1978
Si VE	r	3	d	E	ab	F
	e	1	ad	AMN	a	F
Si	r	2	d	AMN	a	F
VE Si Cl	c	5	a	E	a	F
DM	r	2	a	E AS	a	F, Chrtek & Křísa 1970
	r	3	d	AS	a	F
	r	2	d	AS	a	F
BS	sc	5	d	AS	s	F
Cl Sh Ah Sn VE SO TA	c	5	ad	E AS	a	F
	r	2	d	AS	ts	F, Eitel 1982
Th Sn	e	2	a	E AS AF	a	K
	e	1	d	AS	pe	F, Smrček & Malina 1984
	e	1	d	AMN	pe	F
	r	2	d	E AS	pe	F
BS	r	1	d	AS	s	F
Si	sc	4	d		pe	F
	r	2	d	E AS	t	F
Ai	la	4	d	AMN	t	F
	r	1	d	E	pe	K
	r	1	a	E	a	F
Cl Sh Ah VE	sc	5	a	E AS	a	F

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Fumaria officinalis</i> subsp. <i>wirtgenii</i> (Koch) Arcang.	Fum	nat	arN		H	TM
<i>Fumaria parviflora</i> Lam.	Fum	cas	ar		H	T
<i>Fumaria rostellata</i> Knaf	Fum	nat*	ar		H	TM
<i>Fumaria schleicheri</i> Soyer-Willemet	Fum	nat*	ar		SH	TM
<i>Fumaria vaillantii</i> subsp. <i>schrammii</i> (Aschers.) Nyman	Fum	nat*	arM		SH	TM
<i>Fumaria vaillantii</i> Loisel. subsp. <i>vaillantii</i>	Fum	nat*	arI		SH	TM
<i>Gagea villosa</i> (M. Bieb.) Duby	Lil	nat*	ar		SH	T
<i>Gaillardia pulchella</i> Foug.	Com	cas	neo		H	M
<i>Galega officinalis</i> L.	Fab	nat	neo	1819	SH	TM
<i>Galeobdolon argentatum</i> Smejkal	Lam	inv	neo		NSH	TM
<i>Galeopsis ladanum</i> L.	Lam	nat*	arM		SH	T
<i>Galeopsis segetum</i> Necker	Lam	cas	neo	1852	SH	TM
<i>Galinsoga ciliata</i> (Rafin.) Blake	Com	inv	neo	1901	H	TM
<i>Galinsoga parviflora</i> Cav.	Com	inv	neo	1867	H	TM
<i>Galium parisiense</i> L.	Rub	cas	neo	1835	SH	T
<i>Galium rubioides</i> L.	Rub	cas	neo	1852	S	T
<i>Galium spurium</i> L.	Rub	nat*	arN		SH	T
<i>Galium tricoratum</i> Dandy	Rub	nat	arM		SH	T
<i>Galium verrucosum</i> Hudson	Rub	cas	neo	1822	H	T
<i>Gastridium ventricosum</i> (Gouan) Schinz et Thell.	Gra	cas	neo	1961	H	M
<i>Gaudinia fragilis</i> (L.) P. B.	Gra	cas	neo		H	M
<i>Genista sagittalis</i> L.	Fab	nat	neo	1928	NS	T
<i>Gentiana lutea</i> L. subsp. <i>lutea</i>	Gen	nat	neo		NS	T
<i>Geranium columbinum</i> L.	Ger	nat*	arB		NSH	T
<i>Geranium dissectum</i> L.	Ger	nat*	arN		H	T
<i>Geranium ibericum</i> Cav.	Ger	cas	neo	1965	H	TM
<i>Geranium macrorrhizum</i> L.	Ger	cas#	neo		SH	TM
<i>Geranium molle</i> L.	Ger	nat	ar		H	T
<i>Geranium pusillum</i> Burm. fil.	Ger	nat	arB		H	TM
<i>Geranium pyrenaicum</i> Burm. fil.	Ger	inv	neo	1819	H	TM
<i>Geranium reflexum</i> L.	Ger	cas	neo	1992	SH	T
<i>Geranium rotundifolium</i> L.	Ger	cas	neo	1851	H	TM
<i>Geranium sibiricum</i> L.	Ger	nat	neo	1850	SH	TM
<i>Geranium versicolor</i> L.	Ger	cas	neo	1986	H	M
<i>Geum aleppicum</i> Jacq.	Ros	cas	neo	1923	H	TM
<i>Geum ×gajewskii</i> Smejkal	Ros	cas	neo	1956	S	T
<i>Geum macrophyllum</i> Willd.	Ros	cas	neo	1956	SH	TM
<i>Geum ×spurium</i> Fisch. et Mey.	Ros	cas	neo		H	TM
<i>Gilia capitata</i> Sims.	Pole	cas	neo	1982	S	T
<i>Gilia multicaulis</i> Bentham	Pole	cas	neo		H	TM
<i>Gilia tricolor</i> Bentham	Pole	cas	neo		H	TM
<i>Glaucium corniculatum</i> (L.) J. H. Rudolph	Pap	nat*	ar		NS	T
<i>Glaucium flavum</i> Crantz	Pap	cas	neo	1900	SH	T
<i>Glyceria striata</i> (Lamk.) A. S. Hitchc.	Gra	nat	neo		NSH	T
<i>Glyceria stricta</i> Hook	Gra	cas	neo	1961	H	M
<i>Glycine max</i> (L.) Merrill	Fab	cas	neo	1958	H	TM
<i>Glycyrrhiza glabra</i> L.	Fab	nat#	neo	1900	SH	T
<i>Grindelia squarrosa</i> (Pursh) Dunal	Com	cas#	neo		H	T
<i>Guizotia abyssinica</i> (L. fil.) Cass.	Com	cas	neo		H	TM
<i>Gypsophila elegans</i> M. Bieb.	Car	cas	neo	1968	H	TM
<i>Gypsophila scorzonifolia</i> Ser.	Car	cas	neo	1900	H	TM
<i>Helianthus annuus</i> L.	Com	cas	neo	1872	H	TM
<i>Helianthus ×laetiflorus</i> Pers.	Com	nat	neo		H	TM
<i>Helianthus petiolaris</i> Nutt.	Com	cas	neo	1974	H	M
<i>Helianthus rigidus</i> (Cass.) Desf.	Com	cas	neo		H	M
<i>Helianthus salicifolius</i> A. Dietr.	Com	cas	neo	1973	H	TM
<i>Helianthus strumosus</i> L.	Com	cas	neo		H	M

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Cl Sh Ah VE	c	5	a	E AS	a	F
VE	e	2	ad	E AS AF	a	F
VE	sc	5	a	E	a	F
Cl Sh VE Si GA PS	sc	5	a	E AS	a	F
Cl VE Si PS	r	3	a	E	a	F
Cl Sh VE Si PS	sc	5	a	E AS	a	F
Bd VE BR Ab	sc	4	a	E	pe	K
	r	1	d	AMN	a	K
Sf Pa Ar DM Oa	r	4	d	E	pe	F
Ai BS TA Ae	sc	4	d		pe	F, Smejkal 1975a
Sh Sc Cl CE	sc	4	a	E AS	a	F
	r	2	d	E	a	F
VE PS	c	5	a	AMC AMS	a	K
VE PS	c	5	a	AMS	a	K
	e	2	ad	E AS AF	a	F, Kaplan & Řehořek 1998
	e	1	a	E	pe	F
Cl Sh	sc	5	a	E AS	a	F
Cl	r	4	a	E AF	a	F
	e	1	ad	E AF	a	F
	se	1	a	E AF	a	K, Dvořák & Kühn 1966
	r	1	a	E	a	K
GQ Vc EC Ge	r	3	ad	E	ss	F, Skalická 1993
	r	1	d	E	pe	F
Bd Gs DM Sc AF	sc	5	a	E AS AF	a b	F, Slavík 1997a
VE Sh SO	sc	5	a	E AS	a	F
	r	1	d	AS	pe	F, Slavík 1997a
	r	2	d	E	pe	F, Slavík 1997b
VE Si	r	3	a	E AS AF	a b	F, Slavík 1997b
VE Si SO Cl Sh	c	5	a	E AS	a b	F, Slavík 1997b
Si GA MP	c	5	ad	E AS	b pe	F, Slavík 1997b
Ae	r	1	d	E	pe	F
	r	2	a	E AS	a	F, Slavík 1997a
	r	2	ad	E AS	pe	F, Slavík 1997a
	e	1	a	E	pe	F, Chrtěk 1989
	r	2	ad	AMN E AS	pe	F, Domin 1923, Smejkal 1988, 1989b
	e	1	a		pe	F, Smejkal 1959
	r	1	ad	AMN AS	pe	F
	e	1	a		pe	F
	s	1		AMN	pe	
	r	1	d	AMN	a	F
	r	1	d	AMN	a b	F
Fv Cl AS	r	3	a	E AS	a b	F
	e	2	d	E	b pe	F
	r	2	a	AMN	pe	K, Dančák 2002
	se	1	a	AU	pe	Dvořák & Kühn 1966
	r	1	d	AS	a	F
DM	r	1	d	E AS	pe	F
	r	2	d	AMN	a pe	K
	r	2	ad	AF	a	K, Smejkal 1989a
	r	1	ad	E AS	a	F
	r	2	d	AS	pe	F, Grill & Smejkal 1966
Si	sc	5	d	AMN	a	K, Jehlík 1998
Sf Ar Ae	sc	5	d	AMN	pe	K
	se	1	d	AMN	a	K
	r	1	d	AMN	pe	K
	se	1	d	AMN	pe	K
	se	1	d	AMN	pe	K

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Helianthus tuberosus</i> L.	Com	inv	neo	1885	NSH	TM
<i>Heliopsis helianthoides</i> (L.) Sweet	Com	cas	neo		H	TM
<i>Heliotropium europaeum</i> L.	Bor	nat*	arB		H	T
<i>Helleborus foetidus</i> L.	Ran	cas	neo		H	T
<i>Helleborus niger</i> L.	Ran	cas	neo	1874	SH	T
<i>Helleborus odorus</i> W. et K.	Ran	cas#	neo		N	T
<i>Helleborus viridis</i> L.	Ran	nat	neo	1819	SH	T
<i>Helminthotheca echioides</i> (L.) Holub	Com	cas	neo	1861	H	TM
<i>Hemerocallis fulva</i> (L.) L.	Hem	cas	neo	1883	SH	TM
<i>Hemerocallis lilioasphodelus</i> L.	Hem	cas	neo	1883	SH	TM
<i>Heracleum mantegazzianum</i> Sommier et Levier	Api	inv	neo	1862	NSH	TM
<i>Heracleum persicum</i> Desf. ex Fischer, Meyer et Lalem.	Api	cas	neo	1960	H	TM
<i>Herniaria cinerea</i> DC.	Car	cas	neo		H	M
<i>Herniaria hirsuta</i> L.	Car	nat*	ar		H	T
<i>Hesperis matronalis</i> subsp. <i>candida</i> (Kit. ex Schulzer, Kanitz et Knapp) Thell.	Bra	cas	neo	1909	SH	T
<i>Hesperis matronalis</i> L. subsp. <i>matronalis</i>	Bra	nat	neo	1817	NSH	T
<i>Hesperis matronalis</i> subsp. <i>oblongifolia</i> (Schur) Dvořák	Bra	cas	neo	1933	SH	T
<i>Hesperis matronalis</i> subsp. <i>oblongipetala</i> (Borbás) Dvořák	Bra	cas	neo	1909	SH	T
<i>Hesperis pycnotricha</i> Borbás et Degen	Bra	cas	neo	1950	H	M
<i>Hibiscus trionum</i> L.	Mal	nat	ar		H	TM
<i>Hieracium pannosum</i> Boissier	Com	cas#	neo	1978	S	T
<i>Hippocrepis emerus</i> (L.) Lassen	Fab	cas	neo	1891	S	T
<i>Hippophaë rhamnoides</i> L. subsp. <i>rhamnoides</i>	Ela	cas	neo	1902	H	M
<i>Hirschfeldia incana</i> (L.) Lagreze-Fossat	Bra	cas	neo	1956	H	M
<i>Hordeum distichon</i> L.	Gra	cas	ar		H	TM
<i>Hordeum geniculatum</i> All.	Gra	cas	neo	1961	H	M
<i>Hordeum jubatum</i> L.	Gra	nat	neo		H	M
<i>Hordeum leporinum</i> Link	Gra	cas	neo	1967	H	M
<i>Hordeum marinum</i> Huds.	Gra	cas	neo		H	M
<i>Hordeum murinum</i> L.	Gra	nat*	arI		H	TM
<i>Hordeum secalinum</i> Schreber	Gra	cas	neo		H	M
<i>Hordeum vulgare</i> L.	Gra	cas	ar		H	TM
<i>Hosta plantaginea</i> (Lamk.) Aschers.	Aga	cas#	neo		H	T
<i>Humulus scandens</i> (Lour.) Merrill	Can	cas	neo		H	TM
<i>Hyacinthella leucophaea</i> (C. Koch) Schur	Hya	cas#	neo	1960	N	T
<i>Hyacinthella rumelica</i> Velen.	Hya	cas#	neo	1900	NSH	T
<i>Hylotelephium anacampseros</i> (L.) Ohba	Cra	cas	neo		H	T
<i>Hylotelephium ewersii</i> (Ledeb.) Ohba	Cra	cas	neo		H	T
<i>Hylotelephium spectabile</i> (Boreau) Ohba	Cra	cas	neo		H	T
<i>Hyoscyamus albus</i> L.	Sol	cas	neo	1890	H	T
<i>Hyoscyamus niger</i> L.	Sol	nat*	arN		H	T
<i>Hyparrhenia hirta</i> (L.) Stapf	Gra	cas	neo	1961	H	M
<i>Hyssopus officinalis</i> L.	Lam	cas	neo	1819	SH	T
<i>Iberis amara</i> L.	Bra	cas	neo	1888	SH	TM
<i>Iberis sempervirens</i> L.	Bra	cas#	neo		H	TM
<i>Iberis umbellata</i> L.	Bra	cas	neo	1880	H	TM
<i>Impatiens balfouri</i> Hooker fil.	Bal	cas	neo		S	TM
<i>Impatiens balsamina</i> L.	Bal	cas	neo		H	TM
<i>Impatiens glandulifera</i> Royle	Bal	inv	neo	1896	NSH	TM
<i>Impatiens parviflora</i> DC.	Bal	inv	neo	1870	NS	TM
<i>Impatiens scabriflora</i> DC.	Bal	cas	neo	1986	S	T
<i>Imperatoria ostruthium</i> L.	Api	inv*	neo	1809	SH	T
<i>Imperatoria verticillaris</i> (L.) DC.	Api	cas	neo	1960	H	M
<i>Inula helenium</i> L.	Com	nat	neo	1819	SH	T
<i>Ipomoea hederacea</i> (L.) Jacq.	Con	cas	neo	1972	H	TM
<i>Ipomoea purpurea</i> (L.) Roth	Con	cas	neo	1969	H	TM
<i>Iris germanica</i> L.	Lil	nat	neo	1867	NSH	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Al Ae	c	5	d	AMN	pe	K
	r	1	d	AMN	pe	K
Er VE Oa	e	2	ad	E AS AF	a	F
	r	2	d	E AF	pe	F
	r	2	d	E	pe	F
	r	1	d	E	pe	Dostál 1948-1950, Šuk 2001
	sc	3	d	E	pe	F
DM	r	3	a	E	pe	K
BS BR Ae	r	4	d	E	pe	K
	r	1	d	E	pe	K
Ae BS SS Ct Ar	la	5	d	E	b pe	F, Pyšek 1991, 1994, Pyšek et al. 1995a, Pyšek & Pyšek 1994
	s	1	d	AS	b pe	F
	r	1	a	E AS AF	a	F
MP	r	3	a	E AS AF	a pe	F
	sc	2	d	E	pe	F, Dvořák 1968
Al IS BS Ae Pe	sc	5	d	E	pe	F, Dvořák 1968
	r	1	d	E	pe	F, Dvořák 1968
	r	2	ad	E	pe	F, Dvořák 1968
	s	1	d	E	b pe	F
	r	3	ad	E AS	a	F
Er Si	s	1	d	E AS	pe	
	e	1	d	E AS AF	s	F
	r	1	d	E AS	t s	F
	r	3	a	E AS AF	a b	F, Jehlík 1998
Si MP	r	1	d	E	a	K
	r	1	a	E AS	a	K, Dvořák & Kühn 1966
	sc	4	d	AMN	a	K
Si	se	1	a	E AS	a	
	r	1	a	E	a	K
	c	5	a	E AS	a	K
Pa Si	r	1	a	E	pe	K
	sc	5	d		a	K
Si	r	1	d	AS	pe	K
	r	1	d	AS	a	F
	s	1	d	E	pe	Šuk 2001
	r	1	d	E	pe	Dostál 1989, Šuk 2001
	r	1	d	E	pe	F
	r	1	d	AS	ss	F
	r	1	d	AS	pe	F
	e	1	a	E AS AF	b a pe	F
	sc	5	a	E AS AF	b a	F
	se	1	a	E AS	pe	K, Dvořák & Kühn 1966
Oa Si	sc	2	d	E	ss	F
	r	1	d	E	a	F
	r	2	d	E AS	ss	F
	r	2	d	E	a	F
	r	1	d	AS	a	F
	r	1	d	AS	a	F
	r	1	d	AS	a	F
	la	5	d	AS	a	F, Daumann 1967, Pyšek & Prach 1995a,b, Slavík 1996a
Cr Fg TA CR BS SS IS GA Ae	c	5	d	AS	a	F, Vraštil 1952, Daumann 1967, Slavík 1996a
	e	1	d	AS	a	F
PT Pe Ae Ad	la	5	d	E	pe	F, Kopecký 1973
	e	1	a	E	pe	F
Pa Ae	sc	4	d	AS	pe	K
	r	1	a	AMN AMC AMS	a	K
CA Fv	r	2	d	AMC AMS	a	F
	sc	4	d	E	pe	K

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Iris pallida</i> Lam.	Lil	cas	neo		SH	TM
<i>Iris sambucina</i> L.	Lil	cas	neo	1867	SH	TM
<i>Isatis tinctoria</i> subsp. <i>praecox</i> (Tratt.) Domin et Podp.	Bra	cas	neo	1921	S	T
<i>Isatis tinctoria</i> L. subsp. <i>tinctoria</i>	Bra	nat	ar		NS	T
<i>Ismelia versicolor</i> Cass.	Com	cas	neo		H	TM
<i>Iva xanthiifolia</i> Nutt.	Com	nat	neo		H	TM
<i>Juglans nigra</i> L.	Jug	cas	neo		S	T
<i>Juglans regia</i> L.	Jug	nat	arP		NSH	TM
<i>Juncus tenuis</i> Willd.	Jun	inv	neo	1851	SH	T
<i>Kickxia elatine</i> subsp. <i>crinita</i> (Mabille) Greuter	Scr	cas	neo	1934	H	T
<i>Kickxia elatine</i> (L.) Dumort. subsp. <i>elatine</i>	Scr	nat*	ar		H	T
<i>Kickxia spuria</i> (L.) Dumort. subsp. <i>spuria</i>	Scr	nat*	ar		H	T
<i>Kochia scoparia</i> subsp. <i>densiflora</i> (Moq.) Aellen	Chen	cas	neo	1901	H	M
<i>Kochia scoparia</i> (L.) Schrader subsp. <i>scoparia</i>	Chen	inv	neo	1819	H	M
<i>Kochia scoparia</i> subsp. <i>scoparia</i> f. <i>trichophylla</i> Schinz et Thell	Chen	cas	neo	1819	H	TM
<i>Laburnum anagyroides</i> Med.	Fab	nat	neo	1900	SH	T
<i>Lactuca sativa</i> L.	Com	cas	neo		H	TM
<i>Lactuca serriola</i> L.	Com	nat*	arM		H	TM
<i>Lactuca tatarica</i> (L.) C. A. Meyer	Com	cas	neo	1957	H	M
<i>Lactuca virosa</i> L.	Com	cas	neo	1872	H	T
<i>Lagurus ovatus</i> L.	Gra	cas	neo		H	M
<i>Lamium album</i> L.	Lam	nat	arB		SH	TM
<i>Lamium amplexicaule</i> L.	Lam	nat*	arI		SH	TM
<i>Lamium hybridum</i> Vill.	Lam	cas	neo	1946	H	TM
<i>Lamium moluccellifolium</i> Fries	Lam	cas	ar		H	TM
<i>Lamium orvala</i> L.	Lam	cas	neo		S	T
<i>Lamium purpureum</i> L.	Lam	nat*	arN		H	TM
<i>Lappula patula</i> (Lehm.) Menyh.	Bor	cas	neo	1960	H	M
<i>Lappula squarrosa</i> (Retz.) Dumort	Bor	nat*	ar		NSH	T
<i>Lapsana communis</i> L. subsp. <i>communis</i>	Com	nat*	arN		NSH	T
<i>Lathyrus annuus</i> L.	Fab	cas	neo		H	M
<i>Lathyrus aphaca</i> L.	Fab	nat*	ar		NSH	T
<i>Lathyrus articulatus</i> L.	Fab	cas	neo		H	TM
<i>Lathyrus cicera</i> L.	Fab	cas	neo		SH	TM
<i>Lathyrus clymenum</i> L.	Fab	cas	neo		H	M
<i>Lathyrus odoratus</i> L.	Fab	cas	neo		H	TM
<i>Lathyrus ochrus</i> (L.) DC.	Fab	cas	neo		H	M
<i>Lathyrus sativus</i> L.	Fab	cas	neo	1874	H	T
<i>Lathyrus tingitanus</i> L.	Fab	cas	neo		H	TM
<i>Lathyrus tuberosus</i> L.	Fab	nat*	ar		SH	T
<i>Lavandula angustifolia</i> Mill.	Lam	cas	neo		SH	TM
<i>Lavatera trimestris</i> L.	Mal	cas	neo		H	TM
<i>Lawrenca glomerata</i> Hooker	Mal	cas	neo	1961	H	M
<i>Legousia hybrida</i> (L.) Delarbre	Cam	cas	neo	1809	H	TM
<i>Legousia speculum-veneris</i> (L.) Chaix	Cam	cas	neo	1809	H	TM
<i>Lemna turionifera</i> Landolt	Lem	cas	neo	1997	S	TM
<i>Lens culinaris</i> Med.	Fab	cas	neo	1819	H	TM
<i>Leontopodium alpinum</i> Cass.	Com	cas#	neo	1901	NS	T
<i>Leonurus cardiaca</i> L.	Lam	nat*	arM		H	T
<i>Leonurus intermedius</i> Holub	Lam	nat	neo	1887	H	T
<i>Leonurus japonicus</i> Houtt.	Lam	cas	neo	1934	H	TM
<i>Leonurus villosus</i> Dum.-d'Urv.	Lam	nat	neo	1899	H	T
<i>Lepidium africanum</i> (Burm. fil.) DC.	Bra	cas	neo	1964	H	M
<i>Lepidium campestre</i> (L.) R. Br.	Bra	nat*	arR		SH	T
<i>Lepidium densiflorum</i> Schrad.	Bra	nat	neo	1904	NSH	TM
<i>Lepidium heterophyllum</i> Bentham	Bra	cas	neo		H	M
<i>Lepidium latifolium</i> L.	Bra	cas	neo	1900	H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	s	1	d	E	pe	K
	r	2	d	AS	pe	K
	s	1	a	E	b pe	F
DM Fv AF	la	4	d	E	b pe	F
	r	1	d	AF	a	K
Si	sc	4	a	AMN	a	K, Lhotská & Slavík 1969, Hejný et al. 1973, Jehlík 1998
	r	1	d	AMN	t	F
BS Cr Qp AQ BR Bd Ar Al	la	5	d	E AS	t	F
NJ Cy	c	5	a	AMN	pe	K
	se	1	a	E AS AF	a	F
Sh Cl	r	4	a	E	a	F
Cl	r	4	a	E AS AF	a	F
Si	sc	3	a	AS	a	F, Jehlík 1998
Si Sr	sc	5	a	E AS	a	F, Jehlík 1998
Si	sc	3	d	E AS	a	F
Bd	la	4	d	E	s t	F
Si	r	4	d	AS	a	K
Si	c	5	a	E AS	a	K
Si Al	r	3	a	E AS	pe	K, Hejný et al. 1973, Jehlík 1980, 1998
	e	1	a	E	a	K
	r	2	d	E	a	K
Al Ae	c	5	a	E AS	pe	F
Si Sh Cl VE Ab AS	sc	5	a	E AS AF	a	F, Košťál 1903
	r	1	a		?	F, Otruba 1946
	r	1	a	E	?	F
Ae Ai	r	1	d	E	pe	F
VE Sh Cl Si	c	5	a	E AS AF	a b	F
	e	1	a	E AS AF	a	F, Holub 1974
Oa Si	sc	5	a	E AS	a b	F
GA IS TA	c	5	a	E AS	a	K
	e	1	a	E AS	a	F
Fv Gs Ar CA Pa Cl	r	3	a	E AS AF	a	F, Chrtková et al. 1977
	e	1	a	E AF	a	F
	r	2	ad	E AS AF	a	F
	e	1	a	E AS AF	a	F
	r	1	d	E	a	F
	e	1	a	E AS AF	a	F
	r	2	d	E AS AF	a	F
	r	1	d	E AF	a	F
Cl CA DM	sc	5	a	E AS	pe	F
	r	1	d	E	ss	F
	r	2	d	E	a	F
	se	1	a	AU	pe	F, Dvořák & Kühn 1966
	r	1	d	E	a	F
	r	1	d	E	a	F
Le	s	1	a	AS AMN	a pe	K, Kaplan 2000
	r	3	d		a	F
	r	2	d	E AS	pe	K
Al Ae	sc	5	ad	E AS	pe	F, Holub 1993
Al Ae	sc	4	ad		pe	Holub 1993
	e	1	a	AS	pe	F
Al Ae	r	3	ad	E AS	pe	Holub 1993
	se	1	a	AF	a	F
Bd Ar, KP Ab DM PF	sc	5	a	E AS	a b	F
DM Co Sr	la	4	a	AMN	a b	F, Hejný et al. 1973
Al MP CA Si	r	2	a	E	pe	F
	r	2	ad	E AS	pe	F

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Lepidium perfoliatum</i> L.	Bra	cas	neo	1872	H	TM
<i>Lepidium ruderale</i> L.	Bra	nat*	arR		H	TM
<i>Lepidium sativum</i> L.	Bra	cas	neo	1874	H	TM
<i>Lepidium virginicum</i> L.	Bra	cas	neo	1936	H	TM
<i>Leptochloa chinensis</i> Nees	Gra	cas	neo		H	M
<i>Leptochloa fascicularis</i> (Lamk.) A. Gray	Gra	cas	neo		H	M
<i>Leptochloa filiformis</i> (Lamk.) P. B.	Gra	cas	neo	1961	H	M
<i>Lepyrodiclis holosteoides</i> (C. A. Meyer) Fisch. et Mey.	Car	cas	neo	1967	H	T
<i>Leucanthemella serotina</i> (L.) Tzvelev	Com	cas	neo		N	T
<i>Leucosinapis alba</i> (L.) Spach	Bra	nat	neo	1875	H	TM
<i>Leucosinapis dissecta</i> (Lag.) Zelený	Bra	cas	neo	1953	H	M
<i>Levisticum officinale</i> Koch	Api	cas	neo	1809	H	T
<i>Leymus arenarius</i> (L.) Hochst.	Gra	cas	neo		N	T
<i>Linaria arvensis</i> (L.) Desf.	Scr	nat*	ar		H	T
<i>Linaria maroccana</i> Hooker	Scr	cas	neo		H	TM
<i>Linaria repens</i> (L.) Mill.	Scr	cas	neo	1934	NSH	TM
<i>Linaria vulgaris</i> Mill.	Scr	nat*	ar		SH	TM
<i>Lindernia dubia</i> (L.) Pennell	Scr	cas	neo	1989	S	T
<i>Linum usitatissimum</i> L.	Lin	cas	ar		H	TM
<i>Lithospermum arvense</i> L. subsp. <i>arvense</i>	Bor	nat*	arN		SH	T
<i>Lithospermum arvense</i> subsp. <i>caerulescens</i> (DC.) Rothm.	Bor	cas	neo	1867	H	T
<i>Lobelia erinus</i> L.	Cam	cas	neo		H	TM
<i>Lobularia maritima</i> (L.) Desv.	Bra	cas	neo	1963	H	M
<i>Lolium loliaceum</i> (Bory et Chaub.) Hand. - Mazz.	Gra	cas	neo		H	M
<i>Lolium multiflorum</i> Lamk.	Gra	nat	neo	1883	H	TM
<i>Lolium remotum</i> Schrank	Gra	cas	arI		H	T
<i>Lolium rigidum</i> Gaudin	Gra	cas	neo		H	TM
<i>Lolium temulentum</i> L.	Gra	cas	arB		H	T
<i>Lonicera caprifolium</i> L.	Cap	nat	neo	1809	NSH	T
<i>Lonicera tatarica</i> L.	Cap	cas#	neo	1872	SH	TM
<i>Lunaria annua</i> L.	Bra	nat	neo	1819	NSH	TM
<i>Lupinus albus</i> L.	Fab	cas	neo	1878	H	M
<i>Lupinus angustifolius</i> L.	Fab	cas	neo	1900	SH	TM
<i>Lupinus luteus</i> L.	Fab	cas#	neo	1880	SH	T
<i>Lupinus polyphyllus</i> Lindl.	Fab	inv	neo	1895	NS	T
<i>Luzula nivea</i> (Nath.) DC.	Jun	nat	neo		SH	T
<i>Lychnis chalconica</i> L.	Car	cas	neo		H	TM
<i>Lychnis coronaria</i> (L.) Desr.	Car	nat	neo	1879	NS	T
<i>Lycium barbarum</i> L.	Sol	inv	neo	1870	NSH	TM
<i>Lycium chinense</i> Mill.	Sol	cas#	neo		H	T
<i>Lycopsis arvensis</i> L.	Bor	nat*	arB		H	T
<i>Lycopsis orientalis</i> L.	Bor	cas	neo	1911	H	TM
<i>Lycopus europaeus</i> subsp. <i>menthifolius</i> (Mabille) Skalický	Lam	cas	neo	1880	H	T
<i>Lysimachia punctata</i> L.	Pri	nat	neo	1819	SH	TM
<i>Lythrum junceum</i> Banks et Solander	Lyt	cas	neo	1965	H	M
<i>Macleaya cordata</i> (Willd.) R. Br.	Pap	cas	neo		H	TM
<i>Madia sativa</i> Molina	Com	cas	neo		H	M
<i>Mahonia aquifolium</i> (Pursh) Nutt.	Ber	inv	neo		NSH	TM
<i>Majorana hortensis</i> Moench	Lam	cas	neo		H	TM
<i>Malcolmia africana</i> (L.) R. Br.	Bra	cas	neo	1935	H	M
<i>Malcolmia chia</i> (L.) DC.	Bra	cas	neo		H	M
<i>Malcolmia maritima</i> (L.) R. Br.	Bra	cas	neo	1850	H	M
<i>Malope trifida</i> Cav.	Mal	cas	neo	1969	H	TM
<i>Malus ×dasyphylla</i> Borkh.	Ros	nat	ar		NS	T
<i>Malus domestica</i> Borkh.	Ros	cas	ar		NSH	TM
<i>Malva ×adulterina</i> Wallr.	Mal	cas	ar		H	T
<i>Malva crispa</i> (L.) L.	Mal	cas	neo	1853	H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Si DM	r	3	ad	E	a	F
MP	c	5	a	E	a b	F
	r	3	d	AS AF	a	F
DM Si MP	sc	3	ad	AMN AMC	a b	F, Hejný et al. 1973
	r	1	a	AS	a	K
	r	1	a	AMN AMS	a	K
	r	1	a	AMC AMS	a	K, Dvořák & Kühn 1966
	r	1	a	AS	a	F
	e	1	a	E AS	pe	K
Si VE	sc	3	ad	E AS	a	F
	e	1	a	E	a	F
Ae Ad	r	3	d	AS	pe	F
	r	1	d	E	pe	K
Ah SO	r	4	a	E AF	a	F, Suda 1999, 2001
	r	1	a	AF	a	F
Sc DM	r	2	ad	E	pe	F
DM CA	c	5	a	E AS	pe	F
	s	1	a	AMN	a	F, Kurka 1990
Si	sc	5	d	E AS	a b	F
Cl Sh VE AS	sc	5	a	E AS	a	F
Cl	se	1	a	E AS	a	F
	r	2	d	AF	a	F
	r	2	d	E	pe a	F
	s	1	a	E AS	a	K
Pa Ar	c	5	d		pe	K
Ah	e	3	a	E	a	K
	r	1	a	E AS	a	K, Dvořák & Kühn 1966
Ah	e	4	a	E	a	K
Cr Bd	sc	4	d	E	s	F
	r	2	d	AS	s	F
CR TA Ae GA Si	sc	4	d	E	b	F
	r	2	ad	E	a	F
	r	2	ad	E	a	F
	e	2	d	E	a	F
CE Ar	c	5	d	AMN	pe	F
	r	1	d	E	pe	K
	r	2	d	E AS	pe	F
Fv Bd Qt Ar	r	4	ad	E AS AF	pe b	F
BS BR	sc	5	d	E AS	s	F
	s	1	d	AS	s	F
SO PS	sc	5	a	E	a b	F
	r	1	a	E AS	a b	F, Krahulec 1981
	se	1	a	E	pe	F, Skalický 1968
Ae Ap Ar Ct PT CE	c	4	d	E	pe	F
	se	1	a	E	pe	F, Toman & Starý 1966
	r	1	d	AS	pe	F
	r	1	a	AMC	a	K
Bd Qt CR	la	5	d	AMN	s	F
	r	2	d	E	a b	F
	e	1	a	E AS AF	a	F, Krist 1940
	e	1	d	E	a	F
	r	1	d	E	a	F
	r	2	d	E AF	a	F
Bd Cr	sc	4	a		t	F
PR Bd BS SS	sc	5	d		t s	F
Mn MP	r	2	a		?	F
Si Al	r	4	d	AS	a	F

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Malva neglecta</i> Wallr.	Mal	nat*	arI		H	T
<i>Malva parviflora</i> L.	Mal	cas	neo	1957	H	TM
<i>Malva pusilla</i> Sm.	Mal	nat*	arN		H	T
<i>Malva sylvestris</i> L.	Mal	nat*	ar		H	T
<i>Malva verticillata</i> L.	Mal	cas	neo		H	TM
<i>Malva ×zoemigii</i> Fleischer	Mal	cas	ar		H	T
<i>Mantisalca salmantica</i> (L.) Briq.	Com	cas	neo		H	TM
<i>Marrubium ×paniculatum</i> Desr.	Lam	cas	ar		H	T
<i>Marrubium peregrinum</i> L.	Lam	nat*	ar		SH	T
<i>Marrubium vulgare</i> L.	Lam	nat*	arM		H	T
<i>Matricaria discoidea</i> DC.	Com	inv	neo	1851	H	TM
<i>Matteucia struthiopteris</i> (L.) Tod.	Dry	nat	neo	1820	NSH	T
<i>Matthiola incana</i> (L.) R. Br. subsp. <i>incana</i>	Bra	cas	neo	1877	H	TM
<i>Matthiola longipetala</i> subsp. <i>bicornis</i> (Sibth. et Sm.) P. W. Ball	Bra	cas	neo	1952	H	TM
<i>Matthiola longipetala</i> (Vent.) DC. subsp. <i>longipetala</i>	Bra	cas	neo	1924	H	M
<i>Medicago arabica</i> (L.) Hudson	Fab	cas	neo	1936	H	M
<i>Medicago disciformis</i> DC.	Fab	cas	neo	1963	H	M
<i>Medicago lupulina</i> L.	Fab	nat	ar		SH	TM
<i>Medicago orbicularis</i> (L.) Bartal.	Fab	cas	neo		H	M
<i>Medicago polymorpha</i> L.	Fab	cas	neo	1880	H	M
<i>Medicago rigidula</i> (L.) Desr.	Fab	cas	neo	1923	SH	T
<i>Medicago sativa</i> L. subsp. <i>sativa</i>	Fab	nat	neo	1819	SH	TM
<i>Medicago ×varia</i> Martyn	Fab	nat*	neo		NSH	T
<i>Melampyrum arvense</i> L.	Scr	nat*	ar		NS	T
<i>Melampyrum barbatum</i> Willd. subsp. <i>barbatum</i>	Scr	cas	neo	1893	SH	T
<i>Melica altissima</i> L.	Gra	nat	neo	1955	SH	T
<i>Melilotus albus</i> Med.	Fab	inv	ar		SH	TM
<i>Melilotus indicus</i> (L.) All.	Fab	cas	neo	1913	H	M
<i>Melilotus messanensis</i> (L.) All.	Fab	cas	neo	1929	H	M
<i>Melilotus officinalis</i> (L.) Pallas	Fab	inv	arM		SH	TM
<i>Melilotus sulcatus</i> Desf.	Fab	cas	neo	1929	H	M
<i>Melilotus wolgicus</i> Poir.	Fab	cas	neo	1963	H	M
<i>Melissa officinalis</i> L. subsp. <i>officinalis</i>	Lam	cas	neo	1872	H	TM
<i>Mentha arvensis</i> L.	Lam	nat*	ar		SH	TM
<i>Mentha ×dalmatica</i> Tausch	Lam	cas	ar		SH	TM
<i>Mentha ×gracilis</i> Sole	Lam	nat	neo	1855	H	TM
<i>Mentha ×niliaca</i> Jacq.	Lam	cas	neo	1976	SH	T
<i>Mentha ×piperita</i> L. nothosubsp. <i>piperita</i>	Lam	cas	neo	1840	H	TM
<i>Mentha ×rotundifolia</i> (L.) Huds.	Lam	nat*	neo	1846	SH	TM
<i>Mentha spicata</i> L. subsp. <i>spicata</i>	Lam	nat	neo	1818	SH	TM
<i>Mentha spicata</i> L. s.l.	Lam	nat	neo	1844	SH	TM
<i>Mentha ×verticillata</i> L.	Lam	nat	ar		NSH	T
<i>Mercurialis annua</i> L.	Eup	nat*	arB		H	TM
<i>Mespilus germanica</i> L.	Ros	cas	ar		NS	T
<i>Microrrhinum litorale</i> (Willd.) Speta	Scr	cas	neo	1994	H	M
<i>Microrrhinum minus</i> (L.) Fourr.	Scr	nat*	ar		NSH	T
<i>Mimulus guttatus</i> DC.	Scr	inv*	neo	1853	NS	T
<i>Mimulus moschatus</i> Lindl.	Scr	nat	neo	1868	NS	T
<i>Mirabilis jalapa</i> L.	Nyc	cas	neo		H	M
<i>Miscanthus sinensis</i> N. J. Andersson	Gra	cas	neo		SH	T
<i>Misopates orontium</i> (L.) Rafin.	Scr	nat*	ar		H	T
<i>Monolepis nuttalliana</i> (Schult.) Greene	Chen	cas	neo	1927	H	M
<i>Myagrum perfoliatum</i> L.	Bra	cas	neo	1855	H	TM
<i>Myosotis arvensis</i> (L.) Hill subsp. <i>arvensis</i>	Bor	nat*	arB		SH	TM
<i>Myosotis ×krajinae</i> Domin	Bor	cas	ar		N	T
<i>Myosotis ×pseudohispida</i> Domin	Bor	cas	ar		H	T
<i>Myrrhis odorata</i> (L.) Scop.	Api	inv*	neo	1809	NS	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Mn MP Oa	c	5	ad	AS	b pe	F
	e	1	a	E AS	a	F
Mn MP Oa	sc	5	ad	AS	a	F
Si Oa Al	sc	4	d	E AS	b pe	F
Si DM	r	3	d	AS	a b pe	F
	e	1	a		?	F
	r	1	a	E	pe	K
Oa CA Al	r	1	a		pe	F
Oa CA Al	r	4	a	E	pe	F
Oa Mn Al	r	4	ad	E	pe	F
MP	c	5	a	AS	a	K
Ai An IS Ae	r	3	d	E AS AMN	f	F, Hendrych 1984
	r	1	d	E	a b	F
	r	1	d	E AS	a	F
	e	1	a	E AS	a	F
	r	2	a	E AF	a	F
	e	1	a	E	a	F
MP Si CA Cl VE Cy Ar DM	c	5	ad	E AS AF	a pe	F
	e	1	a	E AS	a	F
	r	2	a	E	a	F
	e	1	a	E	a	F
DM CA Br Ar	c	5	d	AS	pe	F
Br CA	sc	5	ad	AS	pe	F
Gs Tm Br Ar KP	sc	5	a	E AS	ap	F
Cl Sh	e	1	a	E	ap	F
Fv Ar	r	1	d	E AS	pe	K, Pyšek 1997
DM Si Al CA	c	5	ad	E AS	b a	F
	r	2	a	E AS	a	F
	e	1	a	E	a	F
DM Si Al CA	c	5	ad	E AS	b	F
	e	1	a	E	a	F
	e	1	a	E	b	F
Ae BS	r	3	d	E AS	pe	F
Ah Sh VE SO Pa Ae	c	5	a	E AS	pe	F, Štěpánek 1998
Ae Pa	sc	4	ad		pe	F, Štěpánek 1998
Ae Pa	sc	4	d	E	pe	F, Štěpánek 1998
	r	2	d		pe	F, Štěpánek 1998
Pa	sc	3	d	E	pe	F, Štěpánek 1998
Ae Ar Pe Al	c	5	d		pe	F, Štěpánek 1998
Pa	r	3	d?	E	pe	F, Štěpánek 1998
Pa	r	3	d	E	pe	F, Štěpánek 1998
Ap Pa SG Pr Ph	c	5	a		pe	F, Štěpánek 1998
VE	c	5	a	E AF	a	F
GQ Bd	r	3	d	E AS	t s	F
DM	r	1	a	E	a	F, Mikoláš 1997
Sc Sr DM	sc	5	a	E AS AF	a	F
SG CM Ct	sc	5	d	AMN	pe	F
SG CM Pa Ct	r	3	d	AMN	pe	F
Si VE	r	2	d	AMN AMC AMS	pe	F
	r	1	d	AS	pe	K
Cl Sh	r	4	a	E AS AF	a	F
	e	1	a	AMN AS	a	F
	r	2	a	E AS	a	F, Sutorý 1982
Cl Sh Ah PS Si Sr SO Ab	c	5	a	E AS AF	a	F
	s	1	a		a	F
	r	2	a		a	F
Ae PT CE	la	5	d	E	pe	F, Lhotská 1975

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Myrrhoides nodosa</i> (L.) Cannon	Api	cas	neo	1997	H	M
<i>Narcissus poeticus</i> L.	Hya	cas	neo	1867	H	TM
<i>Narcissus pseudonarcissus</i> L.	Amr	cas	neo	1867	H	TM
<i>Nemophila menziesii</i> Hooker fil. et Arnott	Hydp	cas	neo		S	TM
<i>Nepeta cataria</i> L.	Lam	nat*	arN		H	T
<i>Nepeta ×faasenii</i> Stearn	Lam	cas	neo		NSH	T
<i>Nepeta grandiflora</i> M. Bieb.	Lam	cas	neo	1900	H	TM
<i>Nepeta racemosa</i> Lam.	Lam	cas	neo		NSH	T
<i>Neslia paniculata</i> (L.) Desv. subsp. <i>paniculata</i>	Bra	nat*	arR		H	T
<i>Nicandra physalodes</i> (L.) Gaertn.	Sol	cas	neo	1853	H	M
<i>Nicotiana alata</i> Link et Otto	Sol	cas	neo		H	M
<i>Nicotiana rustica</i> L.	Sol	cas	neo	1891	H	TM
<i>Nicotiana tabacum</i> L.	Sol	cas	neo	1891	H	TM
<i>Nigella arvensis</i> L.	Ran	nat*	arR		H	T
<i>Nigella damascena</i> L.	Ran	cas	neo	1874	H	TM
<i>Nigella sativa</i> L.	Ran	cas	neo		H	TM
<i>Nonea lutea</i> (Desr.) DC.	Bor	cas	neo		H	TM
<i>Nonea rosea</i> (M. Bieb.) Link	Bor	cas	neo	1872	H	TM
<i>Obione sibirica</i> (L.) Fischer	Chen	cas	neo	1939	H	M
<i>Ocimum basilicum</i> L.	Lam	cas	neo		H	TM
<i>Oenothera acutifolia</i> Rostański	Ona	cas	neo	1975	H	M
<i>Oenothera ×albipercurva</i> Renner	Ona	cas	neo	1899	H	M
<i>Oenothera ammophila</i> Focke	Ona	cas	neo	1848	H	M
<i>Oenothera biennis</i> L.	Ona	inv	neo	1831	SH	TM
<i>Oenothera canovirens</i> Steele	Ona	cas	neo	1953	H	M
<i>Oenothera coronifera</i> Renner	Ona	cas	neo	2001	H	M
<i>Oenothera depressa</i> Greene	Ona	nat	neo	1936	SH	TM
<i>Oenothera fallax</i> Renner emend. Rostański	Ona	nat	neo	1961	H	M
<i>Oenothera flava</i> subsp. <i>taraxacoides</i> (Woot. et Standl) W. L. Wagner	Ona	cas	neo	2000	H	TM
<i>Oenothera glazioviana</i> M. Micheli	Ona	nat	neo	1890	H	M
<i>Oenothera hoelscheri</i> Rostański	Ona	cas	neo	1975	H	M
<i>Oenothera issleri</i> Rostański	Ona	cas	neo	1949	H	M
<i>Oenothera missouriensis</i> Sims	Ona	cas	neo	1913	H	M
<i>Oenothera moravica</i> Jehlík et Rostański	Ona	cas	neo	1985	H	M
<i>Oenothera oakesiana</i> S. Watson et Coulter	Ona	cas	neo	1962	H	M
<i>Oenothera parviflora</i> L.	Ona	cas	neo	1914	H	M
<i>Oenothera ×punctulata</i> Rostański et Gutte	Ona	cas	neo	1972	H	M
<i>Oenothera pycnocarpa</i> Atkinson et Bartlett	Ona	nat	neo	1960	H	M
<i>Oenothera rubricaulis</i> Klebahn	Ona	nat	neo	1914	H	M
<i>Oenothera stricta</i> Ledeb.	Ona	cas	neo	1825	H	T
<i>Oenothera subterminalis</i> Gates	Ona	cas	neo	1967	H	M
<i>Oenothera tetragona</i> Roth	Ona	cas	neo	1884	H	TM
<i>Oenothera victorini</i> Gates et Catcheside	Ona	cas	neo	1973	H	M
<i>Omphalodes verna</i> Moench	Bor	cas	neo		SH	T
<i>Obobrychis vicifolia</i> Scop.	Fab	nat*	neo	1852	NS	T
<i>Onopordum acanthium</i> L.	Com	nat*	arM		H	T
<i>Onopordum ×beckianum</i> John	Com	cas	neo	1906	H	TM
<i>Opuntia phaeacantha</i> Engelm.	Cac	cas#	neo		S	T
<i>Ornithogalum nutans</i> L.	Lil	nat	neo	1809	SH	T
<i>Ornithopus compressus</i> L.	Fab	cas	neo	1937	H	M
<i>Ornithopus sativus</i> Brot. subsp. <i>sativus</i>	Fab	cas	neo	1889	SH	T
<i>Orobanche crenata</i> Forskål	Oro	cas	neo	1896	H	T
<i>Orobanche gracilis</i> Sm.	Oro	nat	neo	1878	S	T
<i>Orobanche hederæ</i> Duby	Oro	nat#	neo	1945	H	TM
<i>Orobanche lucorum</i> A. Br.	Oro	nat#	neo		H	TM
<i>Orobanche minor</i> Sm.	Oro	inv	ar		H	T
<i>Orobanche nana</i> (Reuter) Beck	Oro	cas	neo	1985	H	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	s	1	a	E AS	a	K, Filippov 1999
Ar Ae	r	4	d	E	pe	K
Ar Ae	r	4	d	E	pe	K
	r	1	d	AMN	a	K
Al Oa Si	sc	5	ad	E AS	pe	F
	r	2	d		pe	F
	r	2	d	E	pe	F, Holub 1991
	r	2	d	E AS	pe	F
Cl Sh Ah VE Si	c	5	a	AS	a	F
	r	2	d	AMS	a	F
	r	1	d	AMS	a	F
	r	1	d	AMN	a	F
	r	1	d	AMS	a	F
Cl Oa	sc	4	a	E AS AF	a	F
	r	3	d	E	a	F
	r	2	d	AS	a	F
	r	1	d	AS	a	F
	r	1	a	E	a	F
	e	1	a	AS	a	F
	r	1	d	AS	a	F
	r	1	a		b	F
	r	1	a		b	F
DM	r	2	ad		b	F
DM	c	5	ad		b a	F, Jehlík & Rostaňski 1980
DM	r	1	a		b	F
DM	s	1	a		b	
DM	r	3	a		b	F
DM	sc	3	a		b	F, Roubal 1972
	s	1	d	AMN	pe	K, Chrtek & Skočdoplová 2001a, Procházka 2002
DM	sc	3	d		b	F
DM	r	2	a		b	F
	r	2	a		b	F
	r	1	d	AMN	pe	F
	r	1	a		b	F
	e	1	a		b	F
	r	1	ad		b	F
	s	1	a		b	F
DM	r	3	a		a b	F
DM	sc	4	a		b	F, Roubal 1968
	e	1	a	AMS	a b	F
	r	1	a		b	F
	e	1	ad	AMN	pe	F
	r	2	a		b	F
	r	2	d	E	pe	F
Br Fv Ar	sc	5	d	E AS	pe	F
Oa	la	5	a	E	b	K
	se	1	a		b	Sutový 2001
	r	1	d		pe	K
Ae An CR	r	2	d	E	pe	K
	e	1	a	E AS AF	a	F
	r	3	ad	E	a	F
	e	1	ad	E	b pe p	F
	e	1	a	E	b pe p	F
	r	1	d	E AS	b pe p	F
	s	1	d	E	b pe p	F
Sh Ah Ar	r	3	a	E AS AF	b pe p	F, Kropáč 1997, Jehlík 1998
	e	1	ad	E	a b pe p	F

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Orobancha ramosa</i> L.	Oro	nat*	arI		H	T
<i>Oxalis corniculata</i> L.	Oxa	nat	neo	1852	H	TM
<i>Oxalis debilis</i> Humboldt, Bonpland et Kunth	Oxa	cas	neo	1963	H	M
<i>Oxalis dilleanii</i> Jacq.	Oxa	nat	neo		H	TM
<i>Oxalis fontana</i> Bunge	Oxa	nat	neo	1852	H	TM
<i>Oxalis latifolia</i> Humboldt, Bonpland et Kunth	Oxa	cas	neo	1963	H	M
<i>Oxalis pes-caprae</i> L.	Oxa	cas	neo	1961	H	M
<i>Oxalis repens</i> Thunb.	Oxa	cas	neo		H	TM
<i>Oxybaphus nyctagineus</i> (Michx.) Sweet	Nyc	nat	neo	1843	H	TM
<i>Paeonia officinalis</i> L.	Pae	cas	neo		H	TM
<i>Panicum capillare</i> subsp. <i>barbipulvinatum</i> (Nash) Tzvelev	Gra	cas	neo	1968	H	M
<i>Panicum capillare</i> L. subsp. <i>capillare</i>	Gra	nat	neo	1940	H	M
<i>Panicum compressum</i> Bivona	Gra	cas	neo	1961	H	M
<i>Panicum dichotomiflorum</i> Michx.	Gra	cas	neo	1970	H	M
<i>Panicum miliaceum</i> subsp. <i>agricolum</i> H. Scholz et Mikoláš	Gra	cas	neo	1975	H	TM
<i>Panicum miliaceum</i> L. subsp. <i>miliaceum</i>	Gra	cas	arN		H	T
<i>Panicum miliaceum</i> subsp. <i>ruderalis</i> (Kitagawa) Tzvelev	Gra	nat	neo	1823	H	M
<i>Panicum obtusum</i> Humboldt, Bonpland et Kunth	Gra	cas	neo	1961	H	M
<i>Panicum oligosanthes</i> Schult.	Gra	cas	neo		H	M
<i>Papaver argemone</i> L.	Pap	nat*	arN		SH	T
<i>Papaver atlanticum</i> subsp. <i>mesatlanticum</i> (Maire) Kadereit	Pap	cas	neo	2001	H	M
<i>Papaver croceum</i> Ledeb.	Pap	cas	neo		H	TM
<i>Papaver dubium</i> L.	Pap	nat*	arM		SH	T
<i>Papaver hybridum</i> L.	Pap	cas	neo	1865	H	TM
<i>Papaver lecoqii</i> Lamotte	Pap	nat	ar		H	T
<i>Papaver pseudo-orientale</i> (Fedde) Medvedev	Pap	cas	neo		H	TM
<i>Papaver rhoeas</i> L.	Pap	nat*	arN		H	TM
<i>Papaver somniferum</i> L. subsp. <i>somniferum</i>	Pap	cas	ar		H	TM
<i>Parapholis strigosa</i> (Dum.) C. E. Hubbard	Gra	cas	neo		H	M
<i>Parentucellia viscosa</i> (L.) Caruel	Scr	cas	neo	1882	SH	T
<i>Parietaria judaica</i> L.	Urt	cas	neo		H	TM
<i>Parietaria officinalis</i> L.	Urt	nat*	ar		NSH	T
<i>Parietaria pennsylvanica</i> Willd.	Urt	cas	neo	2000	H	M
<i>Parthenocissus inserta</i> (Kerner) Fritsch	Vit	inv	neo	1900	NSH	TM
<i>Parthenocissus quinquefolia</i> (L.) Planchon	Vit	nat	neo		NSH	TM
<i>Pastinaca sativa</i> L. subsp. <i>sativa</i>	Api	nat*	ar		SH	TM
<i>Pastinaca sativa</i> subsp. <i>urens</i> (Godron) Čelak.	Api	nat	ar		SH	TM
<i>Paulownia tomentosa</i> (Thunb.) Steudel	Scr	cas	neo		H	M
<i>Peltaria alliacea</i> Jacq.	Bra	cas	neo	1993	N	T
<i>Pentaglottis sempervirens</i> (L.) Tausch ex L. H. Bailey	Bor	cas	neo	1989	H	TM
<i>Persicaria orientalis</i> (L.) Spach	Poly	cas	neo		H	TM
<i>Persicaria pennsylvanica</i> (L.) M. Gómez	Poly	cas	neo	1968	H	M
<i>Persicaria polystachya</i> (Wall. ex Meisner) H. Gross	Poly	inv	neo		SH	T
<i>Petasites japonicus</i> (Sieb. et Zucc.) F. W. Schmidt	Com	cas	neo		SH	TM
<i>Petroselinum crispum</i> (Mill.) A.W. Hill	Api	cas	ar		H	TM
<i>Petunia ×atkinsiana</i> Loudon	Sol	cas	neo		H	M
<i>Peucedanum austriacum</i> (Jacq.) Koch	Api	cas	neo	1837	S	T
<i>Phacelia campanularia</i> A. Gray	Hydp	cas	neo		H	TM
<i>Phacelia ciliata</i> A. Gray	Hydp	cas	neo		H	TM
<i>Phacelia tanacetifolia</i> Bentham	Hydp	cas	neo	1891	H	TM
<i>Phalaris arundinacea</i> var. <i>picta</i> L.	Gra	cas	neo		SH	T
<i>Phalaris brachystachys</i> Link	Gra	cas	neo	1961	H	TM
<i>Phalaris canariensis</i> L.	Gra	cas	neo	1867	H	TM
<i>Phalaris coerulescens</i> Desf.	Gra	cas	neo		H	M
<i>Phalaris minor</i> Retz.	Gra	cas	neo	1961	H	TM
<i>Phalaris paradoxa</i> L.	Gra	cas	neo	1961	H	TM
<i>Phaseolus coccineus</i> L.	Fab	cas	neo		H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Er PS	e	3	a	E AS AF	a b pe p	F, Jehlík 1998
SO	r	4	a	E AS AU AF	a b pe	F
VE	s	1	a	AMS	pe	F, Holub & Holubičková 1980, Jehlík 1995, 1998
VE PS SO	r	2	a	AMN	a b pe	F
SO CE Ae Cr Ai Sa	sc	5	a	AMN	a b pe	F
VE	r	1	a	AMC AMS	pe	F, Jehlík 1995, 1998
	r	1	a	AF	pe	Dvořák & Kühn 1966
MP VE	r	3	ad	AS AU	a b pe	F
Oa Ar	r	2	ad	AMN AMC	pe	F, Jehlík 1998
	r	2	d	E AS	pe	F
	r	1	a	AMN	a	K, Jehlík 1998
Pa Si	sc	2	a	AMN	a	K, Jehlík 1998
	se	1	a	E	a	Dvořák & Kühn 1966
	sc	2	a	AMN	a	K, Jehlík 1998
PS Er Si	sc	2	a	E	a	K, Jehlík 1998
Si PS	r	4	d	AS	a	K
	sc	2	a	AS	a	K, Jehlík 1998
	se	1	a	AMC AMS	pe	K, Dvořák & Kühn 1966
	r	1	a	AMC AMS	pe	K
Ah Cl Sh Fv Ab	c	5	a	E	a	F
	r	1	d	AF	pe	
SO	r	1	d	AS	pe	F
Ah Si Cl Sh AS	sc	5	a	E AS AF	a	F
	e	1	a	E AS AF	a	F
	r	1	a	E	a	F
	r	1	d	AS	pe	F
Cl Sh Ah Si	c	5	a	E AS AF	a	F
Si	sc	4	d	E AS	a	F
	r	1	a	E	a	K
	e	1	a	E	ap	F
	e	1	a	E	a pe	F
TA Ai BS IS GA Al	sc	3	ad	E	pe	F
Si	r	1	a	AMN	a	K
Ai BS CR Ae	la	5	d	AMN	s	F
BS Ae	sc	3	d	AMN	s	F
Ar DM	c	5	ad	AS	b pe	F
Ar DM	sc	4	a	AS	b pe	F
	r	2	d	AS	t	F
	s	1	a	E	pe	K, Mandák 1995
	s	1	d	E	pe	F, Zlámalík 1996, Holub 1996
	r	2	d	AS	a	F
	r	1	a	AMN	a	K
Ae	sc	3	d	AS	pe	F
	r	1	d	AS	pe	K
Si	sc	4	d	E AS	b	F
Si	r	4	d	AMS	a	F
	e	1	a	E	pe	F
	r	1	d	AMN	a	F
	r	1	d	AMN	a	F
Ah Sh Si	r	4	d	AMN	a	F
Ae Al Ar	sc	5	d		pe	K
	se	2	a	E AS	a	K, Dvořák & Kühn 1966
	sc	2	a	E	a	K, Dvořák & Kühn 1966
	r	2	a	E	pe	K
	se	2	a	E AS AF	a	K, Dvořák & Kühn 1966
	se	2	a	E AS AF	a	K, Dvořák & Kühn 1966
	s	1	d	AMC AMS	pe	F

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Phaseolus vulgaris</i> L.	Fab	cas	neo		H	TM
<i>Philadelphus coronarius</i> L.	Phi	cas#	neo	1819	SH	TM
<i>Phleum paniculatum</i> Huds.	Gra	cas	neo		H	M
<i>Phleum subulatum</i> (Savi) A. et Gr.	Gra	cas	neo		H	M
<i>Phlox drummondii</i> Hooker	Pole	cas	neo		H	M
<i>Phlox paniculata</i> L.	Pole	cas	neo	1880	H	TM
<i>Phlox subulata</i> L.	Pole	cas	neo		H	M
<i>Pholiurus incurvus</i> Schinz et Thell.	Gra	cas	neo	1961	H	M
<i>Physalis alkekengi</i> L. var. <i>alkekengi</i>	Sol	nat	ar		NSH	TM
<i>Physalis alkekengi</i> var. <i>franchetii</i> (Masters) Makino	Sol	cas	neo		SH	TM
<i>Physalis angulata</i> L.	Sol	cas	neo	1972	H	TM
<i>Physalis peruviana</i> L.	Sol	cas	neo		H	TM
<i>Physalis philadelphica</i> Lam.	Sol	cas	neo	1935	H	TM
<i>Physalis pubescens</i> L.	Sol	cas	neo	2001	H	T
<i>Physocarpus opulifolius</i> (L.) Maxim.	Ros	inv	neo	1874	NSH	T
<i>Phytolacca americana</i> L.	Phy	cas	neo	1853	NSH	T
<i>Phytolacca esculenta</i> Van Houtte	Phy	nat	neo	1956	H	M
<i>Pimpinella anisum</i> L.	Api	cas	neo		H	T
<i>Pinus nigra</i> Arnold	Pin	nat	neo		NS	T
<i>Pinus strobus</i> L.	Pin	inv	neo	1800	N	T
<i>Pistia stratiotes</i> L.	Ara	cas	neo		N	TM
<i>Pisum sativum</i> L.	Fab	cas	ar		H	TM
<i>Plantago afra</i> L.	Plan	cas	neo	1851	H	TM
<i>Plantago alpina</i> L.	Plan	cas#	neo	1934	N	T
<i>Plantago coronopus</i> L. subsp. <i>coronopus</i>	Plan	cas	neo	1935	H	M
<i>Plantago gentianoides</i> Sibth. et Sm..	Plan	cas	neo		H	TM
<i>Plantago major</i> L. subsp. <i>major</i>	Plan	inv	ar		H	TM
<i>Plantago ×mixta</i> Domin	Plan	cas	arch		SH	T
<i>Plantago ×moravica</i> Chrtk	Plan	cas	arch		SH	T
<i>Platanus ×hispanica</i> Mill.	Plat	cas	neo		H	M
<i>Platycladus orientalis</i> (L.) Franco	Cup	cas	neo	1950	N	T
<i>Polycarpon tetraphyllum</i> (L.) L.	Car	cas	neo	1863	H	M
<i>Polycnemum arvense</i> L.	Chen	nat*	ar		NSH	T
<i>Polycnemum heuffelii</i> A. F. Láng	Chen	nat*	ar		NS	T
<i>Polycnemum majus</i> A. Braun	Chen	nat*	ar		NSH	T
<i>Polygonatum latifolium</i> (Jacq.) Desf.	Lil	nat	neo	1809	H	M
<i>Polygonum aviculare</i> L.	Poly	nat*	ar		H	TM
<i>Polyopogon fugax</i> Nees ex Steud.	Gra	cas	neo	1964	H	M
<i>Polyopogon monspeliensis</i> (L.) Desf.	Gra	cas	neo	1961	H	M
<i>Populus balsamifera</i> L.	Sal	cas	neo	1880	H	M
<i>Populus ×canadensis</i> Moench	Sal	inv	neo		SH	TM
<i>Portulaca grandiflora</i> Hooker	Por	cas	neo	1937	H	M
<i>Portulaca oleracea</i> L. subsp. <i>oleracea</i>	Por	nat*	arR		H	TM
<i>Potentilla fruticosa</i> L.	Ros	cas#	neo	1977	S	T
<i>Potentilla intermedia</i> L.	Ros	nat	neo	1903	H	M
<i>Potentilla supina</i> subsp. <i>paradoxa</i> (Nutt.) Soják	Ros	cas	neo		SH	T
<i>Primula vulgaris</i> Huds. subsp. <i>vulgaris</i>	Pri	nat	neo		SH	T
<i>Prunus armeniaca</i> L.	Ros	cas	ar		H	TM
<i>Prunus cerasifera</i> Ehrh.	Ros	nat	neo		SH	T
<i>Prunus cerasus</i> L.	Ros	nat*	ar		NSH	T
<i>Prunus domestica</i> L.	Ros	nat*	ar		SH	T
<i>Prunus ×emimens</i> G. Beck	Ros	nat*	ar		NS	T
<i>Prunus ×fruticans</i> Weihe	Ros	nat	ar		S	T
<i>Prunus institia</i> L.	Ros	nat*	ar		SH	T
<i>Prunus laurocerasus</i> L.	Ros	cas	neo	2001	H	M
<i>Prunus persica</i> (L.) Batsch	Ros	cas	ar		H	TM
<i>Prunus serotina</i> Ehrh.	Ros	inv	neo		NS	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	r	2	d	AMS	a	F
	r	3	d	E	s	F
	r	1	a	E	a pe	K
	r	1	a	E	a	K
	r	1	d	AMN	a	F
	r	2	d	AMN	pe	F
	r	2	d	AMN	pe	F
	se	1	a	E AS	a	Dvořák & Kühn 1966
CR BS Ai Ae	sc	4	d	E	pe	F, Hendrych 1989
	r	1	d	AS	pe	F
	se	1	d	AMN AMC AMS	a	F
	r	1	d	AMS	pe	F
	r	2	d	AMC	a	F, Pyšek 1995
	s	1	d	AMN AMC AMS	a	F
Ai	la	4	d	AMN	s	F
	r	2	d	AMN	pe	F
Ar Ae Al	r	3	d	AS	pe	F, Skalický 1972
	r	2	d	AS	a	F
Br Fv Qp Cr DS	sc	5	d	E	t	F
GQ	la	5	d	AMN	t	F
Le	e	1	d	AS AF AMC AMS	pe	K
Si	r	4	d	E AS	a	F
	se	1	a	E AS	a	F
Na NA	e	1	d	E	pe	F, Chrtek & Skočdoplová 1995
	e	1	a	E	a pe	F
	e	1	d	E	pe	F
MP Sg Si Cy	c	5	a	E AS AF	pe	F
	e	1	a		pe	F
	r	1	a		pe	F
	r	1	d		t	F
	r	1	d	AS	s t	F
MP	r	1	a	E	a	F
Co PF Ah Sn Ab	r	4	a	E AS	a	F
Cl AS	e	2	a	E	a	F
Fv Ah Si Cl DM	r	4	a	E AS	a	F, Novák 2001
	r	1	d	E	pe	K
VE SO Si	c	5	a	E AS	a	F
	se	1	a	E AS AF	a	
	r	1	a	E AS AF	a	K, Dvořák & Kühn 1966
	r	1	d	AMN	t	K
SS St DM	la	4	ad		t	F
Si VE	r	1	d	AMS	a	F, Domin 1937a, Petřík 2001
Er PS MP	sc	5	a	AS AF	a	F
	s	1	d	AS	s	F
DM MP	r	3	a	E	b pe	F
Bi Pa	e	1	a	AMN AS	a pe	F
	r	2	a	E AS AF	pe	F
	r	3	d	AS	ts	F
BS Bd PR	sc	5	d	E AS	ts	F
PR Bd BS CR	sc	5	d	E AS	ts	F
BS Bd PR	sc	4	d		ts	F
Ps Gs Bd	sc	4	a		s	F
Bd Ai BS	r	4	a		s	F
BS Bd PR	sc	5	d	AS	ts	F
	r	1	d	E AS	s	F
	r	3	d	AS	ts	F
GQ SS	la	4	d	AMN	ts	F

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Prunus virginiana</i> L.	Ros	cas	neo		S	T
<i>Pseudolysimachion incanum</i> (L.) Holub subsp. <i>incanum</i>	Scr	cas	neo		H	TM
<i>Pseudolysimachion</i> × <i>neglectum</i> (Vahl) Trávníček	Scr	cas	neo	1940	N	T
<i>Pseudotsuga menziesii</i> (Mirbel) Franco	Pin	nat	neo		N	T
<i>Puccinellia gigantea</i> (Grossh.) Grossh.	Gra	cas	neo		H	M
<i>Pulmonaria sibirica</i> L.	Bor	cas	neo		SH	T
<i>Pulsatilla vulgaris</i> Mill.	Ran	cas	neo	1852	NS	T
<i>Pulsatilla slavica</i> Reuss	Ran	nat#	neo		N	T
<i>Puschkinia scilloides</i> Adams	Hya	nat#	neo	1856	NS	T
<i>Pyrethrum macrophyllum</i> (W. et K.) Willd.	Com	nat	neo		SH	T
<i>Pyrus</i> × <i>amphigena</i> Domin ex Dostálek	Ros	nat	ar		NS	T
<i>Pyrus communis</i> L.	Ros	nat	ar		NSH	TM
<i>Pyrus</i> × <i>nivalis</i> Jacq.	Ros	nat	ar		NS	T
<i>Quercus rubra</i> L.	Fag	inv	neo		N	TM
<i>Ranunculus acris</i> subsp. <i>friesianus</i> (Jordan) Rouy et Fouc.	Ran	cas	neo	1882	H	TM
<i>Ranunculus arvensis</i> L.	Ran	nat*	arB		H	T
<i>Raphanus raphanistrum</i> L.	Bra	nat*	arN		H	TM
<i>Raphanus sativus</i> L. subsp. <i>sativus</i>	Bra	cas	ar		H	TM
<i>Rapistrum rugosum</i> subsp. <i>orientale</i> (L.) Arcang.	Bra	cas	neo	1940	H	T
<i>Rapistrum rugosum</i> (L.) All. subsp. <i>rugosum</i>	Bra	cas	neo	1850	H	T
<i>Reseda alba</i> L. subsp. <i>alba</i>	Res	cas	neo	1840	H	TM
<i>Reseda lutea</i> L. subsp. <i>lutea</i>	Res	nat	arR		SH	T
<i>Reseda luteola</i> L.	Res	nat*	arN		SH	T
<i>Reseda odorata</i> L.	Res	cas	neo	1900	H	T
<i>Reseda phyteuma</i> L.	Res	nat*	ar		NSH	T
<i>Reynoutria</i> × <i>bohemica</i> Chrtek et Chrtková	Poly	inv	neo	1942	NSH	TM
<i>Reynoutria japonica</i> var. <i>compacta</i> Moldenke	Poly	cas	neo	1995	SH	TM
<i>Reynoutria japonica</i> Hoult. var. <i>japonica</i>	Poly	inv	neo	1892	SH	TM
<i>Reynoutria sachalinensis</i> (F. Schmidt) Nakai	Poly	inv	neo	1869	SH	TM
<i>Rhagadiolus stellatus</i> (L.) Gaertn.	Com	cas	neo	1929	H	M
<i>Rheum rhabarbarum</i> L.	Poly	cas	neo	1967	H	TM
<i>Rhus hirta</i> (L.) Sudw.	Ana	inv	neo	1900	SH	T
<i>Rhus toxicodendron</i> L.	Ana	cas	neo	1874	H	TM
<i>Ribes aureum</i> Pursh	Gro	cas	neo	1900	NSH	T
<i>Ribes odoratum</i> Wendl. fil.	Gro	nat	neo		SH	T
<i>Ribes rubrum</i> L.	Gro	nat*	neo	1809	NSH	T
<i>Ribes spicatum</i> Robson	Gro	cas	neo	1885	NSH	T
<i>Ricinus communis</i> L.	Eup	cas	neo	1996	H	TM
<i>Robinia pseudacacia</i> L.	Fab	inv	neo	1874	NSH	TM
<i>Rodgersia aesculifolia</i> Batalin	Sax	cas	neo	2001	S	T
<i>Rosa</i> × <i>alba</i> L.	Ros	cas#	neo	1874	H	T
<i>Rosa</i> × <i>centifolia</i> L.	Ros	cas#	ar		H	T
<i>Rosa foetida</i> J. Herrmann	Ros	cas	neo	1814	NS	T
<i>Rosa glauca</i> Pourr.	Ros	cas	neo	1874	SH	TM
<i>Rosa rugosa</i> Thunb.	Ros	nat#	neo	1950	SH	TM
<i>Rosa villosa</i> L.	Ros	cas	ar		SH	TM
<i>Rostraria cristata</i> (L.) Tzvelev	Gra	cas	neo		H	M
<i>Rubia tinctorum</i> L.	Rub	cas	neo	1800	H	M
<i>Rubus allegheniensis</i> Porter	Ros	nat	neo		NSH	T
<i>Rubus armeniacus</i> Focke	Ros	nat	neo		H	TM
<i>Rubus canadensis</i> L.	Ros	nat	neo		SH	TM
<i>Rubus illecebrosus</i> Focke	Ros	cas	neo		SH	TM
<i>Rubus laciniatus</i> Willd.	Ros	nat	neo		NSH	TM
<i>Rubus moschus</i> Juz.	Ros	nat	neo		SH	TM
<i>Rubus occidentalis</i> L.	Ros	cas	neo	1997	S	T
<i>Rubus odoratus</i> L.	Ros	nat	neo	1880	NS	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	s	1	d	AMN	t s	F
	r	1	d	E AS	pe	F, Trávníček 1998
	se	1	d		pe	F
Cr GQ LF	r	3	d	AMN	t	F
	r	1	a	AS	a pe	K
	r	1	d	AS	pe	F
Fv HS	r	2	d	E	pe	F
	s	1	d	E	pe	F, Šuk 2001
Ar	r	3	d	AS	pe	K
Ae GA	r	2	d	E	pe	K
Bd	sc	4	ad		t	F
Bd BS	sc	5	d	AS	t	F
Bd	e	2	d	E AS	t	F
GQ LF Cr	sc	5	d	AMN	t	F
Ar Cy	r	1	a	E	pe	F
Cl	sc	5	a	E AS AF	a	F
Ah Sh SO Si	c	5	a	E AS	a	F
	r	2	d	E AS	a b	F
Si Cl Al	r	2	a	E AS	a b	F
Si Cl Al	r	3	a	E AS	a b	F, Hejný et al. 1973
	r	1	d	E	a	F
Oa DM CA Br	sc	5	a	E AS	pe a b	F
Oa Al	sc	5	ad	E	b	F
	r	2	d		a	F
Cl	r	1	a	E	a b	F, Roubal 1984, Hendrych 1978
Sf Ae BS	c	5	ad		pe	F, Mandák & Pyšek 1997
	r	1	d	AS	pe	K, Hlaváček et al. 1996, Mandák & Pyšek 1997
Sf Ae BS	c	5	d	AS	pe	F, Beerling et al. 1994, Mandák & Pyšek 1997
Sf Ae BS	la	4	d	AS	pe	F, Beerling et al. 1994, Mandák & Pyšek 1997, Sukopp & Sukopp 1995
	e	1	a	E AS	a	K
	r	2	d	AS	pe	F
	la	3	d	AMN	s t	F
	r	1	d	AMN	s	F
	r	1	d	AMN	s	F
	sc	3	d	AMN	s	F
Ai BS	sc	5	d	E	s	F
	r	1	d	E	s	F
	r	1	d	AF	a s s t	F
BS TA CR BR	c	5	d	AMN	t s	F
	s	1	d	AS	pe	
	r	3	d	AS	s	F
	r	3	d	E	s	F
	r	1	d	AS	s	F
	r	1	d	E	s	F
	r	3	d	AS	s	F
	r	2	d	E	s	F
	r	1	a	E	a	K
	r	2	d	E AS	pe	F
GQ Bd PR	r	2	d	AMN	s	F
Al PR BS	r	2	d	E	s	F
	r	1	d	AMN	s	F, Žižla & Chán 2001
	s	1	d	AS	ss	F
CE	r	2	d		s	F
	s	1	d	E AS	s	F
	s	1	d	AMN	s	F
GQ Cr CE	sc	3	d	AMN	s	F

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Rubus parviflorus</i> Nutt.	Ros	nat	neo		NSH	TM
<i>Rubus phoenicolasius</i> Maxim.	Ros	cas	neo		H	TM
<i>Rubus sylvaticus</i> Weihe et Nees	Ros	nat	neo		S	TM
<i>Rubus tuberculatus</i> Bab.	Ros	nat	neo		H	TM
<i>Rubus ulmifolius</i> Schott	Ros	cas	neo		SH	TM
<i>Rubus xanthocarpus</i> Bureau et Franchet	Ros	nat	neo	1962	S	T
<i>Rudbeckia hirta</i> L.	Com	nat	neo	1873	H	TM
<i>Rudbeckia laciniata</i> L.	Com	inv	neo	1859	NSH	TM
<i>Rumex acetosa</i> L. × <i>R. thyrsiflorus</i> Fingerh.	Poly	cas	neo		SH	T
<i>Rumex alpinus</i> L.	Poly	inv	neo	1819	SH	T
<i>Rumex brownii</i> Campd.	Poly	cas	neo	1965	H	M
<i>Rumex ×corconticus</i> Kubát	Poly	cas	neo	1981	SH	T
<i>Rumex confertus</i> Willd.	Poly	cas	neo	1965	SH	TM
<i>Rumex dentatus</i> subsp. <i>halacsyi</i> (Rech.) Rech. fil.	Poly	cas	neo	1965	H	M
<i>Rumex ×hybridus</i> Kindberg	Poly	cas	neo	1981	SH	T
<i>Rumex longifolius</i> DC.	Poly	inv	neo	1961	SH	T
<i>Rumex ×mezei</i> Haussknecht	Poly	cas	neo	1980	S	T
<i>Rumex obovatus</i> Danser	Poly	cas	neo		H	M
<i>Rumex patientia</i> L. subsp. <i>patientia</i>	Poly	nat	neo	1861	H	M
<i>Rumex ×propinquus</i> Aresch	Poly	cas	neo	1984	H	T
<i>Rumex scutatus</i> L.	Poly	nat	neo	1818	NSH	T
<i>Rumex thyrsiflorus</i> Fingerh.	Poly	inv	neo		NSH	T
<i>Rumex triangulivalvis</i> (Danser) Rech. fil.	Poly	nat	neo	1943	H	M
<i>Ruta graveolens</i> subsp. <i>hortensis</i> (Mill.) Gams	Rut	cas	neo	1874	NSH	TM
<i>Sagina apetala</i> Ard. subsp. <i>apetala</i>	Car	nat*	ar		S	T
<i>Sagina apetala</i> subsp. <i>erecta</i> (Hornem.) F. Hermann	Car	nat*	ar		SH	T
<i>Sagittaria latifolia</i> Willd.	Alis	cas	neo		N	M
<i>Salix ×sepulcralis</i> Simk.	Sal	cas	neo	2001	N	T
<i>Salix acutifolia</i> Willd.	Sal	nat	neo		N	T
<i>Salsola collina</i> Pallas	Chen	cas	neo		H	M
<i>Salvia officinalis</i> L.	Lam	cas	neo	1880	SH	TM
<i>Salvia reflexa</i> Hornem.	Lam	cas	neo	1934	H	TM
<i>Salvia sclarea</i> L.	Lam	cas	neo	1809	SH	TM
<i>Salvia spinosa</i> L.	Lam	cas	neo	1966	H	M
<i>Salvia splendens</i> Ker-Gawl.	Lam	cas	neo		H	TM
<i>Salvia verbenaca</i> L.	Lam	cas	neo	1965	H	M
<i>Salvia viridis</i> L.	Lam	cas	neo	1908	H	TM
<i>Sambucus ebulus</i> L.	Cap	nat*	ar		SH	T
<i>Sanguisorba minor</i> subsp. <i>polygama</i> (W. et K.) Holub	Ros	nat	neo	1840	NSH	TM
<i>Sanguisorba tenuifolia</i> Fisch. ex Link	Ros	cas	neo	1946	H	TM
<i>Saponaria ocymoides</i> L.	Car	cas	neo	1906	NSH	T
<i>Saponaria officinalis</i> L.	Car	nat*	arP		NSH	TM
<i>Satureja hortensis</i> L.	Lam	cas	neo		H	TM
<i>Saxifraga cuneifolia</i> L.	Sax	cas	neo		H	T
<i>Saxifraga cymbalaria</i> L.	Sax	cas	neo	1955	H	T
<i>Saxifraga ×geum</i> L.	Sax	nat	neo		NSH	T
<i>Saxifraga hostii</i> Tausch subsp. <i>hostii</i>	Sax	nat	neo	1850	N	T
<i>Saxifraga hypnoides</i> L.	Sax	cas	neo	1819	N	T
<i>Saxifraga rotundifolia</i> L.	Sax	cas	neo	1956	S	T
<i>Scandix pecten-veneris</i> L. subsp. <i>pecten-veneris</i>	Api	nat*	ar		H	T
<i>Scilla amoena</i> L.	Hya	cas	neo	1809	H	T
<i>Scilla luciliae</i> (Boiss.) Speta	Hya	cas	neo		SH	T
<i>Scilla sibirica</i> Haw.	Hya	nat	neo	1867	H	T
<i>Scirpus pendulus</i> Mühlenb.	Cyp	cas	neo			
<i>Scleranthus annuus</i> L.	Car	nat*	arN		NSH	T
<i>Scleroblitum atriplicinum</i> (F. Mueller) Ulbrich	Chen	cas	neo	1963	H	M
<i>Sclerochloa dura</i> (L.) P. B.	Gra	nat*	ar		H	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	s	1	d	AMN	s	F
	r	1	d	AS	s	F
	s	1	d	E	s	F
PR	s	1	a	E	s	F
	s	1	d	E AF	s	F
	s	1	d	AS	pe	F, Holub & Palek 1981
	sc	2	d	AMN	a b p	K
Ae Sf St	c	5	d	AMN	pe	K, Francírková 2001
Ar	r	4	a		pe	F
Ra Ae Pe PT CE Ad	la	5	d	E AS	pe	F, Hendrych 2001
	se	1	a	AU	pe	F
	e	1	a		pe	F, Kubát 1985
Ar Al DM	r	1	a	E AS	pe	F, Jehlík & Kopecský 1967
	e	1	a	E AS AF	a	F
PT Ar Ae	r	2	a		pe	F, Kubát 1985
Ae Al Ar PT Pe	la	3	a	E	pe	F, Kubínová & Krahulec 1997, 1999
	e	1	a		pe	F, Kubát 1985
	e	1	a	AMS	a	F
Al DM	sc	3	ad	E AS	pe	F, Jehlík 1998
	r	1	a		pe	F, Kubát 1985
	e	3	d	E AS AF	pe	F
DM Ar PF CA	la	5	a	E AS	pe	F
Si Al DM	r	3	a	AMN	pe	F, Hejný 1949, Hejný et al. 1973, Jehlík 1998
Gs Fv	r	3	d	E	ss	F
Th	e	1	a	E	a	F
Th MP Ah Sg	e	2	a	E	a	F
	e	1	d	AMN	pe	K
Ph St	s	1	d		t	F
	r	2	d	E AS	s	F
	r	2	a	AS	a	F
	r	2	d	E AS	ss	F
	e	1	a	AMN	a	F
	r	1	d	E AS	b pe	F
	se	1	a	E AS AF	pe	F, Štěpánková 1999
	r	1	d	AMS	a	F
	e	1	a	E	pe	F
	r	1	d	E AS	a	F
Al Ae	sc	5	a	E AS	pe	F
Fv Br CA DM	r	4	a	E AS	pe	F, Holub 1978b
	e	1	d	AS	pe	F
	r	2	d	E	pe	F, Domin 1924, Michal 1949
DM CA PF Ar	sc	5	d	E AS	pe	F
Si	r	3	d	E AS	a b	F
	r	1	d	E	pe	F
	r	1	d	E	a b	K, Procházka et al. 1983
	sc	3	d	E	pe	F
DS	s	1	d	E	pe	F
	e	1	a	E	pe	F
	e	1	d	E AS	pe	F
Cl	r	3	a	E AS AF	a	F, Chrtek et al. 1968, Příhoda 2001
	r	1	d		pe	K
Ar	r	3	d	E AS	pe	K
Ar	sc	3	d	E AS	pe	K
	se	1	a	AMN	pe	Dostál 1989
Ah Sn Ab Co Sr PS SO	c	5	a	E AS	a	F
	e	1	a	AU	a	F
MP	r	5	a	E AS	a	K, Chrtek & Žáková 1990

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Scopolia carniolica</i> Jacq.	Sol	nat	neo	1866	NSH	T
<i>Scorpiurus muricatus</i> L.	Fab	cas	neo		H	M
<i>Scrophularia canina</i> L.	Scr	cas	neo	1961	H	M
<i>Scrophularia chrysantha</i> Jaub. et Spach	Scr	cas	neo	1855	SH	T
<i>Scutellaria altissima</i> L.	Lam	nat	neo	1901	SH	T
<i>Secale cereale</i> L.	Gra	cas	arB		H	TM
<i>Sedum aizoon</i> L.	Cra	cas	neo	1880	SH	T
<i>Sedum annuum</i> L.	Cra	cas	neo		SH	T
<i>Sedum anopetalum</i> DC.	Cra	nat	neo		S	T
<i>Sedum hispanicum</i> L.	Cra	inv	neo		SH	TM
<i>Sedum hybridum</i> L.	Cra	nat	neo		SH	T
<i>Sedum pallidum</i> var. <i>bithynicum</i> (Boiss.) Chamberlain	Cra	cas	neo	2001	H	TM
<i>Sedum rupestre</i> subsp. <i>erectum</i> t'Hart	Cra	nat	neo		H	TM
<i>Sedum sarmentosum</i> Bunge	Cra	cas	neo		SH	TM
<i>Sedum spurium</i> M. Bieb.	Cra	nat	neo	1879	NS	T
<i>Sedum stoloniferum</i> S. Gmelin	Cra	cas	neo	2001	SH	TM
<i>Sempervivum tectorum</i> L.	Cra	nat	neo	1819	H	T
<i>Senecio inaequidens</i> DC.	Com	cas	neo	1997	H	M
<i>Senecio rupestris</i> W. et K.	Com	cas	neo	1879	S	T
<i>Senecio vernalis</i> W. et K.	Com	nat	neo	1872	H	TM
<i>Senecio vulgaris</i> L.	Com	nat*	ar		H	TM
<i>Setaria adhaerens</i> (Forskål) Chiovenda	Gra	cas	neo		H	M
<i>Setaria faberi</i> Herrmann	Gra	nat	neo	1961	H	M
<i>Setaria gussonei</i> Kerguelen	Gra	nat*	ar		H	TM
<i>Setaria italica</i> (L.) P. B. subsp. <i>italica</i>	Gra	cas	ar		H	TM
<i>Setaria italica</i> subsp. <i>moharia</i> (Alef.) Körnicke	Gra	cas	ar		H	TM
<i>Setaria pumila</i> (Poiret) R. et Sch.	Gra	nat*	arN		H	TM
<i>Setaria verticillata</i> (L.) P. B.	Gra	nat*	arM		H	TM
<i>Setaria viridis</i> subsp. <i>pyncocoma</i> (Steud.) Tzvelev	Gra	cas	neo		H	M
<i>Setaria viridis</i> (L.) P. B. subsp. <i>viridis</i>	Gra	nat*	arN		H	TM
<i>Sherardia arvensis</i> L.	Rub	nat*	ar		H	T
<i>Schismus barbatus</i> (L.) Thell.	Gra	cas	neo	1961	H	M
<i>Schkuhria pinnata</i> (Lam.) O. Kuntze	Com	cas	neo	1950	H	T
<i>Sicyos angulata</i> L.	Cuc	cas	neo	1880	SH	TM
<i>Sida hermaphrodita</i> (L.) Rusby	Mal	cas	neo	1958	H	TM
<i>Sida rhombifolia</i> L. subsp. <i>rhombifolia</i>	Mal	cas	neo	1979	H	M
<i>Sida spinosa</i> L.	Mal	cas	neo	1972	H	M
<i>Silene armeria</i> L.	Car	cas	neo	1850	H	TM
<i>Silene cretica</i> subsp. <i>annulata</i> (Thore) Hayek	Car	cas	neo	1941	H	T
<i>Silene dichotoma</i> Ehrh.	Car	nat	neo	1841	SH	TM
<i>Silene gallica</i> L.	Car	cas	ar		H	T
<i>Silene ×hampeana</i> Meusel et Werner	Car	nat	ar		NS	T
<i>Silene latifolia</i> subsp. <i>alba</i> (Miller) Greuter et Burdet	Car	nat*	arN		SH	TM
<i>Silene noctiflora</i> L.	Car	nat	arI		H	T
<i>Silene ×grecescuii</i> Gusul.	Car	cas	neo	1972	H	TM
<i>Silene pendula</i> L.	Car	cas	neo	1896	H	TM
<i>Silene viridiflora</i> L.	Car	cas#	neo	1971	NS	T
<i>Silphium perfoliatum</i> L.	Com	cas	neo	1885	N	T
<i>Silybum marianum</i> (L.) Gaertner	Com	cas	neo	1872	H	M
<i>Sinapis arvensis</i> L.	Bra	nat*	arN		H	TM
<i>Sisymbrium altissimum</i> L.	Bra	nat	neo	1815	SH	TM
<i>Sisymbrium austriacum</i> Jacq. subsp. <i>austriacum</i>	Bra	cas	neo	1858	NSH	T
<i>Sisymbrium irio</i> L.	Bra	cas	neo	1851	H	TM
<i>Sisymbrium loeselii</i> L.	Bra	inv	neo	1819	H	TM
<i>Sisymbrium officinale</i> (L.) Scop.	Bra	nat	arN		H	TM
<i>Sisymbrium orientale</i> subsp. <i>macroloma</i> (Pomel) Dvořák	Bra	cas	neo	1958	H	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
TA Ae	r	1	d	E	pe	F
	r	1	a	E AS AF	a	F
	e	1	a	E AS AF	pe	F
	e	1	d	E AS	b pe	F, Chrtek J. & Skočdoplová 1996
GA	sc	3	d	E AS	pe	F
Si	r	5	d		a	K
	r	2	d	AS	pe	F
	s	1	d	E	a b	F
	r	1	d	E AS	pe	F, Holub 1972
MP Ab	sc	4	d	E AS	pe	F
Cm	r	2	d	AS	pe	F
	s	1	d	E AS	pe	
Cm Ab	sc	3	d	E	pe	F
	r	1	d	AS	pe	F
Cm AS Ab	la	4	d	AS	pe	F
	r	1	d	AS	pe	
CA Ab	r	3	d	E	pe	F
Al	r	1	a	AF	pe	K, Mandák & Bimová 2001
	e	2	a	E	a	K
Si Ah Sh	la	5	a	E AS	a	K
VE Si	c	5	a	E AS	a	K
	r	1	a	AS AF AMC AMS	a	K
PS	sc	3	a	AS	a	K, Jehlík 1971a, 1998
Er Cl PS Si	r	2	a		a	K
Si PS	r	3	d		a	K
Si PS	r	3	d		a	K
Er Cl	c	5	a	E AS	a	K
PS Er	la	5	a	E	a	K
	r	1	a	E AS	a	K
PS Er DM Sc AS Si Sh	sc	5	a	E AS	a	K
Sh Cl Ah	sc	5	a	E AS AF	a	F
	se	1	a	E	a	K, Dvořák & Kühn 1966
	r	1	a	AMC AMS	a	K, Chrtek 1981
GA Al BS	r	3	d	AMN	a	F
	r	1	d	AMN	pe	F
	r	2	a	AMN AMC AMS	ss	F
				AS AF		
	r	2	a	AMN AMC AMS	pe ss	F
	r	3	ad	E	a	F
	e	1	a	E	a	F, Šourková 1978
DM VT Ar	sc	4	a	E AS	a b	F
Ah	r	3	a	E	a b	F
Ae IS	sc	4	a		?	F
CA DM Oa PR Bd Al VT	c	5	a	E AS AF	pe a	F
Cl VT Sh	c	5	a	E AS	a b	F
	e	1	a		?	F, Smejkal 1973
	r	1	d	E	a	F
Cr	se	1	d	E	pe	F, Smejkal 1973
	r	2	d	AMN	pe	K
Si Oa	r	3	d	E	a	K
VE SO PS Si	c	5	ad	E AS AF	a	F
Si Sr Er	c	5	a	E AS	a	F
	e	1	a	E	b pe	F
Si	r	2	a	E AS	a	F, Dvořák 1982
Si	c	5	a	E AS AF	a	F
Si Mn	c	5	a	E AS AF	a	F
	e	2	a	E	a	F

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Sisymbrium polymorphum</i> (Murray) Roth	Bra	cas	neo	1959	H	M
<i>Sisymbrium strictissimum</i> L.	Bra	nat	neo	1819	NSH	T
<i>Sisymbrium volgense</i> M. Bieb ex E. Fourn	Bra	nat	neo	1960	H	TM
<i>Sisyrinchium angustifolium</i> Mill.	Iri	nat	neo	1863	S	T
<i>Stium sisarum</i> L.	Api	cas	neo		S	T
<i>Smyrniolum perfoliatum</i> L.	Api	nat	neo	1886	NSH	TM
<i>Solanum americanum</i> Mill.	Sol	cas	neo	1966	H	M
<i>Solanum carolinense</i> L.	Sol	cas	neo	1985	H	M
<i>Solanum cornutum</i> L.	Sol	cas	neo	1899	H	TM
<i>Solanum decipiens</i> Opiz	Sol	nat	neo	1819	H	TM
<i>Solanum linneanum</i> Hepper et Jaeger	Sol	cas	neo		H	TM
<i>Solanum lycopersicum</i> L.	Sol	cas	neo	1880	SH	TM
<i>Solanum melongena</i> L.	Sol	cas	neo		H	TM
<i>Solanum nigrum</i> L.	Sol	nat*	arN		H	TM
<i>Solanum physalifolium</i> Rusby	Sol	cas	neo	1975	H	M
<i>Solanum pseudocapsicum</i> L.	Sol	cas#	neo		H	TM
<i>Solanum pyracanthos</i> Lam.	Sol	cas	neo	1940	H	M
<i>Solanum scabrum</i> Mill.	Sol	cas	neo	1975	H	TM
<i>Solanum sisymbriifolium</i> Lam.	Sol	cas	neo	1935	H	TM
<i>Solanum triflorum</i> Nutt.	Sol	cas	neo	1914	H	M
<i>Solanum tuberosum</i> L.	Sol	cas	neo		H	TM
<i>Solanum villosum</i> Mill.	Sol	cas	neo	1850	H	M
<i>Solidago canadensis</i> L.	Com	inv	neo	1838	NSH	TM
<i>Solidago gigantea</i> Aiton	Com	inv	neo	1851	NSH	TM
<i>Solidago graminifolia</i> (L.) Salisb.	Com	cas	neo		H	TM
<i>Sonchus arvensis</i> L. subsp. <i>arvensis</i>	Com	nat*	arM		SH	TM
<i>Sonchus asper</i> (L.) Hill	Com	nat	arM		H	TM
<i>Sonchus oleraceus</i> L.	Com	nat*	ar		H	TM
<i>Sorbaria sorbifolia</i> (L.) A. Braun	Ros	nat#	neo	1940	NS	T
<i>Sorbus domestica</i> L.	Ros	cas	neo		NS	T
<i>Sorghum bicolor</i> (L.) Moench	Gra	cas	neo		H	M
<i>Sorghum halepense</i> (L.) Pers.	Gra	cas	neo	1927	H	M
<i>Sorghum sudanense</i> (Piper) Stapf	Gra	cas	neo		H	M
<i>Spergula arvensis</i> L. subsp. <i>arvensis</i>	Car	nat*	arN		H	T
<i>Spergula arvensis</i> L. subsp. <i>arvensis</i> × <i>S. sativa</i>	Car	cas	ar		H	T
<i>Spergula arvensis</i> subsp. <i>linicola</i> (Boreau) Janchen	Car	cas	ar		H	T
<i>Spergula arvensis</i> subsp. <i>maxima</i> (Weihe) O. Schwarz	Car	cas	ar		H	T
<i>Spergula arvensis</i> subsp. <i>sativa</i> (Boenn.) Čelak.	Car	nat	arB		H	T
<i>Spinacia oleracea</i> L.	Chen	cas	arM		H	TM
<i>Spiraea alba</i> Duroi	Ros	cas	neo		H	T
<i>Spiraea ×billardii</i> Dippel	Ros	cas	neo		H	TM
<i>Spiraea chamaedryfolia</i> L.	Ros	cas	neo	1900	NSH	T
<i>Spiraea crenata</i> L.	Ros	nat	neo	1889	N	T
<i>Spiraea douglasii</i> Hooker	Ros	cas#	neo	1940	SH	T
<i>Spiraea ×macrothyrsa</i> Dippel	Ros	cas#	neo			
<i>Sporobolus elongatus</i> (Lam.) R. Br.	Gra	cas	neo	1961	H	M
<i>Stachys affinis</i> Bunge	Lam	cas	neo	1924	H	T
<i>Stachys annua</i> (L.) L.	Lam	nat*	arM		H	T
<i>Stachys arvensis</i> (L.) L.	Lam	nat	arN		H	T
<i>Stachys byzantina</i> C. Koch	Lam	cas	neo		H	TM
<i>Stellaria pallida</i> (Dumort.) Murb.	Car	nat*	ar		NS	TM
<i>Symphoricarpos albus</i> (L.) Blake	Cap	inv	neo		NSH	TM
<i>Symphoricarpos orbiculatus</i> Moench	Cap	cas#	neo		H	TM
<i>Symphytum asperum</i> Lepechin	Bor	cas	neo	1941	H	TM
<i>Symphytum ×upplandicum</i> Nyman	Bor	nat	neo	1908	SH	T
<i>Syringa vulgaris</i> L.	Ole	inv	neo	1809	NSH	T
<i>Tagetes erecta</i> L.	Com	cas	neo		H	TM

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	e	1	a	E AS	pe	F, Dvořák 1981
Al GA Bd BS TA	sc	4	ad	E	pe	F
Al DM Si	r	3	a	E	pe	F, Jehlík 1971b, 1981, 1998, Hejný et al. 1973
Ar Ap	r	3	d	AMN	pe	K, Pospíšil 1952, Kotlaba 1952
	s	1	a	E	pe	K
GA CR Ai BS	r	2	ad	E AS	b	F, Křisa et al. 1968, Müller 1998
	r	1	a	AMN AMS	a	F
	e	1	a	AMN	pe	F
	r	1	a	AMN	a pe	F
Si VE Al	sc	4	a	E	a	F
	s	1	a	AF	a ss s	F
Bi Cb Si	la	5	d	AMC AMS	a	F
	r	1	d	AS AF	a pe	F
VE Si	c	5	a	E	a	F
	r	1	a	AMS	a	F
	r	1	d	AMS	ss	F
	e	1	d	AF	s pe	F
	r	1	d	AF	a	F
	r	2	ad	AMS	a pe	F
	e	2	a	AMN	a pe	F
Si VE	c	5	d	AMS	pe	F
	s	1	a	E AS AF	a	F
Ar DM Al Ae BS SS	c	5	d	AMN	pe	K
St Ar Ae Sf BS CR	c	5	d	AMN	pe	K
	r	1	d	AMN	pe	K
Si Pa VE	c	5	a	E	pe	K
VE Si	c	5	a	E	a	K
VE Si	c	5	a	E	a	K
Cr	r	2	d	AS	s	F
Bd	r	3	d	E AS AF	t	F
	r	2	d	AF	a	K
	r	1	a	E AS	pe	K, Hejný et al. 1973, Jehlík 1998
	r	1	a	AF	a	K
SO PS Ah	c	5	a	E AS	a	F
	r	2	a		a	F
Ah	e	1	ad	E AS	a	F
Ah	e	3	ad	E AS	a	F
SO PS Ah	sc	2	ad	E AS	a	F
Si	r	2	d	AS	a	F
	r	2	d	AMN	s	F
	r	2	d		s	F
	r	1	d	E AS	s	F
	s	1	d	E AS	s	F
	r	2	d	AMN	s	F
	r	1	d		s	F
	se	1	a	AS AF AMC AMS	pe	Dvořák & Kühn 1966
	e	2	d	AS	pe	K, Novák 1924, Chrtek 1994
Cl VE Si	r	5	a	E AS	a	F
Ah SO	e	2	a	E AS AF	a	F
	r	2	d	AS	pe	F
Ab MP	sc	5	a	E	a	F
BS	sc	5	d	AMN	s	F
BS Cr Ae	r	3	d	AMN	s	F
Ae Ar Ap	r	2	d	AS	pe	F, Smejkal 1978
Ae Pa	r	4	ad		pe	F
Bd BS	sc	5	d	E	st	F
	r	2	d	AMC	a	K

Taxon	Fam	Stat	Res	1st	Landuse	Landscape
<i>Tagetes patula</i> L.	Com	cas	neo		H	TM
<i>Tanacetum parthenium</i> (L.) Schultz-Bip.	Com	nat	ar		H	T
<i>Tanacetum vulgare</i> L.	Com	inv	ar		SH	TM
<i>Telekia speciosa</i> (Schreb.) Baumg.	Com	inv	neo		SH	T
<i>Tetragonia tetragonioides</i> (Pallas) O. Kuntze	Aiz	cas	neo	1918	H	TM
<i>Teucrium marum</i> L.	Lam	cas	neo		H	TM
<i>Teucrium polium</i> L.	Lam	cas	neo	1960	H	TM
<i>Teucrium scorodonia</i> L.	Lam	nat	neo	1806	SH	T
<i>Thladiantha dubia</i> Bunge	Cuc	cas	neo	1939	SH	TM
<i>Thlaspi arvense</i> L.	Bra	nat*	arN		H	TM
<i>Thlaspi kovatsii</i> Heuff.	Bra	cas	neo		H	M
<i>Thymus drucei</i> Ronniger	Lam	cas	neo	1974	NS	T
<i>Thymus vulgaris</i> L.	Lam	cas	neo		SH	TM
<i>Tilia tomentosa</i> Moench	Til	cas	neo	2001	H	M
<i>Torilis arvensis</i> (Hudson) Link subsp. <i>arvensis</i>	Api	nat*	ar		H	T
<i>Torilis nodosa</i> (L.) Gaertn.	Api	cas	neo		H	T
<i>Tragopogon dubius</i> Scop.	Com	nat	ar		SH	TM
<i>Tragopogon xmirabilis</i> Rouy	Com	cas	neo		H	TM
<i>Tragopogon porrifolius</i> L.	Com	cas	neo	1872	H	TM
<i>Tragus racemosus</i> (L.) All.	Gra	cas	neo		H	TM
<i>Tribulus terrestris</i> L.	Zyg	cas	neo		H	M
<i>Trifolium alexandrinum</i> L.	Fab	cas	neo	1960	H	M
<i>Trifolium angulatum</i> W. et K.	Fab	cas	neo	1976	N	T
<i>Trifolium angustifolium</i> M. Bieb.	Fab	cas	neo	1923	H	T
<i>Trifolium glomeratum</i> L.	Fab	cas	neo	1961	H	M
<i>Trifolium hybridum</i> L. subsp. <i>hybridum</i>	Fab	nat*	neo	1819	SH	TM
<i>Trifolium incarnatum</i> L. subsp. <i>incarnatum</i>	Fab	cas	neo	1870	SH	T
<i>Trifolium lappaceum</i> L.	Fab	cas	neo	1916	H	M
<i>Trifolium ornithopodioides</i> L.	Fab	cas	neo	1960	H	M
<i>Trifolium pallidum</i> W. et K.	Fab	cas	neo	1930	H	M
<i>Trifolium pannonicum</i> Jacq.	Fab	nat	neo	1919	NS	T
<i>Trifolium pratense</i> subsp. <i>americanum</i> (C. O. Harz) Soják	Fab	cas	neo	1880	SH	T
<i>Trifolium pratense</i> subsp. <i>sativum</i> (Schreber) Schübl. et Mart.	Fab	cas	neo		H	TM
<i>Trifolium resupinatum</i> L.	Fab	cas	neo	1853	H	TM
<i>Trifolium squamosum</i> L.	Fab	cas	neo	1930	H	M
<i>Trifolium subterraneum</i> L.	Fab	cas	neo	1962	H	M
<i>Trifolium tomentosum</i> L.	Fab	cas	neo	1961	H	M
<i>Trigonella caerulea</i> (L.) Ser.	Fab	cas	neo	1874	H	T
<i>Trigonella foenum-graceum</i> L.	Fab	cas	neo	1889	H	T
<i>Tripleurospermum inodorum</i> (L.) Schultze	Com	inv	arM		H	TM
<i>Triticum aestivum</i> L.	Gra	cas	ar		H	TM
<i>Triticum dicoccon</i> Schrank	Gra	cas	ar		H	T
<i>Triticum polonicum</i> L.	Gra	cas	neo		H	T
<i>Triticum turgidum</i> L.	Gra	cas	neo		H	T
<i>Tropaeolum majus</i> L.	Tro	cas	neo		H	TM
<i>Tulipa xgesnerana</i> L.	Lil	cas	neo		SH	TM
<i>Tulipa sylvestris</i> L.	Lil	cas	neo	1867	NSH	T
<i>Turgenia latifolia</i> (L.) Hoffm.	Api	nat*	ar		H	T
<i>Ulex europaeus</i> L.	Fab	cas#	neo	1880	S	T
<i>Urtica dodartii</i> L.	Urt	cas	neo		H	TM
<i>Urtica pilulifera</i> L.	Urt	cas	neo	1872	H	TM
<i>Urtica urens</i> L.	Urt	nat*	arN		H	T
<i>Vaccaria hispanica</i> subsp. <i>grandiflora</i> (Ser.) Holub	Car	nat*	arP		H	T
<i>Valerianella dentata</i> (L.) Pollich subsp. <i>dentata</i>	Val	nat*	arN		H	T
<i>Valerianella dentata</i> subsp. <i>eriosperma</i> (Wallr.) Holub	Val	nat*	ar		H	T
<i>Valerianella rimosa</i> Bast.	Val	nat*	ar		H	T
<i>Vallisneria spiralis</i> L.	Hydc	cas	neo		N	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
Si	r	3	d	AMC	a	K
Cm Al Si	sc	5	d	E AS	pe	K
Al CA Ar	c	5	a	E	pe	K
Ae Ai IS	sc	4	d	E	pe	K
	r	1	d	AMS AS AU	a	F
	r	1	d	E	s	F
	e	1	a	E AS AF	s	F
GQ LF CE	r	3	ad	E	pe ss	F, Kubát 1993
Si Ae	r	3	d	AS	pe	F
VE SO Ah	c	5	a	E AS	a b	F
	s	1	a	E	pe	F
	r	1	d	E	ss	F, Čáp 1982
	r	2	d	E	s ss	F
	r	1	d	E	t	F
Oa Al DM Cl	r	3	a	E AS	a	F
	e	1	a	E	a	F
DM Sc	c	5	a	E	a	K
	r	1	a		a pe	K
	r	1	d	E	a	K
Sr	r	1	a	E	a	K
Er MP	r	1	a	E AS	a	F
	e	1	ad	E	a	F
	e	1	a	E	a	F
	se	1	a	E AS	a	F
	e	1	a	E AS	a	F
Ap C Ar Pa	c	5	d	E AS	b pe	F
Pa Ar	sc	5	ad	E	a b	F
	e	1	a	E AS	a	F
	e	1	a	E	a	F
	e	1	a	E AS	a b	F
	r	1	ad	E	pe	F, Hendrych 1968
	e	2	d		pe	F
Pa Ar	sc	5	d		pe	F
	r	3	ad	AS	a	F
	e	1	a	E AS	a	F
	r	2	a	E AS AF	a	F
	e	1	a	E AS	a	F
	sc	3	d	E	a	F
	sc	3	d	E AS	a	F
Si DM	c	5	a	E	a	K
Si SO PS	sc	5	d		a	K
	r	1	a		a	K
	r	1	a	E	a	K
	r	1	a		a	K
Si VE	r	3	d	AMS	a	F
Ar Ae	sc	4	d		pe	K
Cr Ai TA	r	3	d	E	pe	K
Cl	e	3	a	E AS AF	a	F
	r	1	d	E	s	F
	r	1	d	E	pe	F
	r	2	d	E	a	F
Mn Cb	c	5	a	E AS	a	F
Cl	e	4	a	E	a	F
Sh Cl	sc	5	a	E AS AF	a	F, Holub 1978c
Cl Sh Fv	r	4	a	E	a	F
Cl Sh	r	4	a	E AF	a	F
	e	1	d	AS	pe	K

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Verbascum niveum</i> subsp. <i>visianinum</i> (Reichenb.) Murb.	Scr	cas	neo	1914	SH	TM
<i>Verbena bonariensis</i> L.	Ver	cas	neo	1983	H	M
<i>Verbena chamaedryfolia</i> Juss.	Ver	cas	neo	1853	H	M
<i>Verbena</i> × <i>hybrida</i> hort.	Ver	cas	neo		H	T
<i>Verbena officinalis</i> L.	Ver	nat*	arN		H	T
<i>Verbena rigida</i> Sprengel	Ver	cas	neo	1967	H	M
<i>Veronica acinifolia</i> L.	Scr	cas	neo	1908	H	TM
<i>Veronica agrestis</i> L.	Scr	nat*	ar		NSH	T
<i>Veronica arvensis</i> L.	Scr	nat*	ar		NSH	TM
<i>Veronica filiformis</i> Sm.	Scr	inv	neo	1938	SH	TM
<i>Veronica hederifolia</i> L.	Scr	inv	arN		NSH	TM
<i>Veronica opaca</i> Fries	Scr	nat*	ar		H	T
<i>Veronica peregrina</i> L. subsp. <i>peregrina</i>	Scr	cas	neo	1809	NSH	T
<i>Veronica persica</i> Poiret	Scr	inv	neo	1809	H	TM
<i>Veronica polita</i> Fries	Scr	nat*	arM		H	TM
<i>Veronica triloba</i> (Opiz) Wiesb.	Scr	nat*	ar		H	T
<i>Veronica triphyllos</i> L.	Scr	nat*	ar		H	T
<i>Vicia angustifolia</i> L.	Fab	nat*	ar		SH	TM
<i>Vicia articulata</i> Hornem.	Fab	cas	neo	1874	H	TM
<i>Vicia bithynica</i> (L.) L.	Fab	cas	neo	1949	H	TM
<i>Vicia cordata</i> Hoppe	Fab	nat*	ar		H	TM
<i>Vicia ervilia</i> (L.) Willd.	Fab	cas	neo	1874	H	T
<i>Vicia faba</i> L.	Fab	cas	neo		H	TM
<i>Vicia grandiflora</i> Scop. subsp. <i>grandiflora</i>	Fab	nat*	neo	1877	SH	T
<i>Vicia hirsuta</i> (L.) S. F. Gray	Fab	nat	arN		SH	TM
<i>Vicia lutea</i> L.	Fab	cas	neo		NSH	T
<i>Vicia melanops</i> Sibth. et Sm.	Fab	cas	neo	1900	SH	T
<i>Vicia narbonensis</i> L.	Fab	cas	neo		H	TM
<i>Vicia onobrychioides</i> L.	Fab	cas	neo	1980	H	TM
<i>Vicia pannonica</i> Crantz subsp. <i>pannonica</i>	Fab	nat	ar		SH	T
<i>Vicia pannonica</i> subsp. <i>striata</i> (M. Bieb.) Nyman	Fab	nat	neo		SH	T
<i>Vicia</i> × <i>poechhackeri</i> J. Murr	Fab	cas	neo		SH	T
<i>Vicia sativa</i> L.	Fab	nat*	ar		SH	TM
<i>Vicia villosa</i> subsp. <i>varia</i> (Host) Corb.	Fab	nat*	ar		NSH	T
<i>Vicia villosa</i> Roth subsp. <i>villosa</i>	Fab	nat*	arI		NSH	TM
<i>Viola canadensis</i> var. <i>rugulosa</i> (Greene) Hitchc.	Vio	cas#	neo	1948	H	TM
<i>Viola cornuta</i> L.	Vio	cas	neo	1959	NS	T
<i>Viola</i> × <i>haynaldii</i> Wiesb.	Vio	nat	neo	1886	NS	T
<i>Viola</i> × <i>hungarica</i> Degen et Sabr.	Vio	nat	arM		NS	T
<i>Viola</i> × <i>kernerii</i> Wiesb.	Vio	cas	neo	1904	N	T
<i>Viola obliqua</i> Hill.	Vio	cas	neo	1895	NS	T
<i>Viola odorata</i> L.	Vio	inv	arM		NSH	TM
<i>Viola</i> × <i>pluricaulis</i> Borbás	Vio	cas	arM		S	T
<i>Viola</i> × <i>poelliana</i> Murr.	Vio	nat	arM		NS	T
<i>Viola</i> × <i>porphyrea</i> Uechtr.	Vio	nat	arM		NSH	T
<i>Viola</i> × <i>scabra</i> F. Braun	Vio	inv	arM		NSH	TM
<i>Viola</i> × <i>sourekii</i> Procházka	Vio	cas	neo		S	T
<i>Viola suavis</i> M. Bieb.	Vio	nat	neo		NSH	TM
<i>Viola tricolor</i> L. subsp. <i>curtisii</i> (E. Forster) Syme	Vio	cas	neo	1953	N	T
<i>Viola tricolor</i> L. subsp. <i>tricolor</i>	Vio	nat*	ar		SH	T
<i>Viola</i> × <i>vindobonensis</i> Wiesb.	Vio	cas	neo		SH	TM
<i>Viola</i> × <i>wittrockiana</i> Gams	Vio	cas	neo		H	TM
<i>Virga strigosa</i> (R. et Sch.) Holub	Dip	inv	neo	1864	H	TM
<i>Vitis riparia</i> Michx	Vit	cas	neo	1964	SH	TM
<i>Vitis vinifera</i> L. subsp. <i>vinifera</i>	Vit	cas	arM		SH	TM
<i>Vulpia bromoides</i> (L.) S. F. Gray	Gra	nat	ar		SH	T

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	e	1	a	E	b	F
	e	1	ad	AMS	pe ss	F
	e	1	d	AMS	ss	F
	r	1	d		a	F
Al Pa MP	sc	5	a	E AS AF	pe a	F
	se	1	d	AMS	pe	F
Ah	r	1	a	E	a	F, Smejkal 1970
Sh VE Ab	r	5	a	E AF	a	F
AS Ab	c	5	a	E AS	a	F
Ap Ar Cy Pa Ae	sc	4	d	E AS	pe	F, Jehlík 1961, 1998, Jehlík & Slavík 1967, Hejný et al. 1973, Jehlík 1998
Ah Sh Si BS BR CR Cr	c	5	a	E AS AF	a	F
VE	r	4	a	E	a	F
Bi Es Sg	r	2	a	AMN AMC AMS	a	F, Peniašteková & Feráková 1993
VE Sh Ah Si	c	5	a	AS	a	F
VT VE Cl Mn	c	5	a	E	a	F
Cl	r	4	a	E	a	F
Ah Ab Sh Cl	sc	5	a	E	a	F
PS Ab Fv KP AS DM	c	5	a	E AS AF	a	F
	sc	4	d	E AS AF	a	F
	e	1	a	E AS	a	F, Sutorý 1976
	r	3	a	E AS	a	K
	r	2	d	E AS	a	F
	r	3	d	AS AF	a	F
Cl Sh Ah Ar CA	sc	4	a	E AS	a	F
Tm Ah KP PF Ab	c	5	a	E AS	a	F
Cl Fv Br Ar	r	2	a	E AS AF	a	F, Skřivánek 1949
	e	1	a	E	a	F
	r	2	ad	E AS	a	F
Ar	e	1	a	E	pe	F, Saul 1983
Ps Fv Br	sc	4	ad	E	a	F
Cl Fv Br Ar KP	r	3	a	E AS AF	a	F
	r	1	a		a	F
Cl Sh Ah Pa PS VT	c	5	ad	E AS AF	a	F
PF DM Cl Sh Ar	c	5	ad	E	a	F
Ah Sh Ar CA	c	5	ad	E AS	a b	F
	se	1	d	AMN	pe	F, Kirschner & Štěpánek 1984
Vc PT Ar	r	2	d	E	pe	F, Skalický 1973
Fv	r	1	a		pe	F
Fv Br Bd	r	1	a		pe	F
	e	1	a		pe	F
	se	1	d	AMN	pe	F, Kirschner & Štěpánek 1984
GA BS CR Cr Ai Ae	c	5	d	E AS AF	pe	F
	e	1	a		pe	F
	r	1	a		pe	F
Bd Cr Br GA	sc	4	a		pe	F
BS Cr BR Bd GA TA	sc	5	a		pe	F
PT	r	1	a		?	F
GA BS CR Ae	r	3	d	E AS	pe	F
Co	r	1	a	E	pe	F
SO Si Ar	sc	5	a	E	a	F
Bd TA BS GA	r	1	a		pe	F
Si	sc	4	d		a b	F
Al Ae GA	la	4	d	E AS	b	F, Lhotská 1968b
	r	1	d	AMN	s	F
BS CA PR Bd	r	4	d	E AS	t	F
Th	r	2	a	E	a b	K

Taxon	Fam	Stat	Res	Ist	Landuse	Landscape
<i>Vulpia ciliata</i> Dum.	Gra	cas	neo		H	M
<i>Vulpia ligustica</i> (All.) Link	Gra	cas	neo		H	M
<i>Vulpia myuros</i> (L.) C. C. Gmelin	Gra	nat	ar		SH	T
<i>Waldsteinia geoides</i> Willd.	Ros	cas	neo		S	T
<i>Waldsteinia trifolia</i> Rochel ex Koch	Ros	nat	neo		S	T
<i>Xanthium albinum</i> (Widd.) H. Scholz	Com	nat	neo		S	T
<i>Xanthium ripicola</i> Holub	Com	cas	neo	1872	H	TM
<i>Xanthium spinosum</i> L.	Com	nat*	neo	1872	H	T
<i>Xanthium strumarium</i> L.	Com	nat*	arB		H	TM
<i>Xanthophthalmum coronarium</i> (L.) Trehane	Com	cas	neo	1879	H	TM
<i>Xanthophthalmum segetum</i> (L.) Schultz-Bip.	Com	cas	neo	1872	H	T
<i>Zea mays</i> L.	Gra	cas	neo		H	M
<i>Zelkova serrata</i> (Thunb.) Mak.	Ulm	cas	neo	1973	N	TM
<i>Zinnia elegans</i> Jacq.	Com	cas	neo		H	TM

New additions to the alien flora of the Czech Republic:

Agrostis scabra. Třeboň region: sand pit near the village of Halámky in the southern part of the basin (2001, V. Horváthová – M. Štech, pers. com.). — *Alhagi pseudalhagi*. Raspenava, distr. Liberec: 1 specimen in the garden of the house no. 16, SW from the Pekelský vrch hill (1963, V. Jehlík PRA). Wool casual, native from E Europe to Middle Asia. — *Allium atropurpureum* W. et K. was found escaped from cultivation in Maršovice, South Moravia (BRNM – Krahulec, pers. com.). — *Bromus hordeaceus* subsp. *pseudothominii*. Hrdčovice and Nová Paka, distr. Jičín: rarely between tracks at railway stations in both settlements; Praha-Holešovice: W part of the Vltava river port (all localities 1971, V. Jehlík PRA). — *Carduus tenuiflorus*. Liberec: 1 specimen found on the wool-waste deposits of Textilana spinning mill (1967, V. Jehlík PRA). Taxonomic identity of this species requires further study (see Clement & Foster 1994: 300). — *Centaurea xgerstlaueri*. Jasenný, distr. Semily: rarely on railway station; Krásná Lípa: rarely on the main railway station; Tanvald, rarely at the periphery of the main railway station (all localities 1966, V. Jehlík PRA). — *Centaurea nigra* × *phrygia*. Jasenný, distr. Semily: rarely at railway station (1966, V. Jehlík PRA). — *Cerastium xmaureri*. Occurrence of the cross between the neophyte *C. tomentosum* and native *C. arvense* subsp. *arvense* was suspected but not confirmed by Smejkal in Hejny & Slavík (1992). In fact, it has been overlooked as it is rather common around cabins and cottages, in gardens and cemeteries. Selected localities: Praha 8, Trojská street: at a wall base (2001, P. Pyšek & B. Mandák); ruderal habitats in towns, e.g. Otradovice near Lysá nad Labem, Karlštejn, Český Krumlov, Horní Slavkov, Dvůr Králové nad Labem (J. Sádlo). — *Gilia capitata*. Žatec, on the bank of Ohře river along the road to Podbořany (1982, J. Houda PRA). The species is native to North America where it grows from Alaska to California, Arizona, Utah, Idaho (Kartesz & Meacham 1999). Since planting is not mentioned in Czech garden literature, it is uncertain whether it was introduced accidentally or escaped from cultivation. — *Helianthus strumosus*. Reported from two localities (Kutná hora, Březina). Seldom planted in gardens. Native to E North America (J. Kirschner & O. Šída, pers. com.). — *Hieracium pannosum*. It grows on the Kunětická hora hill near Pardubice (J. Chrtek jun., pers. comm.); its occurrence is associated with intentional introductions of many species which were planted at the locality in the 1930s by a natural-historical society from Pardubice (F. Procházka, pers. com.). — *Hordeum leporinum*. Liberec: several plants on a waste deposit near the Textilana spinning mill (1967, V. Jehlík PRA). Native to Mediterranean, including Atlantic Islands. — *Oenothera coronifera*. Zliv, distr. České Budějovice: cca 30 plants grew in a mixed stand with *O. issleri* at a railway station (2001, Mihulka et al. in prep.). — *Papaver atlanticum* subsp. *mesatlanticum*. Sušice, distr. Klatovy: yard of a school canteen (house no. 87/III); hospital: several plants near the entrance gate and by the nutrition department. It formerly occurred on several other localities in the town of Klatovy but disappeared due to building activities (2001, M. Král). The species originates from N Africa (Morocco) and is rarely planted in the Czech Republic. — *Parietaria pennsylvanica*. Praha 5, Na Čechelčce street; about 30 plants were first observed in 2000 (J. Sádlo PRA, in prep.). The species is native to N America, and might have been introduced into Czech Republic from Berlin where it is rather common. — *Polypogon fugax*. Česká Skalice II – Malá Skalice: one specimen on the yard of the Tiba spinning mill on the Úpa river bank (1964, V. Jehlík PRA). Wool or cotton casual native to Mediterranean, including Atlantic Islands. — *Rodgersia aesculifolia*. Příbram: wetland patch with *Menyanthes trifoliata* in a park near a football stadium (2001, P. Pyšek & J. Pyšková PRA). A single clone ca. 0.9 m in diameter was found; the character of the locality indicates accidental introduction. Native to China. — *Sedum pallidum* var. *bithynicum*. Beroun: it was found in 2001 at the Berounka

Syntaxa	Abund	LocNo	Intr	Origin	LH	Source
	r	2	a	E	a	K
	r	2	a	E	a	K
Th	sc	4	a	E	a b	K
	s	1	a	E	pe	F
	s	1	a	E	pe	F
PS Pa	la	4	a	AMN	a	K
	r	1	a	E	a	K
Oa Mn	r	2	a	AMS	a	K
Oa Mn Si PS Er	la	3	a	E AS	a	K, Opravit 1963
	r	2	d	E	a	K
Ah Sn	r	2	ad	E	a	K
Si	sc	4	d	AMC	a	K
	s	1	d	AS	t	K
	r	2	d	AMC	a	K

river bank during the floristic summer school organized by the Czech Botanical Society (det. M. Král). It is rarely planted (e.g. cemetery in Klatovy) and originates from SE Europe and SW Asia. — *Sedum stoloniferum*. Klatovy: two localities near the Větrovna quarry where the plants were probably introduced from the nearby cemetery (2001, M. Král, pers. com.). Native to Caucasus.

Species treated in the Flora of the Czech Republic or in Kubát et al. (2002) but not reported as escaping from cultivation:

Acer ginnala. Liberec: escaped and growing in a park hedgerow (2001, J. Sádlo). — *Acer monspessulanum*. Praha: Vinohrady hospital, young trees about ± 1 m tall were found growing in the park where adult trees are planted (2001, J. Sádlo). — *Aesculus xcarnea*. Semily: two-year old sapling was found at the periphery of railway station (1964, V. Jehlík PRA). — *Astilbe xarendsii*. Jilemnice, distr. Semily: ca 5 km NW of the town, near the Dolní Sytová village, ca 0.2 km from the bridge upstream across the Jizera river (1999, B. Mandák PRA). A single clone ca 1 m² in size was found; it corresponds to the cultivar grown during WWI (M. Opatrná, pers. com.). — *Beta vulgaris* group *Vulgaris*. Escapes by seed on field margins, along roads and paths, on compost piles, rubbish tips and in the vicinity of sugar refineries. — *Beta vulgaris* group *Cicla*. Escapes by seed, usually in garden composts, and occasionally persists for few years (Praha-Satalice, J. Sádlo). — *Buddleja davidii*. Praha 8: numerous population of shrubs of various ages (up to several years) originated from seed was found between the tram stops Trojská and Nad Trojou (2001, J. Pyšková & P. Pyšek). — *Campanula speciosa*. Pec pod Sněžkou (J. Sádlo). A garden escape found in the vicinity of mountain chalets and gardens; the localities represent clonally spreading cultivation relics. — *Castanea sativa*. Praha-Petřín; several localities in the České Středohoří hills (2000, J. Sádlo) and near Litošice in the Železné hory Mts (B. Mandák). — *Celtis occidentalis*. Praha: Vinohrady hospital (2001, J. Sádlo PRA). The species was reported in the Flora of the Czech Republic as very rarely escaping from cultivation, albeit without a concrete locality (an old record by Dostál 1954 from the vicinity of the town of Velvary is erroneous). The record reported here is the first evidence of cultivation escape and the mention in Kubát et al. (2002) is based on the same locality. — *Chamaecyparis lawsoniana*. National park Czech-Saxonian Switzerland (N Bohemia): self-seeding in a sandstone valley NE of the Koliště hill (P. Bauer, pers. com.). — *Chaenomeles japonica*. It grows on Kunětická hora hill near Pardubice as a cultivation relic planted in the 1930s by a natural-historical society from the town of Pardubice; it has spread since then and persists (F. Procházka, pers. com.). — *Chrysanthemum indicum*. Garden escape at rubbish tips in cemeteries and allotments. — *Corydalis alba* subsp. *alba*. Jindřichův Hradec (ca. 1995, J. Kolbek & J. Sádlo), Ronov nad Doubravou (ca 1998, J. Sádlo). Reported neither by the Flora of the Czech Republic nor by Kubát et al. (2002), only by Dostál (1989). It is seldom planted as a garden ornamental and very rarely escapes from cultivation. — *Corylus colurna*. Praha-Petřín: young shrubs escaped from cultivation in the park (2001, J. Sádlo). — *Cotoneaster bullatus*. Praha-Libeň: along tracks at the railway station in a depression between two railway embankments (2001, Z. Kaplan PRA). Native to North America (British Columbia). — *Cotoneaster horizontalis*. Kunětická hora hill near Pardubice. The species grows there as a cultivation relic planted in the 1930s by a natural-historical society from the town of Pardubice; it has spread since then and persists (F. Procházka, pers. com.). Praha-Radotín: as a cultivation relic on a rock in the area of cement works; Brno-Řečkovice: Žitná street, escaped from cultivation,

between a stone wall base and asphalt pavement (2001, V. Řehořek). — *Crataegus flabellata*. Javorník (distr. Jeseník): the Jánský vrch hill ca. 1 km SW of the town, shrubs along a path between fields (1993, J. Vicherek); Praha 5-Černý vrch: several young shrubs in open spaces and among shrub plantations (2001, J. Sádlo). — *Crataegus persimilis*. Lednice, distr. Břeclav: escaping from cultivation in the chateau park (M. Pejchal, pers. com.). — *Crocus flavus*. Morávka, distr. Frýdek-Místek: understory of a beech woodland at the bank of the Morávka river (1999, P. Pyšek & B. Mandák PRA, det. J. Holub). Reported by Dostál (1989) as intentionally planted in the wild near Praha-Hlubočepy. — *Deutzia scabra*. Liberec: escaped by seed in open spaces and in the streets, among cobble stones (2001, J. Sádlo). — *Doronicum orientale*. Naturalized as a cultivation relic in the Terčino údolí valley near Nové Hradky, S Bohemia; it has been known from there since 1884 (Čelakovský 1885), and still grows there (M. Štech, pers. com.). — *Doronicum columnae*. Occasionally planted in the wild and surviving as a cultivation relic, e.g. district of Brno: deciduous forest on the Mniší hora hill E of the Kníničky village (1995, M. Hladíková); allotments in Slabčice near the Vltava river (M. Štech, pers. com.). — *Forsythia suspensa*. Praha-Libeň, slope with scrub above the Rokytka brook; Vínův: in the village. In both localities, the species spreads by rooting of shoots (2001, J. Sádlo). — *Fraxinus ornus*. Praha-Troja: south-oriented slopes above the Trojská street and elsewhere around (approximately since the 1950s); Bohemian Karst: Koněprusy, Beroun (J. Sádlo). In the past, several specimens were reported by Čelakovský (1872) growing on the “Niklasberg” (Mikulášský kopec) hill near Český Krumlov but it was not possible to determine whether or not these were planted and persisted as cultivation relics. Although the species should be considered as naturalized, neither the Flora of the Czech Republic nor Kubát et al. (2002) mention escaping from cultivation. — *Hippophaë rhamnoides*. First reported as garden escape from a hill near Podhradice NE of Břilina (Polívka 1901). Recently, a plant established from a discarded root has been persisting since ca. 1995 in Praha-Jinonice, in a soil deposit near underground station (2000, J. Sádlo). — *Hosta plantaginea*. Escapes occasionally from cultivation in gardens and parks. — *Iris pallida*. Escapes occasionally from cultivation in gardens and parks (J. Holub, pers. com.). — *Juglans nigra*. S Moravia: rather frequent in forest plantations at the confluence of the Morava and Dyje rivers, intensively escaping from cultivation in places, e.g. Ranšpurk nature reserve (Vicherek et al. 2000); Lednice, distr. Břeclav: rather easily escaping from cultivation in the chateau park and its surroundings (M. Pejchal, pers. com.). — *Lycium chinense*. Nebanice, distr. Cheb: it grows as a clonally spreading cultivation relic around a poorly maintained church, and penetrates into surrounding vegetation (J. Sádlo). — *Miscanthus sinensis*. Garden escape at fishpond barrier in Malenice near Volyně, and in ruderal grassland in Praha-Satalice (J. Sádlo). — *Paeonia officinalis*. Srbsko, distr. Beroun: a plant established from rubbish deposited in the limestone quarry Na Chlumu has been persisting for ca. 20 years, and spreads slightly (J. Sádlo). — *Physalis pubescens*. Zlatá Koruna, distr. Český Krumlov: in the village (2001, M. Lepší, in prep.). — *Platanus xhispanica*. Praha: Vinohrady hospital; Praha, Botičská street: an adult tree originated from seed was observed in 1990 and later cut down (J. Sádlo). — *Populus balsamifera*. First reported as a cultivation escape by Čelakovský (1880) along the road from Česká Lípa to Nové Zámky and by Velenovský from the region of Blatná. Recently, three specimens were found in Stará Paka (distr. Jičín) growing at the periphery of the railway station (1964, V. Jehlík PRA). Neither the Flora of the Czech Republic nor Kubát et al. (2002) mention escape from cultivation. — *Potentilla fruticosa*. Kunětická hora hill near Pardubice. The species grows as a cultivation relic planted in the 1930s by a natural-historical society from the town of Pardubice; it has spread since then and persists (F. Procházka, pers. com.). — *Prunus laurocerasus*. Praha-Spořilov, Průhonice: numerous self-seeded young plants around planted adults (2001, J. Sádlo). — *Prunus virginiana*. Lednice, distr. Břeclav: escaped from cultivation in the chateau park (M. Pejchal, pers. com.). — *Ricinus communis*. Kozomín near Kralupy nad Vltavou: two flowering plants on a rubbish tip at the village periphery (1996, P. Pyšek & B. Mandák). — *Salix xsepulcralis*. Praha-Radotín: a single shrub on a Vltava river alluvium (2001, J. Sádlo). — *Scopolia carniolica*. It was reported as long persistent (1866–1880) from school garden in Valteřice near Česká Lípa (Čelakovský 1881). At present, it grows in Praha-Divoká Šárka, probably as a cultivation relic (J. Hadinec, pers. comm.). Recently, a small established population of ca. 10 plants was found in Žampach, distr. Praha-západ (2001, J. Sádlo PRA). — *Sedum annuum*. Old records about escapes from cultivation (Čelakovský 1867–1881, 1900–1904) are doubtful (Grulich in Hejný & Slavík 1992). Recently, it was found as a garden escape in the region of Křivoklát (J. Kolbek, pers. com.). — *Tilia tomentosa*. Praha-Karlovo náměstí square: several young trees from self-seeding, growing in a ruderal habitat by a house (2001, J. Sádlo). — *Tulipa xgesnerana*. Escapes from cultivation in gardens, waste places, rubbish tips and villages.

Remarks on other species:

Acorus calamus. Immigration time of this species requires further study as it probably arrived earlier than usually considered (R. Hendrych, in litt.). — *Allium tuberosum*. The herbarium specimen collected by Čelakovský in Praha-Chuchle 1866, and listed since then in several floras (Dostál 1948–1950, 1989) under the name *Allium odorum* L., belongs to this species (F. Krahulec, in prep.). — *Allim atroviolaceum*. Erroneously reported from South Moravia, near the village of Sokolnice (Dostál 1948–1950, 1989). These plants belong to another taxon; however, herbarium specimen of *A. atroviolaceum* from Pouzdřany, South Moravia, has been discovered recently (F. Krahulec, in prep.). — *Amelanchier lamarckii*. First reported under the name *A. canadensis* as growing wild from the Terezino údolí valley in the vicinity of Nové Hradky (Velenovský 1877 in Čelakovský 1881), also mentioned as garden escape by Polívka (1900–1904). However, *A. canadensis* is cultivated very rarely in Europe, only in few botanical gardens. The old records relate to *A. lamarckii*, grown for its fruits and medicinal use, with which it is often confused. The latter species easily escapes from cultivation in western Europe (M. Pejchal, pers. com.). — *Bromus pumpellianus* × *inermis*. This taxon was first reported as *Bromopsis pumpelliana* subsp. *flexuosa*, syn. *Bromus pumpellianus* (Krahulec & Jiříš 1997) but further studies revealed that it is most probably a hybrid between *B. pumpellianus* and *B. inermis* (F. Krahulec, pers. com.). — *Bunias orientalis*. The species is considered as a neophyte but the time of its immigration into Central Europe requires further study as it is listed by Lang (1994) as an archaeophyte having arrived in the Middle Ages. — *Carex muskingumensis*. Not listed in Kubát et al. (2002). In 1947, two tussocks were collected on a rubbish tip in Brno-Pisárky; later, a more abundant source population was found in a semi-natural reed stand by a small pond (Grüll 1952). — ×*Conygeron huelsenii*. This hybrid was found in 1887 near Ročov, distr. Louny, in a clearing by the road from Netluky to Třeskonice, in the population of both parents (Čelakovský 1888b). — *Cyperus eragrostis*. A single plant appeared on an emergent bottom of the water reservoir in Jablonec nad Nisou (Petřík 2002). — *Eichhornia crassipes*. Osek (distr. Nymburk): ca. 10 plants growing in a water reservoir at the SE margin of the village (2000, J. Hadinec & P. Havlíček). Also reported by Rydlo (1992) as planted in a pool near the village of Račice at Křivoklát region, C Bohemia, in 1991, and later by the same author (Rydlo 2001) from the section of Labe river between Mělník and Hrobc, albeit without further details. — *Oenothera stricta*. Reported from the Vltava river bank (Knaf 1825 PR – Jehlík in Slavík 1997). Recently, a single plant was found growing as a weed on a garden bed in Husova 342, Vroutek, distr. Podbořany (2001, A. Pyšek & P. Pyšek PRA). — *Polygonum aviculare* agg. Pollen of this aggregate species was found in Mesolith (P. Pokorný, pers. com.), which qualifies it as a native species. We believe that this finding concerns *P. arenastrum* Bor. which is considered an apophyte (in the sense of Holub & Jirásek 1967) whereas *P. aviculare* s. str. is considered an archaeophyte, on the basis of its ecology and distribution outside the territory of the Czech Republic. — *Setaria faberi*. Plants treated as *S. macrocarpa* Lucznik by some authors (e.g. Jehlík 1998) are included here within this taxon, according to Kubát et al. (2002). — *Veronica filiformis*. The Caucasian species *V. filiformis* was described by Smith in 1791. Unfortunately, Besser used the same name for *V. persica* (syn. *V. tournefortii*, *V. buxbaumii*) in 1809, as did De Candolle in 1815. For this reason, Presl & Presl (1819) listed *V. filiformis* in their Flora Čechica but the record relates to *V. persica*. The same mistake was made by Pohl (1809) and Opiz (1823). Čelakovský (1867–1881) obviously distinguished between the two species and avoided the confusion as can be inferred from the fact that he does not list *V. filiformis* for the Czech flora (Bohumil Slavík, pers. com.). The “true” *V. filiformis* was introduced as late as in 1938. — *Zelkova serrata*. Erroneously reported as *Z. carpinifolia* in the Flora of the Czech Republic (Hrouda in Slavík 1988). Hundred and twenty specimens were planted on arable land in the arboretum near “Tři grácie” near Lednice in S Moravia. Thousands of saplings have escaped from browsing danger and grow in the surrounding vegetation; where they were first observed in 1973 (M. Pejchal, pers. com.).

Appendix 2. – Structure of the database. See text for more details on characteristics concerning species invasiveness in the Czech Republic. The four main spheres of information are displayed in bold, database fields are printed in upper case.

I. Species identity and taxonomic position: GENUS, SPECIES, SUBSPECIES, SYNONYMS, FAMILY, ORDER, SOURCE OF INFORMATION: floras, important detailed papers; LOCALITY: only in rare species.

II. Invasiveness (features related to the species ecology and behaviour in the Czech Republic) INVASIVE STATUS (sensu Richardson et al. 2000): casual, naturalized, invasive, post-invasive, cultivation relict; RESIDENCE TIME (period of immigration): archaeophyte, neophyte. Recently found hybrid between two archaeophytes is also considered as an archaeophyte; despite of it being reported only recently, the species could have hybridized since the time of arrival of the later immigrant. Hybrid between archaeophyte and neophyte must be, following the same logic, considered a neophyte. YEAR OF INTRODUCTION: in deliberately introduced taxa (applicable namely for woody plants); YEAR OF THE FIRST RECORD in the wild; TYPE OF INVADED HABITAT: natural, seminatural, human-made (Chytrý et al. 2001); TYPE OF INVADED LANDSCAPE: traditional agricultural landscape, industrial urban landscape (Hobsbawm 1991). LIST OF INVADED HABITATS: based on Council Directive 92/43/EEC on the conservation of natural habitats and of wild fauna and flora (1992); SOIL description; PHYTOSOCIOLOGICAL UNITS: list of alliances of the Zürich-Montpellier classification system in which the species is found; NUMBER OF PHYTOGEOGRAPHICAL DISTRICTS from which the species is reported, divided into the three basic types: Thermophyticum, Mesophyticum, Oreophyticum (Skalický 1988); ALTITUDINAL RANGE: minimum, maximum; ABUNDANCE in the wild at the territory of the country: single locality, rare, scattered, locally abundant, common, extinct (if no records have been known for a long period); quantitative estimate of the number of localities using the scale of Clement & Foster (1994): 1–4, 5–14, 15–49, 50–499, at least 500 localities; REGION where the occurrence is concentrated given that species distribution has a regional pattern. TYPE OF INTRODUCTION into the country: deliberate, accidental, or both types. Spontaneously originated hybrids are considered as “accidental”, hybrids escaped from cultivation are considered “deliberate”. PLANTING PURPOSE: ornamental, forestry, agriculture (other than food), food, oil, fodder, medicinal, botanical, bee, textile, dye, landscaping, etc. VECTOR OF ACCIDENTAL INTRODUCTION: grain, seed, fodder, wool, cotton, vine, flax, agricultural products, ore, soya beans, bird-seed, garden material, etc; HISTORY OF INTRODUCTION: description of the introduction into the country and species invasion history. INVASIVENESS ELSEWHERE in the world.

III. Native distribution. AREA OF ORIGIN: classified into geographical regions according to the system used by Brummitt et al. (2001); MAPS available, showing species distribution; LATITUDINAL AND LONGITUDINAL RANGE of primary distribution; HABITATS occupied in primary distribution area; MINIMUM AND MAXIMUM HEIGHT reached in primary distribution area.

IV. Biological and ecological characteristics. LIFE FORM: annual, biennial, monocarpic perennial, polycarpic perennial, shrub, semishrub, tree, climber; RAUNKIAER SCHEME: therophyte, hemicryptophyte, geophyte, chamaephyte, nanophanerophyte, phanerophyte; LIFE STRATEGY: C, S, R, CS, CSR, SR (Grime 1979); CLONALITY TYPE (according to Klimešová & Klimeš 1998). MINIMUM AND MAXIMUM HEIGHT reached in the Czech Republic; SEXUAL REPRODUCTION IN CR: yes, no, rarely; BREEDING SYSTEM: allogamy (protandry, protogyny), autogamy (facultative, obligate), cleistogamy, apogamy, geitonogamy; SEX TYPE: dioecy, monoecy, andromonoecy, gynomonoecy, gynodioecy, polygamy; PLOIDY LEVEL; CHROMOSOME NUMBER; DNA CONTENT: taken from Bennett & Leitch (2001); HYBRID TYPE: none, neophyte × neophyte, neophyte × native, neophyte × archaeophyte, archaeophyte × archaeophyte, archaeophyte × native, originated in cultivation; FLOWERING TIME: start, end; FLOWER COLOUR; FRUIT TYPE: achene, nut, berry, drupe, capsule, follicle, pod, silique, silicula, loment, pome, nutlet, schizokarpium; FRUIT SIZE: minimum, maximum; SEED SIZE: minimum, maximum; PROPAGULE: seed, fruit, fruit fragment; stem, whole plant, root, rhizome, rosette; PROPAGULE WEIGHT; FECUNDITY: number of propagules per plant; SEED BANK TYPE: I–IV (Thompson et al. 1997); PROPAGULE CHARACTERISTIC: description and special features; DORMANCY: non-dormant, morphological, physiological, morphophysiological (Baskin & Baskin 1999); POLLINATION MODE: wind, insect; DISPERSAL MODE: autochory, endozoochory, epizoochory, myrmecochory (dispersal by ants), anemochory (wind), hydrochory (water); ELLENBERG INDICATOR VALUES: light, temperature, moisture, soil reaction, nitrogen (Ellenberg et al. 1991).